

## CURRICULUM VITAE

### I. BIOGRAPHICAL DATA

NAME: Jens R. Coorssen  
 CITIZENSHIP: Canadian & Australian  
 ADDRESS: 110 Dalhousie Avenue  
 St. Catharines, ON L2N4X2, Canada  
 TELEPHONE: 905 688 5550 x6870  
 RANK: Professor (tenured)  
 DEPARTMENTS: Health Sciences and Biological Sciences  
 FACULTIES: Applied Health Sciences and Mathematics & Science  
 INSTITUTION: Brock University  
 (Adjunct Professor, School of Medicine, Western Sydney University)  
 (Honorary Professor, School of Medicine, University of Sydney)  
 (Associate Member, ICORD/Blusson Spinal Cord Centre/UBC)

### II. ACADEMIC RECORD

FINAL DEGREE: Ph.D.  
 DATE COMPLETED: 1993  
 SPECIALTY: Cell Physiology (Medical Sciences)  
 INSTITUTION / CITY / COUNTRY: McMaster University/Hamilton/Ontario/Canada

#### i. UNDERGRADUATE

1986: Brock University, Honours B.Sc., Biological Sciences,  
St Catharines, Ontario, Canada

#### ii. GRADUATE / POST-DOCTORAL / SHORT-TERM APPOINTMENTS

1988: M.Sc., Biological Sciences (Membrane Biophysics; Dr. R.P. Rand),  
 Brock University, St. Catharines, Ontario, Canada  
 1993: Ph.D., Medical Sciences Program (Cell Physiology),  
 McMaster University, Hamilton, Ontario, Canada  
 02/93 – 12/95: PDF, Dept. Molecular Cell Research (Dr. W. Almers), Max-Planck-  
 Institut für Medizinische Forschung, Heidelberg, Germany  
 01/96 – 06/96: Research Associate (BAT II), Dept. Molecular Cell Research, Max-  
 Planck-Institut für Medizinische Forschung, Heidelberg, Germany  
 06/95 – 07/95 &  
 05/96 – 06/96: Visiting Scientist, Neurosciences Lab, Dept. Biological Sciences,  
 Brock University, St. Catharines, Ontario, Canada  
 06/96 – 08/99: Visiting Fellow, Lab. Of Cellular & Molecular Biophysics  
 (Dr. J. Zimmerberg), NICHD, NIH, Bethesda, Maryland, USA

### III. AWARDS AND DISTINCTIONS (\* declined; † terminated with move to Australia)

2019-present: **Senior Research Fellow**, Faculty of Applied Health Sciences, Brock U.  
 2016: **Paul Harris Fellow of Rotary International**; recognition for commitment to  
 Rotary values in supporting community-driven research into Multiple Sclerosis.  
 2016-present: **Adjunct Professor**; School of Medicine, Western Sydney University.  
 2014-present: **Honorary Professor**; School of Medicine, University of Sydney.  
 2012: **Alumnus of Distinction Award**, Brock University.  
 2010-present: **Adjunct Professor**; Faculty of Medicine, University of Calgary.  
 2009-2016: **Head**, Molecular Medicine Research Group, Western Sydney University.

- 2008-2016: **Foundation Professor**, School of Medicine, Western Sydney University.
- 2008: **Celebrating Excellence Award**, University of Calgary; recognition for "inspiration, innovation, and outstanding contributions to teaching & research."
- 2006-2011<sup>†</sup> **Senior Scholar**, Alberta Heritage Foundation for Medical Research.
- 2006-2011<sup>†</sup> **New Investigator**, Canadian Institutes of Health Research.
- 2006\* **Canada Research Chair (Tier II)**, Brock University, ON, Canada.
- 2001-2002 **Young Innovator Award**, Vice-President (Research), University of Calgary.
- 2000-2005 **Research Scholar**, Heart & Stroke Foundation of Canada.
- 2000-2006 **Research Scholar**, Alberta Heritage Foundation for Medical Research.
- 1999-2001 **Visiting Professor** (Exchange Scientist), Laboratory of Cellular and Molecular Biophysics, NICHD, National Institutes of Health, Bethesda, Maryland, U.S.A.
- 1996-1999 **Fogarty International Fellowship**, Laboratory of Cellular and Molecular Biophysics, NICHD, National Institutes of Health, Bethesda, Maryland, U.S.A.
- 1994-1996 **Post-doctoral Fellowship**, Natural Sciences & Engineering Research Council of Canada, Max-Planck-Institut für Medizinische Forschung, Heidelberg, Germany.
- 1994-1996 **Research Fellowship** from the Max Planck Society, Max-Planck-Institut für Medizinische Forschung, Heidelberg, Germany.
- 1993-1994 **Visiting Fellowship** Award from the Max Planck Society, Max-Planck-Institut für Medizinische Forschung, Heidelberg, Germany.
- 1989-1992 **Studentship**, Medical Research Council of Canada, McMaster University, Canada.
- 1988-1992 **Centennial Scholarship**, McMaster University, Canada.
- 1989-1990\* **Research Traineeship**, Heart & Stroke Foundation Canada, McMaster U., Canada.
- 1989-1990\* **Ontario Graduate Scholarship**, McMaster University, Canada.
- 1988-1989 **Governor General's Gold Medal**, Graduate Studies, Brock University, Canada.
- 1988-1989 **Ontario Graduate Scholarship**, McMaster University, Canada.
- 1986-1988 **Postgraduate Scholarship**, Natural Sciences and Engineering Research Council of Canada, Brock University, Canada.
- 1987-1988\* **Ontario Graduate Scholarship**, Brock University, Canada.
- 1985-1986 **Scholarship** for Honours B.Sc. Research, Dept. Biological Sciences, Brock University, Canada.

#### IV. ACADEMIC APPOINTMENTS

**Professor** (tenured)  
Faculty of Applied Health Sciences  
Faculty of Mathematics and Science  
Brock University

**Professor** (tenured)  
Foundation Chair of Molecular Physiology  
School of Medicine  
Western Sydney University

#### July 2016 - present

- Professor, Departments of Health Sciences and Biological Sciences
- Dean, Graduate Studies (07/16-09/17)
- Adjunct Professor at WSU (07/16-present)

#### February 2008 – June 2016

- Head, WSU Molecular Medicine Research Group, 08/09 (first Leader of this Res. Group).
- Conjoint Professor in the School of Biomedical & Health Sciences, 09/08-06/16.
- Associate Member, Nanoscale Organisation & Dynamics Group (2008-2016).

**Adjunct Professor**

Dept. Physiology and Pharmacology  
Dept. Biochemistry and Molecular Biology  
Faculty of Medicine  
University of Calgary

**Jan 2010 – present**

**Honorary Professor**

Medical Sciences  
Sydney Medical School  
University of Sydney

**June 2014 – present**

**Associate Member**

ICORD (International Collaboration On Repair Discoveries)  
Blusson Spinal Cord Centre / UBC  
Vancouver, BC, CANADA

**April 2012 – present**

**Foreign Member**

European Cooperation in Science and Technology (COST-FAP1002)  
European Union

**August 2012 – 2015**

**Associate Professor**

Dept. Physiology and Biophysics  
Hotchkiss Brain Institute  
Faculty of Medicine  
University of Calgary

**July 2004 – Dec 2009** (LOA 3/08-12/09)

- cross appointment in Cell Biology & Anatomy, 03/05.
- co-Leader of the Neuroconnections Program within the Hotchkiss Brain Inst., 10/06–12/07

**Assistant Professor**

Dept. Physiology and Biophysics  
CMNRG  
Faculty of Medicine  
University of Calgary (U of C)

**Dec 2000 - June 2004**

- cross appointment in Biochemistry & Molecular Biology, 08/01.

**Visiting Professor (Exchange Scientist)**

Laboratory of Cellular & Molecular Biophysics  
National Institutes of Health  
Bethesda, MD USA

**Oct. 1999 - May 2001**

- A collaboration with the LCMB (NICHD, NIH)
- responsible for operations in the Cell Physiology section

**V. EDUCATIONAL ACTIVITIES**

**PRIMARY SCHOOL ENGAGEMENT**

Glenbrook Primary;                      Molecules-to-Cells                      Talk (2015)  
Years 5 & 6

**HIGH SCHOOL ENGAGEMENT** — largely through CSIRO Scientists in Schools program, with which I was affiliated from 2011-2016

Wollondilly Anglican                      Independent Research Projects                      Small Group class discussions & lab  
College; Yr 11                      sessions (2014, 2015)  
Chevalier College; Yr                      1)Molecules-to-Cells; 2)  
11                      Imaging                      Talks (2009)

**Invited workshop leader for the Canadian Medical Hall Of Fame Discovery Days** in Health Sciences for gifted High School students from across Canada. At U. Calgary: Introduction to Functional Proteomics (2003-2006)

**Invited workshop leader for the Shad Valley International Program** for gifted high school students. At U. Calgary (07/2003).

**Online Teaching and Community Engagement in Microsoft Teams.** Brock University webinar introducing use of Teams for online teaching. In preparation for teaching in the coming semester (06/2020).

### UNDERGRADUATE COURSES

|                                       |   |   |
|---------------------------------------|---|---|
| PBL                                   | Problem Based Learning & trainer for new tutors.  | Facilitator for Nursing & Medical students (McMaster U, 1988-1992)  |
| MDCN 340                              | Research Methods & Evidence Based Medicine  | Small Group Leader (2001 & 2002)  |
| MDSC 203 B.H.Sc.                      | Inquiry-Based learning component  | Group Facilitator (2003 & 2004)   |
| BCEM 507.23 B.Sc.                     | Independent Research Projects   | Coordinator (2002, 2004-2007)   |
| MDSC 397 B.H.Sc.                      | Independent Study Projects  | Coordinator (2005-2007)   |
| MDSC 507.02 B.H.Sc.                   | Independent Research Project  | Coordinator (2007)  |
| UWS 300552; Mol Biol of Immune System | Intro to Proteomics   | Lecture (2008, 2009)  |
| UWS 300541; Biomolecular Frontiers    | 'Omics & how they are shaping/directing biomedical research   | Lecture (2009-2012)   |
| UWS 300555; Proteins & Genes          | Intro to Membrane Trafficking   | Lecture (2009-2012)   |
| WSU Medicine Year 1                   | 1) Intro to Systems Biology; 2) Molecules of Life; 3) Cell Membranes; 4) Enzymes & Enzyme Kinetics; 5) Cytoskeleton; 6) Exocytosis & Endocytosis; 7) Metabolic response to starvation | Lectures (2009-2016)<br>N.B. changed to a self-directed learning format in 2016 that involved one lecture, recordings, alternate information sources, and a 2h Q&A session. |
| WSU Medicine Year 2                   | 1) Exocytosis & Endocytosis; 2) Axon Guidance & Synapse Formation; 3) Intro to 'Omics   | Lectures (2009-2015)  |
| HLSC 3P30 (Brock U.)                  | Directed Readings   | Mentor; student project required extensive reading (& may include interview of an expert), public presentation & detailed final report + reflection (2018-19; Summer 2020). |
| HLSC 3Q91                             | Special Topics in Medical Sciences  | Mentor; student literature survey & study design + reflection (Spring 2020)   |
| HLSC 4P98 (Brock U.)                  | Biomarkers & Disease Progression  | Lectures/Seminars + flipped classroom (2018 F&W, 2019 F)  |
| HLSC 4P30 (Brock U.)                  | Independent study   | Mentor; student project requiring experimental work + extensive reading, public presentation & detailed final report (2018-19).   |
| BMED 4F90/4F91(Brock U.)              | Honours BSc thesis  | Mentor; literature review and thesis research (2019-2020 x 2)   |

## GRADUATE COURSES

|                                    |   |   |
|------------------------------------|---|---|
| MDSC 755.03                        | Introduction to Functional Proteomics (practical) | Course coordinator (2002 - 2006)<br>- entirely responsible for course design & instruction; hands-on, independent study format. |
| MDSC 755.63                        | Critical Perspectives in Proteomics               | Course co-coordinator (2003 & 2004; with Dr. D. Schriemer)  |
| MDSC 755.06                        | The Exocytotic Pathway                            | Course co-coordinator (2002)<br>(with Dr. J. Braun)   |
| M.B.T. Program                     | Introduction to Functional Proteomics             | Lecture (2003-2005)   |
| PSYC4630A                          | Joint U of C – U. Lethbridge program              | Lecture (9/2003-2006)   |
| MDSC 619.01                        | Neuroscience I                                    | Lecture (9/2003 & 2004); Course coordinator (2005-2007)   |
| MDSC 755.08                        | Biochemistry & Molecular Biology                  | Lecture/Seminar (2006, 2007)  |
| Canadian Medical Education         | Continuing Medical Education for Physicians       | Seminar & course paper (2006)   |
| BIOL 607 - B01                     | Special Problems in Biology                       | Course designer/co-ordinator (2014; with Prof E. Prenner, U. Calgary)   |
| WSU MRes<br>School of Grad Studies | Intro to Mol Physiol & the MMRG                   | Invited talk to 1 <sup>st</sup> MRes cohort (2015)  |
| WSU MRes<br>School of Grad Studies | Stages of Research                                | Key intro graduate lecture (2015)   |
| Brock U.<br>FAHS AHSC7P61          | Practical Top-down Proteomics                     | Independent research study (N. Kurgan, 11/17-08/18; K. Carbonara, 09/19-01/20; B. Baranowski, 09/19-06/20)                      |
| Brock U.<br>AHSC7P96               | Top-Down Proteomics                               | Labs/lecture/seminars (W 2018/19; 2019/20)  |

## SUPERVISOR

### i. UNDERGRADUATE

| <u>Name</u>       | <u>Position</u>     | <u>Period</u>  |
|-------------------|---------------------|--|
| Ms. A. Piazza     | Hon. B.Sc. study    | 09/20 – 04/21  |
| Ms. D. El Jaroudi | Hon. B.Sc. study    | 09/19 – 04/20  |
| Ms. K. Carbonara  | Summer Research     | 05/19 – 09/19 (NSERC USRA)                                     |
|                   | Independent study   | 01/19 – 04/19 (HLSC 4P30 project)                              |
| Ms. D. Sobers     | Summer Research     | 05/19 – 08/19 (FMS International Student Research Scholarship) |
|                   | Independent study   | 01/19 – 04/19 (HLSC 3P30 project)                              |
|                   | Summer Research     | 05/18 – 09/18  |
| Ms. L. Stone      | Independent study   | 09/18 – 12/18 (HLSC 3P30 project)                              |
| Mr. M. Mahrouse   | Hon. B.Sc. study    | 01/18 – 08/18; supervisory committee                           |
| Mr. D. Emmerig    | Hon. M.B.B.S. study | 03/15 – 11/16 (Embedded Honours)                               |
| Mr. T. Dias       | Hon. B.Sc. study    | 03/15 – 11/15; co-supervisor                                   |

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|--------------------------|---|--|
| Ms. N. Noaman            | Biomolecular Projects                   | 08/13 – 11/13 (Yr 3 research project)  |
|                          |   | 02/13 – 05/13 (Yr 3 research project)  |
|                          | Independent Research<br>& Summer Fellow | 07/14 – 03/15 (co-supervised with Prof<br>N. Merrett; UWS Award)                                   |
| Mr. P. Paasila           | Summer Research                         | 12/14 – 02/15 (Year 1 Med Student)   |
| Mr. H. Rodwell           | Biomolecular Project                    | 08/14 – 11/14 (Yr 3 research project)  |
| Mr. B. Smith             | Biomolecular Project                    | 08/13 – 11/13 (Yr 3 research project)  |
| Ms. K. Cox               | Biomolecular Project                    | 08/12 – 11/12 (Yr 3 research project)  |
|                          | Summer Fellow                           | 12/12 – 02/13 (UWS Award)  |
| Ms. B. Schaffran         | Exchange Fellow                         | 09/12 – 02/13 (German Exchange)  |
| Ms. T. Tsourvakas        | Biomolecular Project                    | 08/12 – 11/12 (Yr 3 research project)  |
| Mr. J. Po                | Hon. B.Sc. study                        | 07/11 – 06/12; co-supervisor   |
| Ms. E. Kerketta          | Hon. B.Sc. study                        | 08/11 – 03/12  |
| Mr. F. Morales           | Biomolecular Project                    | 12/11 – 03/12 (Yr 3 research project)  |
| Mr. N. Nikesitch         | Biomolecular Project                    | 08/11 – 11/11 (Yr 3 research project)  |
| Mr. P. Abbineni          | Hon. B.Sc. study                        | 02/11 – 11/11 (Best thesis in Sciences)  |
| Ms. S. Papalia           | Hon. B.Sc. study                        | 02/11 – 11/11  |
|                          | Summer Fellow                           | 11/10 – 02/11 (UWS/CHS Award)  |
|                          | Biomolecular Project                    | 02/10 – 11/10 (Yr 3 research project)  |
| Ms. S. Lim               | Hon. B.Sc. study                        | 02/11 – 11/11; co-supervisor   |
| Mr. P. Wisinski-Bokiniec | Hon. B.Sc. study                        | 02/11 – 11/11; co-supervisor   |
| Mr. L. Marshall          | Hon. B.Sc. study                        | 07/10 – 06/11; co-supervisor   |
| Mr. M. Featon            | Hon. B.Sc. study                        | 07/10 – 06/11; co-supervisor   |
| Mr. H. Bradley           | Summer Fellow                           | 11/10 – 02/11 (from Medicine, U. Notre<br>Dame; UWS/CHS Award)                                     |
| Mr. K. Smith             | Hon. B.Sc. study                        | 02/10 – 11/10; co-supervisor   |
| Mr. S. Stimpson          | Hon. B.Sc. study                        | 02/10 – 11/10; co-supervisor   |
| Mr. T. Deshmukh          | Summer Fellow                           | 11/09 – 02/10 (UWS Medicine; Summer<br>Student Research Scholarship)                               |
| Ms. A. Grunwald          | Co-Op Fellow                            | 01/09 – 05/09 (McMaster U.)  |
| Mr. J. Fahr              | Co-Op Fellow                            | 01/07 - 10/07 (German exchange)  |
|                          |   | 05/08 - 08/08  |
| Ms. K. Dean              | Summer Fellow                           | 05/08 - 08/08 (UC Health &<br>Wellness Research Award)   |
| Ms. A. Hsu               | Summer Fellow                           | 05/08 - 08/08  |
| Mr. G. Marlowe           | Hon. B.Sc. study                        | 09/06 - 05/07 (UC Biochemistry)  |
| Ms. S. Tran              | Hon. B.Sc. study                        | 09/06 - 05/07 (B.H.Sc. program)  |
| Ms. M. Lee               | Hon. B.Sc. study                        | 09/06 - 05/07 (B.H.Sc. program; UC<br>Health & Wellness Research Award)                            |
|                          | Summer Fellow                           | 05/05 - 08/05 (CHR/CLS Summer<br>Research Studentship)   |
|                          |   | 05/04 - 08/04 (AHFMR)  |
| Ms. A. Krioutchkova      | Summer Fellow                           | 05/07 - 08/07 (AHFMR)  |
| Ms. L. Harris            | Co-Op Fellow                            | 05/06 - 12/06 (McMaster U.)  |
| Mr. R. Krishnan          | Summer Fellow<br>(from Cornell)         | 05/07 - 08/07 (AHFMR)<br>& 05/06 - 08/06 (AHFMR; NSERC<br>awarded but declined)                    |
| Ms. M. Chow              | Hon. B.Sc. study                        | 09/05 - 05/06 (UC Biochemistry)  |
|                          | Summer Fellow                           | & 05/06 - 08/06  |
| Mr. J. Lang              | Summer Fellow<br>(from UBC)             | 05/05 - 08/05 (NSERC)<br>& 05/04 - 08/04 (NSERC; best<br>presentation at summer student symposium) |
|                          |   | & 07/03 - 08/03  |
|                          |   | & 06/02 - 08/02 (HYRS/AHFMR)   |
| Mr. J. Bau               | Summer Fellow                           | 05/06 - 08/06 (NSERC)  |
|                          | B.H.Sc. Res. Project                    | & 01/05 - 05/05 (B.H.Sc. program)  |
|                          | Summer Fellow                           | & 05/05 - 08/05 (NSERC)  |

|                       |                                   |  |
|-----------------------|-----------------------------------|--|
| Mr. D. Brandman       | Summer Fellow<br>(from UBC)       | 05/06 - 08/06 (NSERC)<br>& 05/05 - 08/05 (STEP support)  |
| Mr. A. Tang           | Summer Fellow                     | 05/07 - 08/07 (NSERC)<br>& 06/06 - 08/06 (NSERC)<br>& 07/05 - 08/05 (HYRS/AHFMR)                     |
| Mr. P. Tran           | Summer Volunteer                  | 05/05 - 08/05 (Pre-Grad student)   |
| Ms. A. Chan           | B.Sc. Res. Project                | 01/05 - 05/05 (UC Biochemistry)  |
| Mr. J. Lee            | B.Sc. Res. Project                | 09/04 - 12/04 (UC Biochemistry)<br>01/06 - 06/08 (part-time research)                                |
| Ms. B. Sargent        | Summer Fellow                     | 05/04 - 08/04 (B.H.Sc. program)<br>& 09/04 - 12/04 (part-time research)                              |
| Ms. J. Thompson       | Summer Fellow                     | 07/04 - 08/04 (HYRS/AHFMR)   |
| Mr. J. Leung          | Summer Fellow                     | 05/04 - 08/04 (AHFMR; Co-supervised<br>with Dr. Syed; best presentation at summer student symposium) |
| Mr. N. Wu             | Summer Fellow                     | 06/04 - 08/04<br>& 09/03 - 05/04 (Volunteer)<br>& 06/03 - 08/03 (HYRS/AHFMR)                         |
| Mr. K. Marshall       | Co-Op Fellow                      | 09/03 - 04/04 (U. Lethbridge)  |
| Mr. M. Churchward     | Co-Op Fellow                      | 05/03 - 12/03 (McMaster; STEP)   |
| Ms. K. Hsu            | Summer Fellow                     | 07/03 - 08/03  |
| Ms. M. Pratt          | Hon. B.Sc. study<br>Summer Fellow | 09/02 - 05/03 (UC Zoology)<br>05/02 - 08/02  |
| Mr. M. Watson         | IB Student Fellow                 | 09/02 - 05/03 (IB research program)  |
| Mr. R. Butt           | Co-Op Fellow (U of L)             | 05/02 - 12/02 (STEP support)   |
| Ms. E. Barr           | Summer Fellow                     | 05/02 - 08/02 (MacKenzie award)  |
| Ms. J. Hibbert, B.Sc. | Summer Fellow                     | 05/02 - 08/02 (SCPP support)   |
| Mr. C. McLelland      | Summer Fellow                     | 05/02 - 08/02 (Penn West award)  |
| Ms. L. Watkins        | Summer Fellow                     | 05/02 - 08/02 (McMaster exchange)  |
| Mr. R. Sachdeva       | Summer Fellow                     | 05/02 - 08/02  |
| Mr. R. Sharma         | Summer Fellow                     | 05/02 - 08/02  |
| Mr. I. Soo            | Summer Fellow                     | 05/02 - 08/02  |
| Ms. T. Arness         | Summer Fellow                     | 05/00 - 08/00  |
| Ms. M. Gestole        | Summer Fellow                     | 06/00 - 09/00  |
| Mr. J. Kirshtein      | Summer Fellow                     | 05/99 - 08/99  |
| Mr. A. Choi, B.S.     | Summer Fellow                     | 05/98 - 08/98<br>& 05/97 - 08/97   |
| Mr. M. Minardi, B.Sc. | Summer Fellow                     | 06/97 - 08/97  |
| Ms. M. Kasowski       | Summer Fellow                     | 06/97 - 08/97  |

ii. GRADUATE

|                               |                        |  |
|-------------------------------|------------------------|--|
| <b>Ms. T. Clarke</b>          | Graduate Student (MSc) | 05/20 - present  |
| <b>Ms. K. Carbonara</b>       | Graduate Student (MSc) | 08/19 - present; <b>Ontario Graduate Scholarship</b> (2020-21) |
| <b>Mr. J. Coish</b>           | Graduate Student (PhD) | 01/19 - present; supervisory committee                         |
| <b>Mr. C. Watson, MSc</b>     | Graduate Student (PhD) | 08/18 - present; supervisory committee                         |
| <b>Mr. N. Kurgan, MSc</b>     | Graduate Student (PhD) | 05/18 - present; supervisory committee                         |
| <b>Mr. M. Almuslehi, MSc</b>  | Graduate Student (PhD) | 10/15 - present; <b>full scholarship (Iraq gov't)</b>          |
| <b>Mr. M.K. Sen, BSc, MSc</b> | Graduate Student (PhD) | 03/15 - 05/20; <b>IPRS Scholarship</b>                         |

|   |                          |  |
|---|--------------------------|--|
| <b>Mr. M. Withers, MIT</b>                | Graduate Student (PhD)   | 09/14 – present; transferred from MSc to PhD program 12/2018 (NB part-time).   |
| <b>Mr. M. Mueller</b>                     | Graduate Student (MSc)   | 01/18 – 12/19; supervisory committee   |
| <b>Ms. B. Herath</b>                      | Graduate Student (PhD)   | 01/16 – 09/19; co-supervisor with Prof. Sathasivan (WSU Engineering)   |
| <b>Ms. D. Dabral, BSc, MSc</b>            | Graduate Student (PhD)   | 03/15 – 06/19; <b>WSU fee waiver. PDF with Prof. G. van den Bogaart, Groningen Biomolecular Sciences &amp; Biotechnology Institute, The Netherlands.</b>   |
| <b>Ms. N. Noaman, BSc</b>                 | Graduate Student (PhD)   | 05/15 – 03/19; <b>MMRG Scholarship.</b> Travel award to present at HUPO 2015 (Vancouver). Transferred from MSc to PhD program 06/2016. Travel & registration award to present at ICAP 2017 (Lisbon). Travel Award to present at ASBMB 2018 (San Diego). Visiting Scholar, Brock University 03/17 – 03/19. Working in industry. |
| <b>Ms. A. D’Silva, MSc</b>                | Graduate Student (PhD)   | 01/15 – 05/19; <b>Medical Research Scholarship from RPA.</b> Research Scientist, U. New South Wales, Australia.  |
| <b>Mr. M. Radzieta, BSc</b>               | Graduate Student (PhD)   | 03/15 – 02/19; <b>Australian Post-graduate Award.</b> Co-supervisor with Dr. S. Jensen   |
| <b>Mr. M.H. Kabir, MSc</b>                | Graduate Student (PhD)   | 10/15 – 11/18; co-supervisor with Dr. M. O’Connor  |
| <b>Ms. K. Da Silva-Antunes, BSc</b>       | Graduate Student (PhD)   | 03/14 – 06/16; co-supervisor with Dr. P. Mudgil & Prof J. Whitehall  |
| <b>Ms. T. Mukaddam, BSc</b>               | Graduate Student (PhD)   | 11/11 – 06/16; co-supervisor with Prof. D. Chang   |
| <b>Ms. C. Gano, BSc</b>                   | Graduate Student (PhD)   | 11/13 – 12/17; co-supervisor with Dr. P. De Souza  |
| <b>Ms. L. Stroud, BSc</b><br>(nee Cronin) | Graduate Student (PhD)   | 03/12 – 12/17; co-supervisor with Dr. C. Stack   |
| <b>Ms. M. Mansouri</b>                    | Med/Grad student (MPhil) | 12/15 – 06/17; WSU co-supervisor. I arranged a research training exchange with ICORD/Blusson Spinal Centre (Vancouver).  |
| <b>Ms. S. Lim, BSc</b>                    | Graduate Student (PhD)   | 03/12 – 02/17; co-supervisor with Dr. M. O’Connor  |
| <b>Ms. M. Partridge, BSc</b>              | Graduate Student (PhD)   | 06/13 – 06/16 (withdrew); <b>MMRG Scholarship</b>  |
| <b>Mr. P. Abbineni, BSc</b>               | Graduate Student (PhD)   | 03/12 – 11/16; <b>WSU School of Medicine Scholarship.</b> Invited talk & Travel award, 18th International Symposium on Chromaffin Cell Biology (2015). <b>PDF with Prof R. Holz, Medical School, U. Michigan 2015-18) &amp; with Prof D. Ginsburg (2019-present).</b>  |
| <b>Mr. M. Van Leeuwen,</b>                | Graduate Student (MSc)   | 03/15 – 11/16; co-supervisor with BSc Dr. M. Gaborieau   |
| <b>Mr. S. Stimpson, BSc</b>               | Graduate Student (PhD)   | 02/11 – 06/16; co-supervisor with Dr. S. Myers   |
| <b>Ms. D. Laurenti, BSc</b>               | Graduate Student (PhD)   | 04/12 – 06/15; co-supervisor with Prof. G. Münch   |
| <b>Mr. E.M. Meijering, BSc</b>            | Graduate Student (MSc)   | 09/15 – 04/16; exchange from U. Utrecht  |
| <b>Mr. K. Ormerod, BSc</b>                | Exchange Fellow (PhD)    | 04/15 – 10/15 ( <b>Endeavour Fellow;</b> from Brock U., Canada, a WSU Partner institution).  |
| <b>Mr. M. Abomughaid, BSc</b>             | Graduate Student (PhD)   | 03/13 – 03/15; U. Sydney Medical School & Westmead Millennium Institute. Co-supervisor with Dr. M. Douglas   |

- Mr. K. Mahadeo**, BSc Exchange Fellow (PhD) 06/14 – 09/14 (Canadian exchange; from U. Calgary). Co-supervised as part of Graduate studies course with Dr. E. Prenner.
- Mr. M. Featon**, BSc Graduate Student (PhD) 10/11 – 04/14; withdrew, co-supervisor with Dr. S. Myers
- Mr. B. Harper**, BSc Graduate Student (PhD) 01/09 – 11/13; co-supervisor with Prof. J. Aldrich-Wright
- Mr. R. Hyland**, BSc Graduate Student (PhD) 01/09 – 03/14; co-supervisor with Dr. S. Myers
- Mr. M. Burgess**, BSc Graduate Student (PhD) 04/11 – 01/13; co-supervisor with Dr. M. Temple
- Ms. V. Mansour**, BSc Graduate Student (PhD) 06/09 – 06/12; **Australian Post-graduate Award. Associate Lecturer (UWS SoM).** (nee Gauci)
- Ms. E. Wright**, BSc Graduate Student (PhD) 06/09 – 06/11; **Australian Post-graduate Award.** PDF, U. East Anglia & U. Kentucky. Teaching associate at WSU.
- Mr. T.G. Tut**, BSc Graduate Student (PhD) 02/11 – 09/11; Transferred
- Mr. C. Hammang**, BSc Graduate Student (PhD) 09/09 – 09/11; co-supervisor with Dr. A. Salih; Transferred.
- Ms. C. Human**, BSc Exchange Fellow (PhD) 10/10 – 04/11 (German exchange; from Helmholtz Institute, Leipzig)
- Ms. K. Furber**, BSc Graduate Student (PhD) 01/04 – 05/10 (Transferred from MSc; Canada Grad. Scholarship-CIHR Master's Award; AHFMR Studentship; Dean's Research Excellence Award; Alberta Grad. Student Fellowship; MDNS Program Award; UC Travel Award; Canada Grad. Scholarship-CIHR Doctoral Award). **Postdoctoral Fellow, U. Saskatchewan. Assist. Prof., UNBC.**
- Mr. M. Churchward**, BSc Graduate Student (PhD) 09/04 – 12/09 (Transferred from MSc; Dean's Entrance Scholarship – Biochem.; PGS NSERC; AHFMR Studentship; NSERC Doctoral Award; Dean's Res. Excellence Award); **NSERC Postdoctoral Fellowship, U. Edinburgh. RA, U. Alberta. Assist. Prof., Concordia U. (Edmonton)**
- Mr. R. Butt**, BSc Graduate Student (PhD) 01/03 – 08/08 (Transferred from MSc; AHFMR Studentship; BMB Gordon Dixon Excellence Award, Graduate Research Scholarship; Alberta Grad Student Scholarship; Faculty of Graduate Studies Award; **Chancellor's Medal for Graduate Research**). **Professor of Chemistry, Marianopolis College (Montreal).**
- Ms. C. Gutierrez**, MSc Graduate Student (PhD) 12/05 – 06/09 (co-supervisor with Dr. M. Colicos; Dean's Entrance Scholarship – Biochem.)
- Ms. M. Ernst**, MSc Graduate Student (PhD) 06/03 - 06/08 (co-supervisor with Dr. A. Kungl, Karl Franzens University, Graz, Austria; Fellowship from Austrian government)
- Mr. R. Taylor**, BSc Graduate Student (MSc) 09/05 – 06/07 (Dean's Entrance Scholarship – Neurosci.; QEII Scholarship; Graduate Research Stipend). Completed Medical School, U. Calgary.
- Ms. F. Rhemtulla**, BSc Graduate Student (MSc) 09/04 - 11/06 (Alberta Scholarship). Completed Medical School, U. Calgary.
- Ms. H. Munib**, BSc Volunteer 12/05 - 08/06
- Ms. J. Herrmann**, BSc Exchange Grad Student 01/05 - 07/05 (from U. Munich); (nee Höfgen) Completed PhD. Grant/patent Officer, U. Dresden.
- Mr. J. Szule**, MSc Graduate Student (PhD) 09/01 - 07/05 (PGS NSERC; AHFMR Studentship; Canada Grad. Scholarship-CIHR Doctoral Award); **NSERC Post-doctoral Fellowship at Stanford. Research A/Prof, Texas A&M.**

|                                |                            |  |
|--------------------------------|----------------------------|--|
| <b>Ms. J. Hibbert</b> , BSc    | Graduate Student (MSc)     | 01/03 - 11/04 (Alberta Graduate Student Award. Staff, Alberta Bone & Joint. Clinical Trials Coordinator, Centre for Neonatal Research and Education, U. Western Australia) |
| <b>Mr. J. Kirshtein</b> , BSc  | Exchange Graduate Student  | 08/02 - 08/03<br>(Visiting from Johns Hopkins University; completed PhD & working as scientist in industry)  |
| <b>Ms. N. Orton</b> , BSc      | Graduate Student (MBT)     | 10/01 - 08/02  |
| <b>Ms. S. Horn</b> , BSc       | Exchange Grad Student      | 07/02 - 12/02 (from U. Berlin)   |
| <b>Mr. Y. Belmalha</b> , BSc   | Exchange Student           | 03/02 - 04/02 (from U. Amsterdam)  |
| <b>Ms. L. Ma</b> , BSc         | Graduate Student (MSc)     | 09/01 - 11/01 (BMB rotation)   |
| <b>Ms. D. K. Song</b> , BSc    | Graduate Student (MSc)     | 09/01 - 12/01 (withdrew; health)   |
| <b>Mr. F. Albertorio</b> , BSc | Predoc Fellow (MSc equiv.) | 01/99 - 05/01<br>(MARCS Fellow; completed PhD (Chemistry) at Texas A&M and a PDF at Harvard)   |

### iii. POSTGRADUATE

|                                  |                     |  |
|----------------------------------|---------------------|--|
| <b>Dr. Zahra Farahnak</b>        | Postdoctoral Fellow | 06/20 - present                                  |
| <b>Dr. Anu Shanu</b>             | Postdoctoral Fellow | 04/15 - present†                                 |
| <b>Dr. Elise Wright</b>          | Postdoctoral Fellow | 06/11 - 12/11 & 03/12 - 05/14; HUPO Travel Award |
| <b>Dr. Victoria Gauci</b>        | Postdoctoral Fellow | 07/12 - 02/13                                    |
| <b>Dr. Kendra Furber</b>         | Postdoctoral Fellow | 06/10 - 02/12                                    |
| <b>Dr. Sina Ahmadi Pirshahid</b> | Postdoctoral Fellow | 04/04 - 02/05                                    |
| <b>Dr. Amrisha Verma</b>         | Postdoctoral Fellow | 09/99 - 06/02 (Fogerty Fellow)*                  |
| <b>Dr. Miroslava Stastna</b>     | Postdoctoral Fellow | 05/99 - 09/01 (Fogerty Fellow)*                  |
| <b>Dr. Irina Kolosova</b>        | Postdoctoral Fellow | 09/99 - 09/00 (Fogerty Fellow)*                  |

†co-supervised with Dr. S. Myers (WSU)

\* co-supervised with Dr. J. Zimmerberg (NIH)

### SUPERVISORY COMMITTEES

- I currently supervise/co-supervise 5 PhD and 2 MSc.
- Sat on 16 PhD student supervisory committees (chaired 6) in Canada.
- Have graduated 11 PhD and 5 MSc from my group; I have co-supervised another 23 PhD (many doing most of their research within my group).
- Have sat on 6 MSc supervisory committees (chaired 3; UC); 3 MSc committees at Brock.
- Have sat on 2 MBT student supervisory committees (UC).
- Mentor for first year students in the Faculty of Medicine BHSc program (UC).

## VI. ADMINISTRATIVE RESPONSIBILITIES

### i. DEPARTMENTAL (including thesis exams internationally)

- Examiner, Habilitation thesis: Dr. M. Frick, **University of Ulm, Germany** (04/12)
- Member of 19 PhD Examining Committees (other than as Supervisor):
  - Ms. K. Wongtrakul-Kish (Dept Chemistry & Biomolecular Sciences, Faculty of Science, Macquarie University, Sydney, Australia; 08/15)
  - Ms. X. Li (Nanyang Technological University, **Singapore**; 12/14)
  - Mr. C-M. Clausson (**External Examiner** ('Faculty Opponent') for Biomedical Centre, **Uppsala University, Sweden**; 03/14)
  - Ms. A.B.H. Quan (CMRI / Sydney Medical School, University of Sydney, Sydney Australia; 01/14)
  - Ms. N.A. Leneva (Institute for Molecular Bioscience, University of Queensland, Brisbane, Australia; 10/13)
  - Ms. L.-S. Chan (CMRI / Sydney Medical School, University of Sydney, Sydney Australia; 01/13)

Mr. M.J. Medynskyj (Dept Chemistry & Biomolecular Sciences, Faculty of Science, Macquarie University, Sydney, Australia; 10/12)  
Mr. T. Couttas (School of Biotechnology and Biomolecular Sciences, University of New South Wales, Sydney, Australia; 07/12)  
Ms. Y. Zhou (Nanyang Technological University, **Singapore**; 03/12)  
Ms. D. Cheng (School of Medical Sciences, University of New South Wales, Australia; 01/11)  
Ms. R. Flynn (UC Faculty of Medicine; 08/10)  
Ms. M. Vorland (**External Examiner** ('First Opponent') for Faculty of Medicine, **University of Bergen, Norway**; 05/08)  
Mr. A. Klimowicz (UC Faculty of Medicine; 01/08)  
Mr. G. Slysz (UC Faculty of Medicine; 09/07)  
Ms. K. Maxwell (UC Faculty of Social Sci; 09/07)  
Mr. T. Dunn (UC Faculty of Medicine; 06/07)  
Mr. D.W. Munno (UC Faculty of Medicine)  
Mr. D. Schibli (UC Faculty of Science, Biology)  
Ms. A. Ulke-Lemee (UC Faculty of Science, Biology)

- Member of 7 MSc Examining Committees (other than as Supervisor):
    - Mr. M. Bader (UC Faculty of Medicine)
    - Ms. K. Cochrane (UC Faculty of Medicine)
    - Ms. N. Van (UC Faculty of Medicine)
    - Ms. J. Howell (UC Faculty of Science, Biology)
    - Mr. B. Vartien (UC Faculty of Medicine)
    - Ms. P. Pratikhya (UC Faculty of Medicine)
    - Mr. M. Mueller (Brock Faculty of Mathematics & Science)
  - Member of 27 Candidacy Examining/Review Committees (other than as primary Supervisor):
    - D.W. Munno, S. Sangha, A. Klimowicz, G. Slysz, J. MacCallum, T. Dunn, K. Maxwell, R. Flynn, T. Christie, T. Nguyen (UC Chemistry, Qualifying Oral Exam); C. Hammang; M. Burgess; S. Stimpson; T. Mukaddam; D. Laurenti; S. Lim; M. Mangala; L. Cronin; M. Featon; A. Sutton; E. Whitty; M. Toutounji; M. van Leeuwen; S. Aggarwal; K. Da Silva-Antune; N. Kurgen; C. Watson
  - Member of 3 PhD proposal defence committees at Brock U. (other than as primary Supervisor):
  - Neutral Chair for 19 PhD/MSc examinations (2016-present)
  - 10 Honours Thesis examinations (2005-present); Introduction reviews — 4 (2012-13).
  - Supervision and examination of 8 Third Year Biomolecular Projects (2010-2013).
  - Reserve examiner for PhD thesis, APAF/Macquarie University (2011)
  - Chair for 2 Candidacy Examining Committees (Ms. J. Beveridge, U of C Med. Sci. Grad Program; Ms. S. Aggarwal, WSU (2015))
  - Member of 6 MBT examining committees, and have also served as a reviewer for final formal student presentations in this program (in 2006)
  - Reserve examiner for PhD thesis in Dept Obstetrics & Gynaecology, University of Melbourne /Royal Women's Hospital, Victoria, Australia (2007)
  - U of C Secondary appointments: Depts. Biochemistry & Molecular Biology and Cell Biology & Anatomy (2001 – 2008)
  - Member, NRG website update committee (UC; 2001-2002)
  - Member, National Graduate Student Competition subcommittee (UC; Neuroscience; 2003-2006)
  - Member, BMB Committee on Modular Courses (UC; 2004-2005)
- ii. SCHOOL / COLLEGE / FACULTY
- Member, Epidemiologist search committee; Dept. Health Sciences, FAHS (01/2020-present)
  - Member, advisory committee for FAHS Associate Dean of Research & Graduate Studies (11/2019 – 2/2020)
  - Member, FAHS proposal panel for new CRC Tier I and Tier II positions.

- Member, FAHS ranking panel for NSERC Doctoral Scholarship applicants (10/2018)
- **Dean, Faculty of Graduate Studies, Brock University** (07/2016-09/2017)
- **Panel member for Counsellors Day**; represented Brock University Graduate Studies
  - presentation to regional high school and college guidance counsellors (8/12/2016).
- Member, search panels for FGS Communications, Marketing & Recruitment Officer, and FGS Vitae Support Assistant (08-09/2016)
- **WSU School of Medicine Executive Committee** (03/2008-05/2016)
  - Research Committee restructure working group (2010)
  - MBBS/MD restructure working group (2016)
- **WSU School of Medicine Research & Higher Degree Committee** (05/2011-05/2016)
- **WSU School of Medicine Curriculum Committee** (03/2008-05/2016)
  - Biomedical Science Program Review subcommittee (01/2011- 01/2012)
- **Coordinator, WSU School of Medicine Internationalisation meetings in Canada** (03 & 10/2014; 03-04, 06, 09, and 10/2015).
- WSU School of Medicine Year 1 & 2 Management Committee (01/2012-05/2016)
- WSU School of Medicine Year 3 & 5 Management Committee (01/2012-05/2016)
- **Conjoint appointment in the WSU School of Biomedical and Health Sciences** (now School of Sciences and Health, 09/2008-05/2016)
- Member, WSU Nanoscale Organisation & Dynamics Group (09/2008-05/2016)
- **WSU School of Medicine Lab User Group Committee** (04/2008-05/2016)
- **WSU Mass Spectrometry Facility oversight committee** (09/2009-05/2016)
- **WSU School of Medicine**, admission interview panel, MBBS candidates (12/2012-05/2016)
- Member, WSU School of Medicine internal grant review panel for NHMRC submissions (2015-2016).
- **Requested Member**, review panel for Narellan Rotary Club Scholarship for first year medical students: a community-based award (2009-2015).
- **Chair**, three separate MMRG grant review panels (2015)
- **Chair**, Review panel for PhD Scholarship applicants to the School of Medicine (05/2015)
  - To review applications as well as train junior faculty & postdoctoral fellows as to the operation of international calibre grant review panels.
- Member, Selection Committee for MMRG Postdoctoral Fellow (Neuro; Myers' lab; 05/2015)
- Member, Selection Committee for Acting Head of the Centre for Health Research (01/2015)
- Member, Selection Committee for a Coordinator of the SoM Animal Facilities (07/2013)
- UWS Bio-Imaging Facility oversight committee (03-12/2011)
- College mentor (with Prof. Sucher) for new Lecturer/Manager of the UWS CHS Mass Spectrometry Facility (02/2010-03/2011)
- UWS School of Medicine Year 2 Management Committee (03/2008-12/2011)
- UWS School of Medicine Year 4 Management Committee (01-12/2011)
- UWS School of Medicine Year 5 Management Committee (01-12/2011)
- UWS School of Medicine Year 3-5 (revised to Year 4/5) Curriculum Planning Committee (03/2008-12/2010)
- **Spearheaded establishment and signing of an MOU between the UWS School of Medicine / Molecular Medicine Research Group and Bio-Rad Australia** (signed 16/09/2010).
- **Talk on Professionalism and Career Development at the UWS School of Medicine/CHS Research Day** (2/12/2010).
- Chair, Selection Committee for a Senior Scientific Officer in Molecular Physiology (04/2010).
- Chair, Selection Committee for an Associate Lecturer in Molecular Physiology, UWS (10/2009).
- **Acting Dean of UWS School of Medicine** (28/2005 – 6/2006, 2009)
- **Member, search committee for new UWS Dean of Medicine** (03-05/2009)

- UWS School of Medicine retreat to review Years 1 & 2 of the Medical program (09/2008)
- UWS School of Medicine retreat - Planning for Success & Progressing Research (04/2008)
- Chair, Selection Committee for a Scientific Officer in Molecular Physiology, UWS (2008).
- Chair, Selection Committee for an Administrative Officer in Molecular Physiology, UWS (2008).
- **Co-Leader of the Epilepsy & Brain Circuits Research Program** within the Hotchkiss Brain Institute, U of C (10/2006 – 01/2008)
- **U of C Faculty of Medicine subcommittee for internal accreditation review** (2007)
- **HBI representative on the Finance & Review Committee for the Centre for Advanced Technologies** (2006–2007)
- **Member, UC Steering Committee for Proteomics & Functional Genomics** (2005-2007)
- **Member, Steering Committee, Southern Alberta Mass Spectrometry Centre** (2004-07)
- **Lead PI; Faculty of Medicine CFI proposal: Centre for Molecular Interactions in (Patho)Physiology**
- **Member, Biomedical Technical Support Centre Committee** (07/2001 - 06/2004); **Chair** 06/2003 - 06/2004.
- **Member, Graduate Education Committee** (Neuroscience 2001 - 2006)
- **Member, Graduate Education Committee** (MBT Program)
- **Member, Graduate Course Review Committee** (Neuroscience, 2006-2007)
- **Neuroscience Candidacy Review Committee** (10/2004-12/2005)

iii. UNIVERSITY

- **Board of Trustees** (elected), Brock University (06/2018-06/2021)
  - University and Community Experience Committee (07/2020-06/2021)
  - Capital Infrastructure Committee (06/2018-06/2021)
- **Senator** (elected), Brock University Senate (06/2018-06/2021)
  - **Chair, Senate Governance Committee** (06/2018-06/2022)
  - Planning Priorities and Budget Advisory Committee (06/2018-06/2022)
  - Two-Row Council (Indigenous Engagement; 09/2019-06/2022)
  - Working Group on External review of Senate (10/2018-07/2019)
- **Member**, Advisory Committee on the Appointment of a Vice-President, Administration (04/2020-present).
- **Member**, Advisory Committee on the Appointment of a Vice-Provost, Indigenous Engagement (04/2018-03/2019).
- CAUT Grievance Handling Workshop (27-28/02/2019).
- **Member**, review panel, OGS Doctoral Scholarships (02/2019).
- **Chair** (elected), Senate *ad hoc* committee to address the Maclean's rankings (01-05/2018).
- **Brock Representative**, DAAD (German Academic Exchange Service) Special Information Program/Tour: Germany Today 2018 - Getting to know Germany's Universities of Applied Sciences. One of only 6 Canadians with 16 American institutions represented (06/2018).
- **Spearheaded establishment of Academic Cooperation Agreements** between Brock University and (i) University of Ljubljana (Slovenia); (ii) NOVA University (Portugal); and (iii) Mount Royal University (Calgary) (2016-2018).
- **Dean, Faculty of Graduate Studies, Brock University** (07/2016-09/2017)
- **Brock University Senior Administrative Council** (07/2016-09/2017)
  - **Brock University Senate** (07/2016-09/2017; *Ex Officio*, as Dean of Graduate Studies)
    - Teaching & Learning Policy Committee (09/2016-09/2017)
    - Graduate Studies Committee (07/2016-09/2017)
    - Research & Scholarship Policy Committee (07/2016-09/2017)
- **Brock University Human Rights Task Force** (10/2016-01/2017)
- **Brock University Council of Academic Deans** (07/2016-09/2017)
- **Brock University Graduate Council** (Chair; 07/2016-09/2017)

- **Brock University Academic Review Committee** (07/2016-09/2017)
- **Brock University Spring/Summer Review Committee** (09/2016-09/2017)
- **Brock University SMA Enrollment planning committee** (11/2016-03/2017)
- **Brock University Advisory Committee on Space** (11/2016-09/2017)
- **Brock University Graduate Bursary Committee** (07/2016-09/2017)
- **Member, Selection panel for Canada 150 Research Chair applicants** (08/2017).
- **Chair, Brock Chancellor's Chair for Research Excellence and Distinguished Research Awards Committee** (06/2017)
- Member, Counsellors Day Panel. Representing FGS in presentations to high school and college guidance counsellors (08/2016)
- Member, search panels for Director of Brock International, FGS Communications, Marketing & Recruitment Officer, FGS Vitae Support Assistant (07-09/2016; 05/2017)
- **Extensive international recruitment trips for WSU**, with emphasis on the School of Medicine (2009-2016).
- **Named to Head the new WSU Molecular Medicine Research Group** (08/2009-05/2016).
  - Successfully coordinated first external 5 yr review (05-09/2015); recognized as highly successful and comparable to many large research institutes in Australia.
- **School of Medicine representative on the School Academic Committee** for Science & Health (04/2012-05/2016).
- Search committee for Mass Spectrometry Lecturer / Facility Manager (2016; 2012; 2009)
- Reviewer for promotion to Senior Lecturer, School of Science & Health (2015).
- **Panel member, 4<sup>th</sup> Vice-Chancellor / Chair of Academic Senate Forum - Enhancing the Student Experience** (02/2015).
- **WSU Professorial representative for a meeting with IDP Education Ltd.; Q & A with staff concerning the WSU Graduate and Medical Programs** (Singapore, 15/12/2014).
- **Member, Coordination Advisory Group to Executive: Master of Research/Pathway to PhD Program** (07-09/2014). Expert panel of 3 + chair, to design effective transition program.
- Reviewer for promotion to Associate Professor, School of Medicine (2014).
- **Panel member, Graduate Research Supervisors Forum** (07/2014).
- CO-ORGANIZER (with Prof Münch and Dr. O'Connor) of the WSU Molecular Medicine Research Group Annual Meeting (WSU Campbelltown, 09/2013).
- **WSU representative at Biomedical Science Education into the 21<sup>st</sup> Century**, a National Forum on Education in the Biomedical Sciences hosted by the Academy of Science. This also formally opened the Australian Collaborative Universities Biomedical Education network (CUBENet), a new national initiative to improve education in the Biomedical Sciences (Canberra, 12/2011 and 12/2013).
  - Member, CUBENet working parties on Emerging/Future Health Research & Academic Prerequisites (2011-2016)
- Search committee for Lecturer/Mass Spectrometry Facility Manager (09-10/2012)
- **Founding Member, Integrated Development Program** to enhance Honours & Graduate teaching & professional development for UWS biomedical/science/health students (10/2011).
- **Member, UWS Research Studies Committee of Academic Senate** (04/2009-12/2011)
- Member, Level E (Full Professor) Promotion Review Committee (10/2010).
- **Spearheaded establishment and signing of an Academic Cooperation Agreement between WSU and Brock University** (12/2010); renewed 10/2015.
- **Assigned Member, Australian Trade Commission Life Sciences Delegation to Canada** (11/2010).
- **Spearheaded establishment and signing of an Academic Cooperation Agreement between UWS and McMaster University** (08/2010).
- Brief research overview for official opening of the WSU/CHS Mass Spectrometry Facility (07/2010); a **team led by Profs Sucher & Coorssen secured the initial funding for these instruments and thus for establishment of this critical new facility at UWS.**

- College review/hiring panel for Research Lectureships; principal assessor of Molecular Medicine applications (07/2009) - successful recruitments in NMR & Stem Cells/Regeneration Medicine.
  - These have proven to be two of the top young research/teaching faculty at WSU.
- **Research Review Panel for the WSU College of Health and Science** (Internal panel member, 11/2008).
- Official 'mentor' to new junior professor in U of C Dept. Chemistry (09/2007-01/2008)
- **Invited academic, the President's Life Sciences Research Workshop** (U. Calgary; 11/2003); identifying areas of strength and weakness, identifying priority areas for development & strategic planning. Also asked to join second focus group workshop (01/2004).

iv. (INTER)NATIONAL

- **Member**, Council of Graduate Studies; Council of Ontario Universities (07/2016-09/2017).
- **Adjunct Professor**, School of Medicine, Western Sydney University (07/2016-present).
- **Adjunct Professor**, Faculty of Medicine, University of Calgary (01/2010-present).
- **Honorary Professor**, School of Medicine, University of Sydney (06/2014-present).
- **Associate Member**, ICORD, Blusson Spinal Centre / U. British Columbia (04/2012-present).
- **Invited delegate**, Precision Medicine & Biomarkers Leaders Summit: USA, Boston, USA (7-8/05/2018); asked to consult with industry leaders on translational 'omics approaches.
- **International Reviewer, Knowledge Foundation (KK-Stiftelsen), Sweden** (2017).
- **International Reviewer, Polish National Science Centre** (2016).
- **External reviewer for promotion to Associate Professor**, School of Science & Health, Western Sydney University, Australia (2016).
- **Foreign Member**, EU Cooperation in Science & Technology (COST-FAP1002; 2012-2015).
- **Requested External Reviewer, CIHR Foundation Scheme (Stage 3): 1<sup>st</sup> Live Pilot competition**; by arrangement between the NHMRC (Australia) and CIHR (Ottawa, 06/2015).
- **Member, Rebecca L Cooper Medical Research Foundation Scientific Advisory Committee** (2014-2015).
- **International Reviewer, German-Israel Foundation for Scientific Research & Development** (2015).
- **External reviewer for promotion to Professor**, Faculty of Medicine, U. Calgary, Canada (2015).
- **External reviewer for promotion to Associate Professor**, Faculty of Medicine, Kuwait U., Kuwait (2015).
- **Invited delegate**, 2<sup>nd</sup> International Precision Medicine Congress, London, UK (2015); asked to consult with industry leaders on translational 'omics approaches.
- **Member, MRC (UK) International College of Experts** (in partnership with Department of Health, Wellcome Trust, Cancer Research UK, British Heart Foundation, Arthritis Research UK and UK Research Councils; 2014).
- **Member, NHMRC Assigners Academy** (Biochemistry & Cell Biology; 2013 & 2014).
- Reviewer for the **Qatar National Research Fund** (2009-2013).
- External reviewer for promotion to Professor, Case Western Reserve U., USA (2013).
- Appointed co-Supervisor (i.e. 'Auxiliary') at Sydney Medical School, the Faculties of Dentistry & Pharmacy, and Sydney Nursing School (including Royal Prince Alfred Hospital; 03/2013).
- Habilitation examiner for the University of Ulm, Germany (03/2012).
- Member, NHMRC Neurosciences Peer Review Panel, Canberra (08/2011 & 2012).
- Member, NHMRC Biochemistry Peer Review Panel, Canberra (08/2010).
- Member, CIHR Cell Biol & Mechanisms of Disease Peer Rev Panel, Ottawa, Canada (05/10).
- Appointed as an ARC Assessor (04/10); ad hoc operating and fellowship reviews (2009-13).
- Reviewer for the Royal Society of Chemistry, Joseph Black Award (2010).
- Reviewer for the Royal Society, International Joint Project Grant Program (2009, 2010).

- Australian link, Royal Society of Chemistry travel award application by Dr. Carlos Lodeiro Espino, Dept. Physical Chemistry, University of Vigo, Spain (2010).
- Invited participant (one of only two from Australia) in the Asian Protein Science Forum sponsored by Waters Corporation (Beijing, August, 2009).
- External member, Canadian Institutes of Health Research Cell Physiology grant review committee (2007 - 2009).
- Reviewer for the Deutsche Forschungsgemeinschaft (2008-2010)
- Ad hoc external reviewer for NHMRC grants (2009-13)
- Ad hoc external reviewer for Natural Sciences & Engineering Research Council of Canada, Discovery Grants Program (2009)
- **Invited participant in the Australian Academy of Sciences – Theo Murphy High Flyers Think Tank** 2008 on "Preventative Health: The use of Science & Technology in Prevention and Early Detection of Disease (University of Sydney, November, 2008).
- **Invited expert, Health Genomics and Proteomics Workshop sponsored by Genome Alberta and the Alberta Heritage Foundation for Medical Research**; 30 experts asked to identify priority areas and strategic funding opportunities to guide Alberta investment in health related genomics and proteomics (2007).
- **Requested participant in American Society of Cell Biology workshop on international membership interests** (2006)
- Member, PDF Fellowship Review Committee, CIHR (2006-2008)
- Member, Graduate Studentship Review Committee, Canadian League Against Epilepsy (06-09)
- Member, Calgary Health Region Perinatal Grant Review Committee (2007)
- Member, Graduate Studentship Review Committee, Alberta Heritage Foundation for Medical Research (2004-2007)
- Member, Ph.D. Fellowship Rev. Committee, Heart & Stroke Foundation of Canada (2004-2006)
- Member, Operating Grant Review Committee, Canadian Breast Cancer Foundation (AB/NWT Chapter; 2004-2005)
- **Invited workshop leader for the Canadian Medical Hall Of Fame Discovery Days in Health Sciences at U of C: Introduction to Functional Proteomics** (2003-2006)
- **Invited workshop leader for the Shad Valley International Program**, July 8-10, 2003
- **Supporting speaker for the Canadian National Site Licensing Project** (2001-2005); comments on innovation in the 21<sup>st</sup> century used as the opening statement on the Canadian Research Knowledge Network website.

## VII. PROFESSIONAL ACTIVITIES

### i. MEMBERSHIP IN PROFESSIONAL AND LEARNED SOCIETIES

Biochemical Society  
American Society for Cell Biology  
American Society for Biochemistry and Molecular Biology  
International Society for Neurochemistry (2008-2017)  
Human Proteome Organisation (2009-2016)  
Australasian Proteomics Society (2008-2016)  
Collaborative Universities Biomedical Education Network (CUBENet; 2010-2016)  
Australian Physiological Society (2008-2016); Founder of the Membranes Special Interest Group (with Drs. Thorn & Keating; 2010)  
New York Academy of Sciences (2000-2010)  
Canadian Society of Biochemistry, Molecular & Cellular Biology (2001-2008)

### ii. PROFESSIONAL SERVICE

- MEMBER, Editorial Board, **High-Throughput** (2017 – present)
- MEMBER, Editorial Board, **Proteomes** (2015 – present)
  - **Guest editor**, special issue on Feature Review Papers ('status of the field') (2020-21).
  - **Guest editor**, special issue on Top-Down Proteomics (2016-17, 2018-19)

- MEMBER, Editorial Board, **DNA and Cell Biology** (2011 – present)
- Editor-in-Chief, **Journal of Integrated OMICS** (2009 – present; a founding editor)
  - -Regional Editor for Australia & New Zealand (07/2011 – 09/2012)
- MEMBER, Editorial Board (Biochemistry), Dataset Papers in Biology (2012 – 2017)
- MEMBER, Editorial Board, **PLoS ONE** (2012 – 2017)
- MEMBER, Editorial Board, **Journal of Chemical Biology** (2007 – 2017); in 01/2018, journal merged with SpringerOpen Chemistry Central Journal.
- MEMBER, Editorial Board, **Open Proteomics Journal** (2007 – 2013)
- MEMBER, **Scientific Committee of the 6th International Congress on Analytical Proteomics** (Portugal, 07/2019).
  - Opening Plenary Address: Defining a better mouse model for Multiple Sclerosis research.
  - Session chair.
- MEMBER, **Scientific Committee of the 5th International Congress on Analytical Proteomics** (Portugal, 07/2017).
  - Keynote Address: Post-Translational Modifications Predictive for Preterm Birth.
- MEMBER, **International Scientific Committee of the 4<sup>th</sup> International Congress on Analytical Proteomics** (Portugal, 09/2015).
  - Closing Plenary Address: High Resolution Top-Down Proteomics.
  - Evaluator, Young Researcher Prizes.
  - Session chair.
- MEMBER, Local Organising Committee for the **18th International Symposium on Chromaffin Cell Biology** (Cairns, Australia; 08/2015).
- **Invited Presenter/Moderator/Instructor, International Proteomics Conference/6<sup>th</sup> Indian Proteomics Society Meeting**, Indian Institute of Technology (Mumbai, India, 2014)
  - INVITED TALK (Education Day): Functional proteomics.
  - INVITED MODERATOR (Biopharma Industry day): Proteomics perspective on development & characterization of Biosimilars and Biotherapeutics.
  - INVITED INSTRUCTOR (Proteomics Workshop): Top-down, gel-based proteomics.
- MEMBER, scientific review panel and session chair, 2<sup>nd</sup> Proteomics & Beyond Symposium (Macquarie U., 12/11/2014).
- **Session organiser & co-chair, by request of the Chinese Academy of Sciences, 18<sup>th</sup> International Union of Pure and Applied Biophysics Congress** (Australia, 08/2014).
- MEMBER, **International Scientific Committee of the 1<sup>st</sup> International Symposium on Nanoparticles/Nanomaterials and Applications** (Portugal, 2014).
- MEMBER, **International Scientific Committee of the 3<sup>rd</sup> International Congress on Analytical Proteomics** (Brazil, 2013). **Keynote address.**
  - SESSION CHAIR: Functional Proteomics.
- **GUEST EDITOR, special issue of Cell Calcium** on 'Regulated Exocytosis' (2012)
- MEMBER, organising committee of the Sydney Cell Imaging Forum (05/2011 – 06/2016)
- MEMBER, **International Committee of Chairs, 2<sup>nd</sup> International Congress on Analytical Proteomics** (Spain, 2011).
  - SESSION CHAIR: Quantitative Proteomics
- CO-ORGANIZER (with Dr. P. Thorn) of the symposium 'Exocytosis and Endocytosis' for the **2011 Hunter Cell Biology Meeting** (Hunter Valley, 03/2011).
- CO-ORGANIZER (with Dr. P. Thorn) of the symposium 'Molecular Physiology & Membrane Dynamics' for the **2010 Joint Annual Meeting of the Australian Physiological and Biophysics Societies** (Adelaide, 11/2010).
- FOUNDER (with Drs. Thorn & Keating) of the **Membranes Special Interest Group**, Australian Physiological Society (2010).
- MEMBER, **Local Organizing Committee for the Human Proteome Organization 2010 Annual Meeting** in Sydney, Australia (09/2010).
- CO-ORGANIZER, WSU MMRG symposium 'International Efforts in Translational Medicine' (UWS, Campbelltown, 16-18/09/2010). **Satellite Meeting of the Human Proteome Organization 2010 Annual Meeting.**
- MEMBER, **International Advisory Committee for the 4<sup>th</sup> Shanghai International Conference on Biophysics and Molecular Biology** (China, 08/2010).
- SESSION CHAIR, 'Protein Screening, Bioassay and Discovery' in the Protein Technologies symposium at the **3<sup>rd</sup> Annual Protein and Peptide Conference** (Beijing, 03/2010).

- ORGANIZER, WSU MMRG Symposium 'New Directions in Membrane Trafficking Research' (WSU, Campbelltown, 4/02/2010). **Official Satellite Meeting of the 2010 Joint Annual Meeting of the Australian Physiological and Neuroscience Societies.**
- CO-ORGANIZER (with Dr. D. Keating) of the symposium 'Mechanisms Underlying Exocytosis & Endocytosis: Potential Relevance to Health & Disease.' **2010 Joint Meeting of the Australian Physiological and Neuroscience Societies** (Sydney, 02/2009).
- MEMBER, Editorial Advisory Board, **Biochemical Journal (2002 – 2009)**
- External reviewer for Faculty Promotion to tenured Professor, Dept Chemistry and Biochemistry, University of Minnesota (2009).
- External reviewer for Faculty Promotion to tenured Professor, Cornell University (2008).
- External reviewer for Faculty Promotion to 'tenured status' in the Faculty of Veterinary Medicine, U. Calgary (2008)
- Member, Selection Committee for a Biophysical Chemist, Chemistry Dept., U. Calgary (2007)
- External reviewer for Faculty Promotion (Associate Professor) at University of Alberta (2006)
- Member, Selection Committee for a Senior Lecturer, Medical School, University of Western Sydney, Campbelltown, NSW, Australia (2006)

#### Industry Interactions

- Proteomics for PlantForm Corp. (2018-present); batch characterization and recommendations for process refinement.
- Proteomics for BioDiem Ltd (2014-2017), to define mechanism of action of an antimicrobial agent; in collaboration with A/Prof S. Jensen (UWS SoM & Ingham Research Institute).
- Pre-publication review for an engineering text on Physiology; Springer-Verlag (2015).
- Beta test site for Bio-Rad Laboratories (2010); new isoelectric focussing unit.
- Proteomics consultation work for PerkinElmer Life Sciences (2002-2007).
- Consultant for Garland Science/Taylor & Francis Grp; reviewed planned scientific text (2006).
- Beta test site for Pierce Chemical Company; membrane protein extraction reagents (2005).
- Consultation work for Pfizer Canada, HTS Technologies Ltd (UK), and Coalesce Corp. (Biological Microscopy).

#### iii. GRANT REVIEWS

- Knowledge Foundation (KK-Stiftelsen, Sweden), University-Industry Project Grant (2017).
- Polish National Science Centre, New Investigator Research Grants (2016)
- Women's & Children's Hospital Foundation (SA), Research Grants (2015)
- German-Israeli Foundation for Scientific Research and Development (2015)
- Canadian Institutes of Health Research 1<sup>st</sup> Pilot Foundations Scheme (2015)
- Rebecca L Cooper Medical Research Foundation, Australia (2014 Grants; 2015 Fellowships).
- MRC (UK) International College of Experts. Enhancing UK's Clinical Research Capabilities and Technologies (2014).
- Royal Society, International Joint Project Grant Program (2009 & 2010)
- Deutsche Forschungsgemeinschaft (2008-2011)
- Royal Society of Chemistry, Joseph Black Award (2010)
- NHMRC Operating Grants Program (2009-2013)
- ARC Discovery Grants and Fellowship Programs (2009-2013)
- NHMRC Fellowships Program (2009)
- Natural Sciences & Engineering Research Council of Canada (Cell Physiology)
- Canadian Institutes of Health Research (Neuroscience; Cell Physiology; Pharmacology & Therapeutics); external member of Cell Physiology panel from 2007-2009.
- Heart and Stroke Foundation of Canada
- Telethon Foundation (Italy)
- Kentucky Science and Engineering Foundation

iv. PATENT REVIEWS

- External expert review and declaration for resubmission of a patent in the area of protein detection (2005). Product entered the market shortly thereafter and still holds a unique position.

v. JOURNAL REVIEWS

- ad hoc reviews for:

Journal of Cell Science  
Electrophoresis  
Proteomics  
J. Neuroscience Res.  
Molecular Biology of the Cell  
Trends in Cell Biology  
Cancer Letters  
Future Medicine  
FEBS Journal (Eur. J. Biochem.)  
BBA-Biomembranes  
Experimental Lung Research  
Biochem & Cell Biol.  
Neurochem. Intl.  
J. Cellular Physiology  
Journal of Chromatography B  
Molecular & Cellular Proteomics  
J. Molecular Neuroscience  
Cell Calcium  
BMC Genomics  
Journal of Integrated OMICS  
Nature Protocols  
Proteomics: Clinical Applications  
Proteomes  
Med One  
Pregnancy Hypertension

Biochemical Journal  
Journal of Neurochemistry  
Physiological Reviews  
Journal of Neurophysiology  
Biophysical Journal  
J. Proteome Res.  
Drugs in R & D  
Biochemistry  
Arch. Biochem & Biophys  
Channels  
Toxicon  
Open Proteomics Journal  
PLoS One  
Journal of the American Chemical Society  
Journal of Neuroscience Methods  
BBA-Cell Biology of Lipids  
Journal of Proteomics  
Environment & Natural Resources Research  
Hypertension in Pregnancy  
Curr. Protein & Peptide Science  
Neuroscience  
Expert Review of Proteomics  
Biology Methods & Protocols  
High-Throughput  
Scientific Reports (NPG)

OTHER

- During pandemic lockdown, 'attended' ASBMB Spotlight Webinars "Enzyme Regulation by Filamentation and Other Alternate & Emerging Mechanisms" (27/05/20) & "Shape Shifting in the Control of Protein Function" (28/05/20); Technology Networks webinar "Speeding Up Biological Imaging Using the Infinity Mirror".
- Faculty/Dept representative, Brock Open House (04/2019).
- Moderator/Host for the 12<sup>th</sup> Mapping the New Knowledges Graduate Conference; Brock University (04/2017).
- Poster judge, 18<sup>th</sup> ISCCB Meeting; Cairns, QLD (08/2015).
- Poster judge, 20<sup>th</sup> Lorne Proteomics Symposium; Lorne, Vic (02/2015).
- Poster judge, Proteomics & Beyond Symposium, Macquarie University (11/2012 & 2014).
- **Seminars and small research project with local high school students** (via CSIRO Scientist in Schools Program); Wollondilly Anglican College (05-07/2014 & 2015).
- Poster judge, WSU SSH/SoM Graduate Research Forum (06/2014).
- **Invited Attendee, Natural Products Research Conference**, RMIT; to facilitate interactions across universities and discipline areas (5/12/2013).
- **Session chair, SoM/SSH Research Futures Forum** (12/07/2013).
- Poster judge, European Cooperation in Science & Technology Meeting. Slovakia (04/2013).
- **Introductory research talk as part of an acknowledgement to the Narellan Rotary Club** for funding to the MMRG to initiate a research program on Multiple Sclerosis (11/2012).

- Poster judge, WSU College of Health & Science Student & Postdoctoral Conference (11/2011).
- **Adjudicator for NSW Brain Bee**, 2008 & 2011
- Attended, "Intracellular Signalling, Calcium Ions and Membrane Trafficking" meeting, Scientific Institute San Raffaele, Milan, Italy (30/03/2011). By invitation; only Australian attendee.
- Attended, Bootes Course on Translational Medicine, John Curtin School of Medical Research, Australian National University (17 – 20/09/2010).
- Poster judge, WSU College of Health & Science Research Future Postgraduate Forum. 2010.
- Poster judge, 1<sup>st</sup> International Congress on Analytical Proteomics/5<sup>th</sup> Congress of the Portuguese Proteomics Network. Lisbon, Portugal. 2009.
- **Guest lecturer / community outreach at area high school, Chevalier College** (06/2009); two lectures ('Molecules-to-Cells' and 'Imaging')
- Established, managed, and further developed a state-of-the-art Molecular Physiology Lab and a Cellular Analysis Facility that together provided Cell Biological, Proteomics and Lipidomics capacity to the WSU School of Medicine, the WSU Molecular Medicine Research Group, and a range of other collaborators; supported a large number of publications and successful grants.
- **Moderator, 2<sup>nd</sup> NeuroConnections Research Retreat, Hotchkiss Brain Institute**, 01/07
- Poster judge for Minority Students Program, ASCB 2006
- **My 4<sup>th</sup> Year BHSc Project Student, Ms. Maggie Lee, was awarded a \$2,000 research grant from the U. Calgary Undergraduate Student Research Program (USRP)** in Health & Wellness; these local health research grant competitions are incredibly stiff.
- **My team of International Baccalaureate students awarded first prize** in the 1<sup>st</sup> annual Canadian Proteome Society Student Awards for Enabling Technologies in Proteomics (09/05)
- **Lab tour/discussion: use of protein & lipid analyses to understand brain function & disease states; for students from U of C Centre for Gifted Education** (07/21/04)
- **Supporting speaker for the Faculty of Medicine B.H.Sc. program**; introduced the program in support of future recruitment interactions with Medicine Hat College (09/09/03)
- Established state-of-the-art Proteomics and Lipidomics Facilities that were housed in my laboratory; this resulted in collaborations with several key research programs in the Faculty, the University of Calgary, and other Institutions (Drs. Power, Zamponi, Wiebe, Teskey, Dyck, Hill, Childs, Wildering, Zochodne, Cross, Beattie, Rancourt, Storey, Yong, Nguyen, and Weselake), including generating preliminary data for subsequently successful grants to many of these groups.
- Arranged for visiting speakers at Brock University:
  - Prof. O. Jahn (Max Planck Institute for Experimental Medicine, Göttingen, Germany)
  - Dr. A. Yergey (NICHD, NIH, USA)
  - Prof. R. Zorec (U. Ljubljana, Slovenia); VP Slovenian Academy of Sciences
  - Dr. R. Seltner (Mount Royal U., Calgary) - to establish formal institutional relations
  - Dr. O. Söderberg (U. Uppsala, Sweden)
  - Dr. P. Abbineni (U. Michigan)
  - Dr. S. Cologna (U. Chicago)
  - Dr. P. Shortland (Western Sydney U.)
- Arranged for visiting speakers in WSU School of Medicine:
  - Prof. P. Robinson (CMRI, Sydney)
  - Dr. T. Rogasevskaia (U. Calgary, Canada)
  - Prof. J. Keast (Kolling Institute, Sydney)
  - Dr. K. Furber (U. Calgary, Canada)
  - Prof. M. Wilkins (UNSW, Sydney)
  - Prof. A. Yergey (NICHD, NIH, USA); 2010, 2011, 2013
  - Prof. N. Packer (Biomolecular Frontiers Centre, Macquarie U.)
  - Prof. R. Zorec (U. Ljubljana, Slovenia) - est. formal institutional relations
  - Prof. R. French (U. Calgary, Canada)
  - Prof. J. Rettig (Physiology, Saarland University, Germany)
  - Prof. D. Stock (Victor Chang Cardiac Research Institute, U. Sydney)
  - Prof. B. McCarry (McMaster University, Canada) - est. formal institutional relations
  - Prof. G. Zamponi (U. Calgary, Canada)
  - Prof. J. Stow (U. Queensland)

- Dr. S.T. Cooper (Children's Hospital Westmead, and U. Sydney)
  - A/Prof. A.J. Hannan (Florey Institute of Neuroscience and Mental Health)
  - Prof. K. Gaus (UNSW)
  - Prof. H. Herzog (Garvan Institute of Medical Research)
  - A/Prof. M.A. Colicos (U. Calgary & Riken Institute, Tokyo)
  - Prof. M. Lindau (Cornell U. & Max Planck Institute for Biophysical Chemistry)
  - Prof. C. Barg (Uppsala U., Sweden)
  - Dr. U. Becherer (Physiology, Saarland University, Germany)
- Arranged for visiting speakers in both the U of C Dept. of Physiology & Biophysics, and the Dept. Biochemistry & Molecular Biology:
- Dr. R. Rahamimoff (University of Jerusalem, Israel); AHFMR Visiting Lecturer
  - Dr. T.F.J. Martin (U. Wisconsin-Madison); AHFMR Visiting Lecturer
  - Dr. A. Mayer (Friedrich-Miescher-Labor, Germany); AHFMR Visiting Lecturer
  - Dr. A. Yergey (NICHD, NIH, USA)
  - Dr. P.S. Blank (NICHD, NIH, USA)
  - Dr. S.S. Vogel (NIAAA, NIH, USA); AHFMR Visiting Lecturer
  - Dr. P. Dietl (University of Innsbruck, Austria); AHFMR Visiting Lecturer
  - Dr. B. Innocenti (University of Tübingen, Germany)
  - Dr. W. Singer (University of Innsbruck, Austria)
  - Dr. G. Rapp (EMBL; Rapp Optoelectronics)
  - Dr. G. Eitzen (University of Edmonton, Alberta)
  - Dr. E. Stanley (University Hospitals Research Network/U of T); AHFMR Visiting Lecturer
  - Dr. M. Lindau (Cornell University, USA); AHFMR Visiting Lecturer
  - Dr. R. Chow (Keck Medical School, USC, USA); AHFMR Visiting Lecturer
  - Dr. M. Morency (Foley & Lardner, Boston, USA)
  - Dr. J. Rizo-Rey (Texas Southwestern Medical Center); AHFMR Visiting Lecturer
  - Dr. M.A. Cortez (Division of Neurology, University of Toronto)
  - Dr. M. Charlton (University of Toronto); AHFMR Visiting Lecturer
  - Dr. J. Klingauf (Max Planck Institute, Germany); AHFMR Visiting Lecturer
  - Dr. A. Hennessy (U. Western Sydney, Australia) - est. formal institutional relations
  - Dr. P. Robinson (CMRI, Australia); AHFMR Visiting Lecturer
  - Dr. C. Borchers (UBC Proteomics Facility)
  - Dr. A. Yergey (NICHD, NIH, USA); AHFMR Visiting Lecturer
  - Dr. J. Rettig (Physiology, Saarland University, Germany); AHFMR Visiting Lecturer

## VIII. RESEARCH SUPPORT

### CURRENT

**Discovery Grant**, Natural Sciences and Engineering Research Council of Canada  
Proteomes of proteoforms — improving routine top-down proteomic analyses.

**\$210,000 CND (2019-2024); PI**

**Brock University**, Explore Grant

Early biomarkers predictive of preterm birth + technology development to refine proteomics

**\$10,000 CAD (2019-2020); PI**

**Brock University**, Lab start-up

Top-down Proteomics Lab

**\$150,000 CAD (2016-present); PI**

**ARC**, Linkage Project (LP160100909)

Smart management of disinfectant in chloraminated water-supply systems.

**\$2,191,000AUS** (\$710,000 ARC + \$951,000 industry + \$530,000 CSIRO & WSU;

**2016-2021); CID** (CIA – A/Prof Arumugam Sathasivan (UWS) and a team including members from universities across Australia + Arizona State University)

**Rotary Club of Narellan**, Biomedical Research Grant

The Multiple Sclerosis (MS) Project

**\$440,000 AUS (2013-2020); co-CI** (with Dr Myers, MMRG); Dr. Mahns co-CI since 2017.

PREVIOUS

**Anonymous Private Foundation, Biomedical Research Grant**

Phosphoproteins critical to Ca<sup>2+</sup>-triggered exocytosis  
**\$391,820 AUS (2011-2019); CIA**

**International Retinal Research Foundation**

Defining the molecular pathways regulated by endogenous BEST1 in Best disease  
**\$200,000 US/\$271,075 AUS (2015-2018); CIE (CIA – Dr. M. O'Connor, WSU MMRG)**

**NHMRC, Project Grant (APP1065328)**

Membrane attachment and components of the Ca<sup>2+</sup>-triggered release mechanism  
**\$373,447 AUS (2014-2017); CIA**

**WSU – RPA Partnership Funding**

Identification of post-translational modifications of serum proteins predictive for preterm birth  
**\$15,000 AUS (2015-2016); CIA (CIB – Prof J. Hyett, RPA/USyd)**

**Research Foundation of the Cerebral Palsy Alliance (PG4115)**

Post-Translational modifications of serum proteins that are predictive for preterm labour  
**\$15,000 AUS (2015-2017); CI (with A. D'Silva (PhD student) & Prof J. Hyett)**

**WSU Molecular Medicine Research Group, Facility Funding**

Renewal and Upgrading of Top-Down Proteomics Infrastructure  
**\$37,000 AUS (2015-2017); CIA - with equal joint funding from the WSU SoM**

**German Academic Exchange Service (DAAD), Universities Australia – Germany Joint Research Co-operation Scheme**

Critical roles of lipids in the late steps of Ca<sup>2+</sup>-triggered exocytosis  
**\$50,000 AUS (2016-2017); with Prof M. Lindau, Max Planck Institute for Biophysical Chemistry (Göttingen, Germany)**

**Rebecca L Cooper Research Foundation, Equipment Grant**

The Multi-Spectral Proteome Imager: driving the next generation of Top-down Proteomic analyses  
**\$21,600 AUS (2015); co-CI with M. Withers (Graduate student)**

**Miltenyi Biotech, Scholarship Grant**

Miltenyi-WSU Molecular Medicine Research Group PhD Scholarship  
**\$75,000 AUS (2013-2015); MMRG (student co-supervised by Dr Myers & Prof Coorssen)**

**Rebecca L Cooper Research Foundation, Equipment Grant**

Real-time analyses of single cell functions using electrophysiology: dissecting the central mechanism of neuronal function  
**\$20,800 AUS (2013); CIA**

**Australian Academy of Science, COST Action Program grant-in-aid**

Travel to speak at a European proteomics meeting and visit collaborators  
**\$4,300 AUS (2013); CIA**

**Western Sydney University, Research Grant**

*Tritrichomonas foetus*-Epithelial Interactions: Dissecting the Roles of Attachment, Cytotoxicity and Host Factors in the Infection Process  
**\$23,050 AUS (2013); CIB (CIA-Dr. C. Stack, SSH)**

**Western Sydney University, Research Grant**

*Using protein analyses and ESI-MS to identify molecular mechanisms of metal complexes*  
**\$5,000 AUS (2013); CIB (CIA – Prof J. Aldrich-Wright, SSH)**

**Bellberry / WSU School of Medicine, Targeted Research Funding**

Using cholesterol to dissect the mechanism of fast Ca<sup>2+</sup>-triggered membrane fusion  
**\$120,000 AUS (2012-2013); CI**

**Western Sydney University, Research Grant**

Quantitative analysis of differential gel electrophoresis (DIGE)-based protein detection  
**\$7,500 AUS (2012-2013); co-CI with Dr. E. Wright (postdoctoral fellow)**

**ARC, LIEF Grant**

Fluorescence microscopy with optical tweezers: imaging cellular responses.  
**\$300,000 AUS (2011)**; one of 17 CI's from institutions in the Sydney area.

**WSU College of Health & Science, Targeted Research Funding**

Cholesterol is a critical component in the mechanism of Ca<sup>2+</sup>-triggered membrane fusion.  
**\$55,000 AUS (2011); CI**

**WSU School of Medicine, Special Infrastructure Funding**

Automated freeze substitution workstation for electron microscopy.  
**\$71,666 AUS (2011); CIA** with Prof Lee and A/Prof Killingsworth).

**WSU College of Health & Science, Research Infrastructure Funding**

Establishment of a zebrafish and microinjection facility.  
**\$142,000 AUS (2011); CIB** (CIA – Dr. M. O'Connor, with cross-College team (Profs. Lee, Aldrich-Wright, Price, Chan, A/Profs Chang & Ross, and Dr. Myers).

**Slovenian Government, Promotion of Science and International Agreements**

Cell sciences & advanced cell therapies; meetings to foster Australian-EU scientific collaborations.  
**1,700 EURO (2011); co-CI** with Prof R. Zorec (U. Ljubljana)

**ARC, LIEF Grant**

Beyond Proteomics: structure and function of protein modifications.  
**\$500,000 AUS (2010)**; one of 25 CI's from institutions in the Sydney area.

**Western Sydney University, International Research Initiatives Scheme**

UWS MMRG symposium 'International Efforts in Translational Medicine' (**Satellite Meeting of the International Human Proteome Organization 2010 Annual Meeting**).  
**\$10,800 AUS (2009-2010); CIA** (with the UWS Molecular Medicine Research Group)

**Western Sydney University, Research Infrastructure Funding**

Cell and protein analysis facility (flow-cytometer and electrophoresis set-up).  
**\$250,000 AUS (2009-2010); CIB** (CIA – Dr. S. Myers, with cross-College team (Profs. Lee and Hennessy, A/Ps Aldrich-Wright and Muench, and Dr. Piller).

**Western Sydney University, Research Grant**

Proteomic analysis of human preterm labour: dissecting molecular mechanisms.  
**\$20,000 AUS (2009-2010); CI**

**Operating Grant (renewal), Natural Sciences and Engineering Research Council of Canada**

Roles of native membrane matrix components in regulated secretion.  
**\$211,965 CND (2008-2013); PI** (terminated with move to Australia)

**Operating Grant (renewal), Canadian Institutes of Health Research**

Role of Cholesterol in the mechanism of calcium-triggered membrane fusion  
**\$734,190 CND (2006-2013); PI**

**Operating Grant, Canadian Institutes of Health Research**

Dietary restriction and spinal cord injury  
**\$363,000 CND (2008-2010); collaborator** – PI Dr. W. Tetzlaff & ICORD/UBC team.

**Operating Grant, Nielson Foundation**

Genomic and proteomic analyses of spinal cord injury.  
**\$250,000 USD (2008-2009); co-PI** with Dr. W. Tetzlaff and ICORD / UBC team.

**Team Start-up Grant, Michael Smith Foundation**

Spinal cord injury proteomics  
**\$200,000 CND (2007-2009); collaborator** – PI Dr. W. Tetzlaff & ICORD/UBC team.

**Western Sydney University, Research Infrastructure Funding**

Establishing a mass spectrometry facility for proteomics, lipidomics, and natural products research and teaching.  
**\$470,000 AUS (2008-2009); co-CI** with Prof. Sucher and including a College team (Prof. Price, A/Ps Muench, Shalliker, and Drs. Higgins, Chang, Piller, Myers, Stack, & Moffitt).

**WSU School of Medicine**, Special Infrastructure Funding  
Confocal Microscope, Electrophysiology Rig, Liquid Chromatography System, & Nanodrop Spectrophotometer.

**\$483,500 AUS (2008); co-CI** with Profs. Hennessy, Lee, Morley, A/P Muench, and Drs. Lind, Preis, & Mahns.

**WSU College of Health & Science**, Targeted Research Funding  
Roles of Native Membrane Matrix Components in Regulated Secretion.

**\$30,000 AUS (2008); CI**

**Operating Grant**, Natural Sciences and Engineering Research Council of Canada  
Roles of membrane matrix components in the calcium-triggered steps of exocytosis

**\$160,000 CND (2003-2008); PI**

**Multi-user Equipment & Maintenance Grant**, Canadian Institutes of Health Research  
The Southern Alberta Mass Spectrometry Centre

**\$243,150 CND (2004-2007); co-PI** (PI, Dr. D.C. Schriemer, with co-PI's Drs. Deans, Fujita, Gravel, Power, Rancourt, Surette, and Walsh)

**Operating Grant (renewal)**, Canadian Institutes of Health Research  
The protein machinery of calcium-triggered membrane fusion

**\$299,889 CND (2003-2006); PI**

**Project (short term)**, University Research Grants Committee, University of Calgary  
Direct role of cholesterol in the molecular mechanism of Ca<sup>2+</sup>-triggered membrane fusion

**\$6,000 CND (2005-2006); PI**

**Operating Grant (Pilot)**, Calgary Health Region Perinatal Funding, Child Health Research Office  
Proteomic analysis of human myometrial tissue in term labouring and non-labouring patients

**\$30,000 CND (2005-2006); PI**, with Dr. Wood (co-investigator)

**Operating Grant (Pilot)**, Calgary Health Region Perinatal Funding, Child Health Research Office  
Proteomic analysis of human fetal membranes in term and preterm labour

**\$30,000 CND (2004-2005); PI**, with Dr. S. Wood (co-investigator)

**Operating Grant**, Fonds zur Förderung der Wissenschaftlichen Forschung (Austrian Science Fund)  
Exocytosis in alveolar type II cells of the lung (FWF P12974)

**367,000 Euro (~ \$500,000 CND; 2002-2005);** sole International Collaborator on this project with Prof. Dr. Paul Dietl, U. Innsbruck.

**Operating Grant (Pilot)**, CIHR (Genomic Research)

A functional analysis of long-term memory

**\$120,000 CND + \$45,308 CND in equipment (2002-2004); PI**, with Drs. Lukowiak and Syed (co-investigators)

**Collaborative Research Grant**, University Research Grants Committee, University of Calgary  
The aging brain: a multidisciplinary study of learning deficiencies

**\$9,910 CND (2003); co-PI** (with Dr. Wildering)

**Laboratory Establishment & Implementation Grant**, Alberta Heritage Found. for Medical Res.

Identification of membrane components essential to Ca<sup>2+</sup>-triggered exocytosis

**\$360,000 CND (2000 - 2003); PI**

**Operating Grant**, Canadian Institutes of Health Research

Identifying the minimal molecular components of regulated membrane fusion

**\$245,112 CND (2000-2003); PI**

**Facility Establishment Grant**, Provincial Res. Excellence Envelop / U of C Faculty of Medicine  
Establishing an automated, quantitative HPTLC facility

**\$105,000 CND (2001); PI**

**Faculty of Medicine Endowment Grant**, Lam Foundation and the Rose Scotter Memorial Fund  
Establishing a quantitative high performance thin layer chromatography facility  
**\$10,000 CND (2001); PI** (equipment support for a dedicated N<sub>2</sub> generator).

**New Opportunities/Initiatives Facility Establishment Grants**, Canada Foundation for Innovation (CFI), Alberta Network for Proteomics Innovation (ANPI), Alberta Cancer Board, and University of Calgary Faculty of Medicine

Functional proteomics of synaptic vesicle trafficking: An integrated approach to CNS plasticity and functional recovery; establishing a functional proteomics facility.

**\$671,365 CND (2001); Lead PI**, with Drs. Whelan and Lu (co-PI's)

- I directly administered \$329,295 for establishment of a proteomics facility and \$150,860 for equipment related to functional assays.

**Faculty of Medicine Endowment/Start-up Grant**, Ruth Rannie Memorial Fund

The essential proteins of regulated membrane fusion

**\$4,000 CND (2000-2001); PI**

**Starter Grant**, University Research Grants Committee, University of Calgary

A molecular analysis of learning and memory

**\$9,832.74 CND (2001); PI**

**Young Innovator Award** (Vice-President, Research), Hunter Exploration and Petro Canada.

Toward an integrated molecular understanding of learning and memory

**\$20,000 CND (2001); PI**

## IX. INVITED ADDRESSES

1. What is the Aetiology of Multiple Sclerosis? Systematic Studies to Define a Better Mouse Model. Plenary address: **6th International Congress on Analytical Proteomics**. Lisbon, Portugal. July 2019.
2. First & foremost – It's about the Biology. Quantitative high-resolution top-down proteomics to analyse molecular mechanisms and identify biomarkers. Medical Faculty, University of Ljubljana and the **Slovenian Academy of Sciences**, Slovenia. November 2017.
3. Identification of post-translational modifications of first trimester maternal serum proteins that are predictive for spontaneous preterm birth. Keynote address: **5th International Congress on Analytical Proteomics**. Lisbon, Portugal. July 2017.
4. Community-driven research: Translating an Australian experience into new directions at Brock University. St. Catharines Central Rotary Club. St. Catharines, ON. April 2017.
5. The global graduate student (with Dr. S. Henderson). Managing Cultural Diversity Meeting: "Leadership on the Move." Hochschule Koblenz, University of Applied Sciences, Remagen, Germany. November 2016.
6. Alternate approaches to identify components critical to fast, Ca<sup>2+</sup>-triggered exocytosis. **Max Planck Institute for Biophysical Chemistry**, Göttingen, Germany. May 2016.
7. High resolution quantitative top-down proteomics: 2DE to assess protein species. **Max Planck Institute for Biophysical Chemistry**, Göttingen, Germany. May 2016.
8. Effects of hypertension on the brain: an initial top-down proteomic analysis. 25<sup>th</sup> HUPO Human Brain Proteome Project Workshop. Nobel Forum, **Karolinska Institute**, Stockholm, Sweden. May 2016.
9. First & foremost – It's about the Biology. Quantitative high resolution top-down proteomics to analyse molecular mechanisms and identify biomarkers. **Illawarra Health and Medical Research Institute**, Wollongong, NSW, Australia. March 2016.
10. Multiple Sclerosis: What do we really know? How to proceed? Rotary Club of Macarthur Sunrise. Campbelltown, NSW, Australia. March 2016.
11. High resolution quantitative top-down proteomics: Resolving protein species as a key to identifying biomarkers and analyzing molecular mechanisms. ICORD / **Blusson Spinal Cord Research Centre** / University of British Columbia, Vancouver, Canada. Nov 2015.

12. Alternate approaches to identify components critical to fast, Ca<sup>2+</sup>-triggered exocytosis. Faculty of Science & Technology, and **Biophysics Centre of Excellence**, Universidad del Pais Vasco/Euskal Herriko Unibertsitatea, Bilbao, Spain. Sept, 2015.
13. High resolution quantitative top-down proteomics: Resolving protein species as a key to identifying biomarkers and analyzing molecular mechanisms. **Closing Plenary address: 4<sup>th</sup> International Congress on Analytical Proteomics**. Lisbon, Portugal. Sept 2015.
14. What is the docked state of a release-ready vesicle? **18<sup>th</sup> ISCCB Meeting / International Symposium on Chromaffin Cell Biology**. Cairns, Australia. August 2015.
15. Quantitative top-down proteomics: A high resolution 2D gel-based approach to analyze biomolecular mechanisms. **Australian Centre for Research on Separation Sciences Annual Meeting. Opening Plenary**. Hobart, CA, AUS. July, 2015.
16. Quantitative top-down proteomics: A high resolution 2D gel-based approach to analyze biomolecular mechanisms. **Bio-Rad Laboratories**, Inc. Hercules, CA, USA. March, 2015.
17. Alternate approaches to identify components critical to fast, Ca<sup>2+</sup>-triggered exocytosis. **Biopolis (Helios)**, Singapore. December, 2014.
18. Introduction to Proteomics: large-scale protein analyses. St. Xavier's College. Mumbai, India. December, 2014.
19. Systems Biology = 'BIG' Biology. St. Xavier's College. Mumbai, India. December, 2014.
20. Quantitative top-down proteomics to dissect cellular and molecular mechanisms. **International Proteomics Conference/6<sup>th</sup> Indian Proteomics Society Meeting**. Mumbai, India. December 2014.
21. Introduction to Two-dimensional gel electrophoresis (2DE). Education Day plenary at the **International Proteomics Conference/6<sup>th</sup> Indian Proteomics Society Meeting**. Mumbai, India. December 2014.
22. Lipid regulation of exocytosis. Annual Meeting of the Australian Physiological Society. U. Queensland, Australia. December 2014.
23. Quantitative top-down proteomics to dissect cellular and molecular mechanisms. The NSW Reproduction Forum. U. Sydney, Australia. November 2014.
24. Driving Top-Down Proteomics to dissect molecular mechanisms. Proteomics Research Group, Life Sciences Centre, U. British Columbia. Vancouver, BC, Canada. Sept 2014.
25. Mechanisms of exocytosis, a new research-intensive medical school, and driving top-down Proteomics. Marianopolis College. Montreal, QC, Canada. Sept 2014.
26. A role for PLD in the late steps of exocytosis. **18<sup>th</sup> International Union of Pure and Applied Biophysics Congress**. Brisbane, Australia. August 2014.
27. Could 2DE help expose missing protein species? **Inaugural HUPO Human Proteome Project "Missing Proteins" Workshop**. Australian School of Advanced Medicine, Macquarie University. Sydney, Australia. July 2014.
28. Quantitative top-down proteomics: A high resolution 2D gel-based approach for the analysis of biomolecular mechanisms. **Centenary Institute / U. Sydney / Royal Prince Alfred Hospital**. Sydney, Australia. June 2014.
29. Quantitative top-down proteomics: A high resolution 2D gel-based approach for the analysis of biomolecular mechanisms. Faculty of Science, Mount Royal University, Calgary, Canada. April 2014.
30. Quantitative top-down proteomics: A high resolution 2D gel-based approach for the analysis of biomolecular mechanisms. **SciLife Biomedical Centre**, Uppsala University, Sweden. March 2014.
31. Vesicular 'Omics: Defining the release-ready and Ca<sup>2+</sup>-triggered fusion steps of regulated exocytosis. **Australasian Proteomics Society 19<sup>th</sup> Lorne Proteomics Symposium**, Lorne, VIC. Feb 2014. Keynote; highlighted in the Australian Life Scientist 11, 14-16.
32. High resolution gel-based proteomics for the analysis of biomolecular mechanisms. Faculty of Health, U. Newcastle, NSW. Nov 2013.
33. Alternate approaches to identify components critical to fast, Ca<sup>2+</sup>-triggered exocytosis. **Florey Institute of Neuroscience and Mental Health**, U. Melbourne, VIC. Oct 2013.

34. Alternate approaches to identify components critical to fast, Ca<sup>2+</sup>-triggered exocytosis. Health Innovations Research Institute, RMIT University, VIC. Oct 2013.
35. Top-down Proteomics: 2D gels are an integral part of dissecting molecular mechanisms. i3 Institute, University of Technology, Sydney, NSW, Australia. September 2013.
36. Top-down Proteomics: 2D gels are an integral part of dissecting molecular mechanisms. Ingham Institute, Liverpool, NSW, Australia. September 2013.
37. Alternate approaches to identify components critical to the late steps of fast, Ca<sup>2+</sup>-triggered exocytosis. Centre for Neuroscience, School of Medicine, Flinders University, SA. Aug 2013.
38. The Omics of Exocytosis: Defining the release-ready and Ca<sup>2+</sup>-triggered fusion steps. **Keynote address: 3<sup>rd</sup> International Congress on Analytical Proteomics.** São Pedro, Brazil. July 2013.
39. Alternate approaches to identify components critical to the late steps of fast, Ca<sup>2+</sup>-triggered exocytosis. Faculty of Medical Sciences, UCA-CONICET, Buenos Aires, Argentina. July 2013.
40. Top-down Proteomics: 2D gels are an integral part of the process. 3<sup>rd</sup> Annual **European Cooperation in Science & Technology FAP Meeting.** Kosice, Slovakia. April 2013.
41. Alternate approaches to identify components critical to the late steps of fast, Ca<sup>2+</sup>-triggered exocytosis. Dept of General Physiology, University of Ulm, Germany. April 2013.
42. Alternate approaches to identify components critical to the late steps of fast, Ca<sup>2+</sup>-triggered exocytosis. Institute of Physiology, Saarland University. Homburg, Germany. April 2013.
43. Systems Biology: What it is, and where it is taking biomedical research and healthcare. Camden Science Cafe, NSW, Australia. March 2013.
44. Understanding human labour: myometrial proteomics. Invited platform presentation at the Biomarker Discovery Conference. Shoal Bay, NSW, Australia. December 2012.
45. Top-down Proteomics: 2D gels are an integral part of the process. NICHD & NIH Mass Spectrometry Interest Group, **NIH**, Bethesda, Maryland, USA. September 2012.
46. Alternate approaches to identify components critical to the late steps of fast, Ca<sup>2+</sup>-triggered exocytosis. **Children's Medical Research Institute** (University of Sydney), Westmead, NSW, Australia. June 2012.
47. Dissecting molecular mechanisms — examples of how and why. McMaster University, Ontario, Canada. May 2012.
48. Fertilisation - Proposed advances to human IVF using lessons learned from studies of sea urchin oocytes. Genea Australia, Sydney. February 2012.
49. High resolution gel-based proteomics for the analysis of molecular mechanisms. McMaster University, Ontario, Canada. December 2011.
50. Learners & Leaders: Hands-on training and long-term career success. Brock University, St. Catharines, Ontario, Canada. September 2011. **Invited Homecoming address.**
51. Pathways to a career in science: A candid alumnus perspective. Faculty of Mathematics and Science. Brock University, St. Catharines, Ontario, Canada. September 2011. **Invited Homecoming address.**
52. High resolution gel-based proteomics for the analysis of molecular mechanisms. **Instituto de Tecnologia Química e Biológica.** Lisbon, Portugal. September 2011.
53. Optimisation of CBB staining for sensitive near IR protein detection. **2<sup>nd</sup> International Congress on Analytical Proteomics.** Ourense, Spain. July 2011.
54. Alternate Approaches to Analysing Lipid Contributions to Docking, Priming, and Fusion. **16<sup>th</sup> ISCCB Meeting / International Symposium on Chromaffin Cell Biology.** Beijing, China. July 2011.
55. Refining gel-based proteomics to optimize analyses. Brock University, St. Catharines, Ontario, Canada. May, 2011.
56. Alternate approaches to identify components critical to the mechanisms of docking, priming, and Ca<sup>2+</sup>-triggered membrane fusion. **Institute of Neuroscience, University of Milan.** Milan, Italy. March 2011.
57. High resolution gel-based proteomics for the analysis of molecular mechanisms. **Institute of Molecular Medicine, Peking University.** Beijing, China. Dec. 2010.

58. High resolution gel-based proteomics for the analysis of molecular mechanisms. **Beijing Institute of Genomics, Chinese Academy of Sciences**. Beijing, China. Dec. 2010.
59. New approaches to dissecting the mechanism of Ca<sup>2+</sup>-Triggered Membrane Fusion. University of Alabama at Birmingham, USA. May 2010.
60. Refining gel-based proteomics to optimize analyses. In the Protein Technologies Symposium at the **3<sup>rd</sup> Annual Protein and Peptide Conference**. Beijing, China. March 2010.
61. Alternate approaches to dissecting the mechanism of fast, Ca<sup>2+</sup>-triggered membrane fusion. In the UWS MMRG Symposium 'New Directions in Membrane Trafficking Research'. **Official Satellite Meeting of the 2010 Joint Annual Meeting of the Australian Physiological and Neuroscience Societies**. Campbelltown, Australia. February, 2010.
62. An unbiased approach to identifying proteins critical to the mechanism of Ca<sup>2+</sup>-triggered membrane fusion. In the session 'Mechanisms underlying exocytosis and endocytosis: potential relevance to health and disease' at the **2010 Joint Annual Meeting of the Australian Physiological and Neuroscience Societies**. Sydney. February, 2010.
63. New approaches to dissecting the mechanism of Ca<sup>2+</sup>-triggered membrane fusion. Brock University, St. Catharines, Ontario, Canada. November, 2009.
64. New approaches to dissecting the mechanism of Ca<sup>2+</sup>-Triggered Membrane Fusion. **Instituto de Biotecnologia/UNAM**. Cuernavaca, Mexico. November 2009.
65. Refining gel-based proteomics: Towards optimized methods of analysis. **Instituto de Biotecnologia/UNAM**. Cuernavaca, Mexico. November 2009.
66. Integrating studies of lipids and proteins to identify components critical for calcium-triggered membrane fusion. **15<sup>th</sup> ISCCB Meeting / International Symposium on Chromaffin Cell Biology**. Merida, Mexico. November 2009.
67. Perspectives and approaches to dissecting the mechanism of Ca<sup>2+</sup>-Triggered Membrane Fusion: What's new at UWS. University of Ljubljana and the Slovenian Physiology and Biochemistry Societies, Slovenia. October 2009.
68. Refining gel-based proteomics: Towards optimized methods of analysis. **1<sup>st</sup> International Congress on Analytical Proteomics/5<sup>th</sup> Congress of the Portuguese Proteomics Network**. Lisbon, Portugal. October 2009.
69. Studying Proteins and Lipids to Dissect the Molecular Mechanisms of Ca<sup>2+</sup>-Triggered Membrane Fusion. **Anderson Stuart research seminars for Anatomy & Physiology, University of Sydney**. Sydney, Australia. Sept. 2009.
70. Dissecting the Molecular Mechanisms of Calcium-Triggered Membrane Fusion: Perspectives & Approaches. **Centre for Vascular Research, University of New South Wales**. Sydney, Australia. Sept. 2009.
71. Using thiol-reactivity as a tool to dissect the mechanism of calcium-triggered membrane fusion. **Institute of Molecular Medicine, Peking University**. Beijing, China. Aug. 2009.
72. Enhancing resolution & detection for gel-based proteomics. **Centenary Institute / Royal Prince Alfred Hospital**. Sydney, Australia. July 2009.
73. Enhancing resolution & detection for gel-based proteomics. **Blusson Spinal Cord Centre / University of British Columbia**. Vancouver. April 2009.
74. Thiol-Reactivity: A tool to dissect the mechanism of calcium-triggered membrane fusion. **Annual Hunter Cellular Biology Meeting**. Hunter Valley, Australia. March 2009.
75. Dissecting the Molecular Mechanisms of Calcium-Triggered Membrane Fusion: Perspectives & Approaches. **International Collaboration On Repair Discoveries (ICORD) / University of British Columbia**. Vancouver. Feb. 2009.
76. Integrating studies of proteins and lipids: dissecting the mechanism of Ca<sup>2+</sup>-triggered membrane fusion. Invited symposium talk at the **Annual Meeting of the Australian Physiology Society**. Melbourne, Australia. December 2008.
77. Dissecting the molecular mechanisms of Ca<sup>2+</sup>-triggered membrane fusion: perspectives and approaches. **Kolling Institute of Medical Research, University of Sydney at Royal North Shore Hospital**. Sydney, Australia. August 2008.

78. Studying proteins and lipids to identify critical components of native Ca<sup>2+</sup>-triggered membrane fusion. **3<sup>rd</sup> International Conference of Neurons and Brain Disease**. Seoul, S. Korea. August 2008.
79. Enhancing resolution and detection for gel-based proteomics. **Yonsei Proteome Research Center & Biomedical Proteome Research Center**. Seoul, Korea. August 2008.
80. Dissecting the molecular mechanisms of Ca<sup>2+</sup>-triggered membrane fusion: perspectives and approaches. Faculty of Medical Sciences, University of Bergen, Norway. May 2008.
81. Proteins and lipids: integrative studies of membrane fusion. Faculty of Medical Sciences, University of Ulm, Germany. May 2008.
82. Identifying critical components of native Ca<sup>2+</sup>-triggered membrane fusion: integrative studies of proteins and lipids. **International EU Meeting on Mechanism(s) of Exocytosis**. Ljubljana, Slovenia. May 2008.
83. Enhancing resolution and detection for gel-based proteomics. **Medical University of Vienna**, Vienna, Austria. May 2008.
84. Enhancing the native Ca<sup>2+</sup> sensitivity and kinetics of triggered membrane fusion. **London Research Institute**, London, UK. May 2008.
85. Enhancing resolution and detection for gel-based proteomics. **8th Annual International Conference of the Canadian Proteomics Initiative (CPI 2008)**. Vancouver. May 2008.
86. 'Omics – beyond biomarkers? Medical Genetics Grand Rounds, Alberta Children's Hospital / Calgary Health Region. Calgary, Canada. Feb. 2008.
87. Specific lipids contribute critical negative curvature — an essential component of Ca<sup>2+</sup>-triggered native membrane fusion. Biochemistry and Biomedical Sciences Seminar Series, McMaster University. Hamilton, Canada. Nov. 2007.
88. Application of proteomic approaches to the analysis of spinal cord injury. **Annual meeting, International Collaboration On Repair Discoveries (ICORD)**. Vancouver. Oct. 2007.
89. Specific lipids provide critical negative curvature to enable Ca<sup>2+</sup>-triggered native membrane fusion. **Society of General Physiologists 61<sup>st</sup> Annual Meeting & Symposium: Membrane Biophysics of Fusion, Fission, and Rafts in Health & Disease**. MBL Woods Hole, USA. Sept. 2007.
90. Identifying an essential component of calcium-triggered vesicular fusion: Specific lipids contribute a critical negative curvature. **IBRO 2007 Satellite Meeting: The Secretory Vesicle Cycle and Novel Approaches to its Analysis**. Brisbane, Australia. July 2007.
91. Cholesterol, curvature, rafts, and regulated membrane fusion: getting at mechanism. **Membrane Biology Group, University of Edinburgh**. May 2007.
92. Refining 2D-PAGE for the dissection of molecular mechanisms. **Helmholtz Centre (Proteomics)**, Leipzig, Germany. May 2007.
93. Cholesterol, curvature, rafts, and regulated membrane fusion: getting at mechanism. **London Research Institute**, London, UK. May 2007.
94. Refining 2D-PAGE for the dissection of molecular mechanisms. **Dundalk Research Institute**, Dundalk, Ireland. May 2007.
95. Cholesterol, curvature, rafts, and regulated membrane fusion: getting at mechanism. Lab of Physical & Structural Biology, **NICHD, National Institutes of Health**, USA. April 2007.
96. Cholesterol, curvature, rafts, and regulated membrane fusion: getting at mechanism. Dalhousie University, Halifax. April 2007.
97. Refining 2D-PAGE for the dissection of molecular mechanisms. Proteomics research interest group, Dalhousie University, Halifax. April 2007.
98. Cholesterol, curvature, rafts, and regulated membrane fusion: getting at mechanism. **OHRI Moses and Rose Loeb Research Centre / University of Ottawa Faculty of Medicine**, Ottawa. April 2007.
99. Dissecting the mechanism and modulation of regulated membrane fusion. **Children's Medical Research Institute**, University of Sydney, Australia. February 2007.
100. Assessing Dogma: Recent Advances in 2D-PAGE. Invited platform presentation at the **12<sup>th</sup> Lorne Proteomics Symposium** (annual meeting of the Australasian Proteomics Society), Lorne, Victoria, Australia. February, 2007.

101. Refining 2D-PAGE with the goal of dissecting molecular mechanisms. **Australian National Proteomics Facility** (APAF) and Macquarie University, Sydney, Australia. October, 2006.
102. Advances in understanding the mechanism of calcium-triggered exocytosis. Canadian Centre for Behavioural Neuroscience, University of Lethbridge. September, 2006.
103. The role of cholesterol in synaptic release. **1<sup>st</sup> International Conference on Synapse, Memory, Addiction, and Pain**. Toronto, Ontario. August 2006.
104. Using proteomics to understand epilepsy. **41<sup>st</sup> Meeting of the Canadian Congress of Neurological Sciences**. Montreal, Quebec. June 2006. **One of 6 speakers invited by the Canadian League Against Epilepsy**.
105. What cholesterol and rafts teach us about regulated membrane fusion. Dept. of Physiology, University of Alberta. May 2006.
106. Cholesterol, rafts, and calcium-triggered membrane fusion. **CNRS Centre for Neurochemistry**, Strasbourg, France. May 2006.
107. Dissecting molecular mechanisms: from basic to translational research. Faculties of Medicine and Natural Sciences, University of Ulm, Germany. April, 2006.
108. Cholesterol, rafts, and the mechanism of regulated membrane fusion. **1<sup>st</sup> Lipid and Membrane Biology Symposium**. Graz, Austria. March, 2006. **One of 12 invited international speakers for this inaugural symposium**.
109. Cholesterol, rafts, and the mechanism of regulated membrane fusion. **San Raffaele Scientific Institute**. Milan, Italy. March 2006.
110. Rafts and other roles of cholesterol in regulated membrane fusion. Faculty of Medical Sciences, University of Ulm, Germany. December, 2005.
111. A direct role of cholesterol in regulated membrane fusion. Dept. of Biopolymer Chemistry, University of Munich, Germany. November, 2005.
112. Rafts and other roles of cholesterol in Ca<sup>2+</sup>-triggered vesicular release. Dept. of Physiology, University of Innsbruck, Austria. November, 2005.
113. Roles of cholesterol in membrane fusion. Faculty of Natural Sciences, Universidad del Pais Vasco/Euskal Herriko Unibertsitatea, Bilbao, Spain. November, 2005.
114. High resolution analyses of membrane proteomes using 2D-PAGE. Proteomics Interest Group, **National Institutes of Health**, USA. November, 2005.
115. 2D-PAGE protocols for high resolution analyses of membrane proteomes. University of British Columbia, Faculty of Life Sciences. September, 2005.
116. Quantitative analyses of membrane proteomes. Invited platform presentation at the Perkin Elmer symposium 'Proteomics & Alternative Biomarkers.' Toronto, September, 2005.
117. Regulated Exocytosis: Models and Mechanisms. **Canadian Centre for Behavioural Neuroscience**, University of Lethbridge. September, 2005.
118. Using urchin cortical vesicles to understand regulated membrane fusion. Invited platform presentation at the **Gordon Research Conference** "Fertilization & Activation of Development." July, 2005.
119. Roles of cholesterol in regulated membrane fusion. Toronto Western Hospital Research Institute. May, 2005.
120. Alternate approaches to molecular mechanisms: Dissecting the Ca<sup>2+</sup>-triggered steps of regulated release. Behavioural Neuroscience Research Group, Dept. Psychology, University of Calgary. March, 2005.
121. Cholesterol is an essential component of the minimal native fusion machine. Invited platform presentation at the 49<sup>th</sup> **Biophysical Society Annual Meeting**. Long Beach, CA, USA. Feb., 2005.
122. New directions in dissecting the Ca<sup>2+</sup>-triggered membrane fusion steps of regulated exocytosis. University of California, Riverside. Feb., 2005.
123. 2D-PAGE Protocols for high resolution analyses of membrane proteomes. Invited platform presentation at the **Proteome Society Meeting** "New Technologies, Novel Approaches in Proteomics Research." Dec. 2004.

124. Dissecting the calcium triggered membrane fusion steps of regulated exocytosis: New perspectives on mechanism. **Robarts Research Institute**, University of Western Ontario, Ontario, Canada. Dec., 2004.
125. Coupled functional and molecular analyses of regulated membrane fusion: From dogma to mechanism. Dept. Biological Sciences, Brock University, Ontario, Canada. May, 2004.
126. Introduction to proteomics. Invited address at the Pfizer Canada/Canadian Medical Hall of Fame Discovery Days in Health Sciences. University of Calgary. May, 2004.
127. A functional-molecular analytical approach to regulated exocytosis: From Dogma to Mechanism. 1<sup>st</sup> Annual CMNRG Retreat, Banff, Alberta, Canada. Oct. 2003.
128. Integrated functional-molecular analyses to explore models of regulated membrane fusion. Invited platform presentation at the **HUPO 2nd Annual & IUBMB XIX Joint World Congress**, Montreal, Quebec, Canada. Oct. 2003.
129. Quantitatively assessing models of regulated membrane fusion. Dept. of Biochemistry and Molecular Physiology, **University of Massachusetts Medical Centre**, Worcester, Mass., USA. Sept., 2003.
130. Testing models of regulated exocytosis. **XV Developmental Biology Meeting**, Marine Biological Laboratory, Woods Hole, Mass., USA. Sept., 2003.
131. Integrated Analyses of Regulated Release: Separating Mechanism from Dogma. Faculty of Sciences (Neurosciences Program), University of Lethbridge, Alberta, Canada. June, 2003.
132. Lessons in the establishment of a successful lab and research program. Invited address at the symposium "Sidestepping the Hazards of an Academic Career," hosted by the Cancer Biology Research Group, Faculty of Medicine, University of Calgary. May, 2003.
133. Proteomics: The next wave. Invited address at the Pfizer Canada/Canadian Medical Hall of Fame Discovery Days in Health Sciences. University of Calgary. May, 2003.
134. Getting started: directions in the successful start-up of junior faculty. Invited address to the University of Calgary Senate. March 2003.
135. Dogma and Mechanism: Coupling quantitative functional and molecular analyses to dissect fundamental molecular mechanisms. **Dahlem Colloquium** at the Max Planck Institute for Molecular Genetics, Berlin, Germany. December, 2002.
136. A functional proteomic approach to the dissection of fundamental molecular mechanisms. Faculty of Medicine, McGill University, Montreal, Quebec, Canada. June, 2002.
137. A functional proteomic approach to the dissection of fundamental molecular mechanisms. Centre de Recherche du CHUL and the Eastern Quebec Centre for Proteomics, University of Laval, Quebec City, Quebec, Canada. June, 2002.
138. The role of SNARE proteins in exocytosis. Faculty of Medicine, University of Calgary, as part of the seminar program in the Department of Biochemistry & Molecular Biology. May, 2002.
139. Role of the **Canadian National Site Licensing Project** in research innovation. CNSLP evaluation committee, Winnipeg, Manitoba, Canada. May, 2002.
140. Using functional proteomics to dissect essential molecular mechanisms. Invited platform presentation at the annual meeting of the **Canadian Proteomics Initiative**, Edmonton, Alberta, Canada. May, 2002.
141. Introduction to proteomics: Potential clinical applications. 13<sup>th</sup> annual Clara Christie Research Day (Dept. Obstetrics & Gynaecology), University of Calgary, Alberta, Canada. May, 2002.
142. Applied functional proteomics in the dissection of the calcium-triggered fusion steps of exocytosis. Medical Faculty, and the **State MicroArray Facility**, University of Western Australia. Perth, Western Australia. March, 2002.
143. Establishing an academic facility for quantitative functional proteomics. Murdoch University and the **Australian National Agricultural Research Centre**. Melville, Western Australia. March, 2002.
144. The role of SNARE proteins in exocytosis. Invited platform presentation at the annual meeting of the **Canadian Physiological Society** in a symposium honouring Dr. Harold Atwood. Vernon, BC, Canada. February, 2002.

145. The Canadian National Site Licensing Project: a central role in research innovation. **Launch ceremonies for the CNSLP**, Halifax, Nova Scotia, Canada. September, 2001.
146. Studying the Ca<sup>2+</sup>-triggered fusion steps of exocytosis: problems and strategies. Department of Pharmacology, University of Graz, Graz, Austria. November, 2000.
147. Identifying proteins essential to Ca<sup>2+</sup>-triggered exocytosis. Department of Anatomy, Histology and Cell Biology, University of Innsbruck, Innsbruck, Austria. November, 2000.
148. Studying the Ca<sup>2+</sup>-triggered fusion steps of exocytosis: problems and strategies. **Max Planck Institute for Molecular Genetics**, Berlin, Germany. November, 2000.
149. Identifying proteins essential to Ca<sup>2+</sup>-triggered exocytosis. Department of Biochemistry, University of Alberta, Edmonton, Alberta, Canada. August, 2000.
150. Identifying proteins essential to Ca<sup>2+</sup>-triggered membrane fusion. Invited platform presentation at the **Steenbock Symposium** on Intracellular Protein & Lipid Traffic, University of Wisconsin, Madison, WI, USA. August, 2000.
151. Identifying proteins essential to Ca<sup>2+</sup>-triggered exocytosis. Invited platform presentation at the **FASEB Meeting** entitled Molecular Biophysics of Cellular Membranes, Saxtons River, VT, USA. July, 2000.
152. Identifying proteins essential to Ca<sup>2+</sup>-triggered membrane fusion. Laboratory of Experimental and Computational Biology, **National Cancer Institute/FCRDC**, Frederick, MD, USA. June, 2000.
153. Studying the Ca<sup>2+</sup>-triggered fusion steps of exocytosis: problems and strategies. Laboratory of Cell Biology, **National Heart, Lung, and Blood Institute, NIH**, Bethesda, MD, USA. March, 2000.
154. Identifying factors essential to the late steps of Ca<sup>2+</sup>-triggered exocytosis. Department of Biological Sciences, Simon Fraser University, Burnaby, BC, Canada. April, 1999.
155. Studying the late steps of Ca<sup>2+</sup>-triggered exocytosis: evaluating the role of SNARE proteins. **Research Institute, Sick Children's Hospital/University of Toronto**, Toronto, Ontario, Canada. April, 1999.
156. Evaluating the role of SNARE proteins in the late steps of exocytosis. Department of Physiology, University of Montreal, Montreal, Quebec, Canada. April, 1999.
157. Studying the late steps of Ca<sup>2+</sup>-triggered exocytosis. Neuroscience Research Institute, University of Ottawa, Ottawa, Ontario, Canada. April, 1999.
158. Combined biochemical and functional studies of Ca<sup>2+</sup>-triggered exocytosis. MARCS Program, Pontifical Catholic University of Puerto Rico, Ponce, Puerto Rico. April, 1999.
159. Studying the late steps of Ca<sup>2+</sup>-triggered exocytosis. Neuroscience Research Group, University of Calgary, Calgary, Alberta, Canada. March, 1999.
160. Studying the late steps of Ca<sup>2+</sup>-triggered exocytosis. Protein Trafficking Interest Group, **NICHD, NIH**, Bethesda, MD, USA. February, 1999.
161. Studying the late steps of Ca<sup>2+</sup>-triggered exocytosis. **Center for Molecular Neurobiology Hamburg (ZMNH)**, Hamburg, Germany. February, 1999.
162. A reduced system to study the molecular mechanism of Ca<sup>2+</sup>-triggered exocytosis. **Max-Planck-Institute for Biophysical Chemistry**, Göttingen, Germany. February, 1999.
163. Identifying the essential molecular components of Ca<sup>2+</sup>-triggered exocytosis. **Max-Planck-Institute for Brain Research**, Frankfurt, Germany. February, 1999.
164. Studying the molecular mechanisms of docking and Ca<sup>2+</sup>-triggered fusion. **Max-Planck-Institute for Medical Research**, Heidelberg, Germany. February, 1999.
165. A reduced system to study the final steps of Ca<sup>2+</sup>-triggered exocytosis. Synaptic Mechanisms Section, **National Institutes of Neurological Disease and Stroke**, NIH, Bethesda, MD, USA. July, 1998.
166. A simple model system for exocytosis: Testing hypotheses and identifying essential components. **SARS International Centre for Molecular Marine Biology**, Bergen, Norway. February, 1998.
167. Testing the SNARE hypothesis. Dept. of Cell Biology, University of Alberta, Edmonton, Alberta, Canada. February, 1998.

168. Exocytosis: Testing the SNARE hypothesis. Dept. of Physiology, **Trinity College Dublin**, Dublin, Ireland. December, 1997.
169. Is there a fusion complex or is fusion just complex? Dept. of Biological Sciences, Brock University, St. Catharines, Ont., Canada. September, 1997.
170. Late events in the exocytotic pathway of neuroendocrine cells. Dept. of Neurobiology and Anatomy, Medical College of Pennsylvania and Hahnemann University, Philadelphia, PA, U.S.A. January, 1997.
171. Ca<sup>2+</sup>-triggered exocytosis: Characterization of a conserved mechanism. Dept. of Medical Physiology and Biophysics, University of Sevilla Medical School, Sevilla, Spain. April, 1996.
172. Ca<sup>2+</sup>-triggered exocytosis: Where you'd expect it and where you might not! Dept. of Physiology, University of Edinburgh Medical School, Edinburgh, U.K. April, 1996.
173. The last (milli)seconds in the life of a secretory granule: ATP-independent, Ca<sup>2+</sup>-triggered exocytosis implies post-docking steps in neuroendocrine cells. Max-Planck-Institute / Friedrich Miescher Institute Seminar Group, the **Max-Planck-Institute for Developmental Biology**, Tübingen, Germany. August, 1995.
174. Time-resolved studies of Ca<sup>2+</sup>-triggered exocytosis in neuroendocrine and epithelial cells: The simultaneous use of capacitance measurement, Ca<sup>2+</sup> quantitation by microfluorimetry, and flash photolysis of caged Ca<sup>2+</sup>. Calcium Signalling Group, Dept. of Biological Sciences, Brock University, St. Catharines, Ont., Canada. July, 1995.
175. Bits and pieces of secretion and exocytosis: A 10 year odyssey. Laboratory of Theoretical and Physical Biology, **National Institutes of Health**, Bethesda, MD, U.S.A. June, 1995.
176. Ca<sup>2+</sup>-independent secretion and evidence that PLA<sub>2</sub>, PLC and PLD activities are not essential to the exocytotic mechanism. Dept. of Biochemistry and Molecular Biology, The University of Bergen, Bergen, Norway. May, 1995.
177. Secretion and phospholipase D activity: Correlations and characterization of Ca<sup>2+</sup>-independent pathways in permeabilized human platelets. Dept. of Pharmacology, The Biozentrum, University of Basel, Basel, Switzerland. April, 1994.
178. Ca<sup>2+</sup>-independent secretion and phospholipase D activity in permeabilized human platelets. The Pharmacology Institute, University of Mainz, Mainz, Germany. October, 1993.
179. The apparent role of phospholipase D activity in platelet activation/secretion: Comparisons and contrasts with neutrophils. Granulocyte Research Laboratory, Dept. Hematology, Rigshospitalet, Copenhagen, Denmark. September, 1993.
180. Ca<sup>2+</sup>-independent secretion from permeabilized platelets and the possible involvement of phospholipase D. Dept. of Medical Physics, Gothenburg University, Gothenburg, Sweden. August, 1993.
181. Factors influencing secretion from permeabilized platelets. Dept. of Biological Sciences, Brock University, St. Catharines, Ont., Canada. October, 1992.
182. Factors affecting secretion from human platelets: Ca<sup>2+</sup>-independent actions and evidence that phospholipase D activity may mediate secretion. Vessel Wall Research Group, Health Sciences Centre, McMaster University, Hamilton, Ont., Canada. March, 1992.
183. Second messengers regulating secretion from electropermeabilized human platelets: Evidence that Ca<sup>2+</sup> is not essential to the exocytotic mechanism. Cell Biophysics/Secretory Mechanisms Group, University of Washington, Seattle, WA, U.S.A. December, 1991.
184. Structural effects of neutral lipids on the divalent cation-induced interactions of phosphatidylserine-containing bilayers. Dept. of Biological Sciences, Brock University, St. Catharines, Ont., Canada. December, 1988.
185. Effects of cholesterol on model bilayer membrane systems. Dept. of Biological Sciences, Brock University, St. Catharines, Ont., Canada. May, 1986.

## X. PUBLICATIONS / PRESENTATIONS

i. PEER REVIEWED MANUSCRIPTS (names of my trainees & research associates are underlined)

1. Carbonara, K., and **Coorsen, J.R. (2020)** A 'green' approach to fixing polyacrylamide gels: improvement of top-down proteomic analyses. *Analytical Biochemistry* (Epub 17/7/2020).
2. Coish, J.M., Crozier, R.W.E., Schieffelin, J.S., **Coorsen, J.R.**, Hunter, F.F., and MacNeil, A.J. **(2020)** Zika virus replication in a pre-basophilic/mast cell model is augmented by Dengue virus 6 antibody-dependent enhancement and features selective CCL5 secretion. Submitted (*J. Virology*).
3. Rituper, B., Guček, A., Lisjak, M., Gorska, U., Šakanović, A., Bobnar, S.T., Lasič, E., Božič, M., Abbineni, P.S., Yilmaz, G., Jorgačevski, J., Kreft, M., Verkhatsky, A., Platt, F.M., Anderluh G, Stenovc, M., Božič, B., **Coorsen, J.R.**, Zorec, R. **(2020)** Redistribution of cholesterol from vesicle to plasmalemma controls fusion pore geometry. Submitted (*J. Cell Biol.*).
4. Sen, M.K., Almuslehi, M.S., Shortland, P.J., **Coorsen, J.R.**, and Mahns, D.A. **(2020)** Revisiting the Pathoetiology of Multiple Sclerosis: Has the Tail been Wagging the Mouse? Submitted (*Annals of Neurology*).
5. Shanu, A., Ng, N., Lauto, A., **Coorsen, J.R.**, and Myers, S.J. **(2020)** Calcium mediated calpain activation and microtubule dissociation in Hereditary Sensory Neuropathy Type-1 expressing V144D SPTLC1 mutation. Submitted (*Nature Research*).
6. Almuslehi, M.S., Sen, M.K., Shortland, P.J., Mahns, D.A., and **Coorsen, J.R. (2020)** CD8 T-cell recruitment into the central nervous system of cuprizone-fed mice: Relevance to modelling the aetiology of Multiple Sclerosis. *Frontiers in Cellular Neuroscience* 14:43; doi: 10.3389/fncel.2020.00043. *A top-ranked article during first quarter after publication.*
7. D'Silva, A.M., Hyett, J.A., and **Coorsen, J.R. (2020)** Initial validation testing of four potential early serum biomarkers of preterm labour. *Fetal Diagnosis and Therapy* 47:497-506 (Epub 28/2/2020).
8. Sen, M.K., Almuslehi, M.S., Mahns, D.A., **Coorsen, J.R.**, and Shortland, P.J. **(2020)** Behavioural and histological changes in cuprizone-fed mice. *Brain, Behavior, and Immunity* 87: 508-523 (Epub 31/01/2020).
9. **Coorsen, J.R.**, and Padula, M.P. **(2020)** Top-down Proteomics: In Memory of Dr. Alfred Yergey. *Alfred Linwood Yergey, III, 17 September 1941–27 May 2018. Proteomes* 8: 1; doi:10.3390/proteomes8010001. Editorial for Special Issue.
10. Zhan, X., Li, B., Zhan, X., Schlüter, H., Jungblut, P.R., and **Coorsen, J.R. (2019)** Innovating the concept and practice of two-dimensional gel electrophoresis in the analysis of proteomes at the proteoform level. *Proteomes* 7, 36; doi:10.3390/proteomes7040036. Special Issue: "Top-down Proteomics: In Memory of Dr. Alfred Yergey."
11. Sen, M.K., Almuslehi, M.S., Gyengesi, E., Myers, S.J., Mahns, D.A., Shortland, P.J. and **Coorsen, J.R. (2019)** Suppression of the peripheral immune system limits the central immune response following cuprizone-feeding: Relevance to modelling multiple sclerosis. *Cells*, 8(11), 1314; doi:10.3390/cells8111314 (Special Issue: "Cellular and Molecular Mechanisms in Pathogenesis of Multiple Sclerosis").
12. Furber, K.L., Backlund, P.S., Yergey, A.L., and **Coorsen, J.R. (2019)** Unbiased thiol labeling and Top-Down Proteomic analyses implicate multiple proteins in the late steps of regulated secretion. *Proteomes* 7(4), 34; <https://doi.org/10.3390/proteomes7040034>. Requested for Special Issue: "Top-down Proteomics: In Memory of Dr. Alfred Yergey."
13. Sen, M., Mahns, D., **Coorsen, J.R.**, and Shortland, P. **(2019)** Behavioural phenotypes in the Cuprizone model of central nervous system demyelination. *Neuroscience & Biobehavioral Reviews* 107, 23-46.

14. Dabral, D. and **Coorssen, J.R. (2019)** Arachidonic acid and lysophosphatidylcholine inhibit multiple late steps of regulated exocytosis. *Biochem. Biophys. Res. Comm.* 515, 261-267 (Epub 21/05/2019).
15. Mazinani, S.A., Noaman, N., Pergande, M., Cologna, S.M., Coorssen, J.R.\*, and Yan, H.\* **(2019)** Exposure to microwave irradiation at constant culture temperature slows the growth of *Escherichia coli* DE3 cells, leading to modified proteomic profiles. *RSC Advances* 9, 11810-11817. \*Co-corresponding authors.
16. Kurgan, N., Noaman, N., Pergande, M.R, Cologna, S.M., Coorssen, J.R.\*, and Klentrou, N.\* **(2019)** Changes to the Human Serum Proteome in Response to High Intensity Interval Exercise: A Sequential Top-Down Proteomic Analysis. *Frontiers in Physiology*, April, Vol. 10, paper 362. doi: 10.3389/fphys.2019.00362. \*Co-corresponding authors.
17. Dabral, D. and **Coorssen, J.R. (2019)** Combined Omic and functional assays identify Phospholipases A<sub>2</sub> that regulate docking/priming in Calcium-triggered exocytosis. *Cells* 8, 303; doi:10.3390/cells8040303.
18. Abbineni, P.S. and **Coorssen, J.R. (2018)** Sphingolipids modulate docking, Ca<sup>2+</sup> sensitivity and membrane fusion of native secretory vesicles. *Int. J. Biochem. Cell Biology* 104,43-54 (Epub 5/09/2018).
19. Noaman, N. and **Coorssen, J.R. (2018)** Coomassie does it (better): a Robin Hood approach to total protein quantification. *Analytical Biochemistry* 556, 53-56 (Epub 12/05/2018).
20. D'Silva, A.M., Hyett, J.A., and Coorssen, J.R. (2018) Proteomic analysis of first trimester maternal serum to identify candidate biomarkers potentially predictive of spontaneous preterm birth. *J. Proteomics* 178, 31-42 (Epub 13/02/2018).
21. Noaman, N., Abbineni, P.S., Withers, M., and Coorssen, J.R. (2017) Coomassie staining provides routine (sub)femtomole in-gel detection of intact proteoforms: expanding opportunities for genuine Top-down Proteomics. *Electrophoresis* 38, 3086–3099 (Epub 5/09/2017).
22. Abbineni, P.S. and **Coorssen, J.R. (2017)** Application of High-Throughput Assays to Examine Phospho-Modulation of the Late Steps of Regulated Exocytosis. *High Throughput* 13;6(4). doi: 10.3390/ht6040017 (Epub 13/11/2017).
23. **Coorssen, J.** and Yergey, A. **(2017)** Approaches to Top-Down Proteomics: In Honour of Prof. Patrick H. O'Farrell. *Proteomes* 5, 18. doi:10.3390/proteomes5030018. Editorial for Special Issue.
24. D'Silva, A.M., Hyett, J.A., and Coorssen, J.R. (2017) A routine 'Top-Down' approach to analysis of the human serum proteome. *Proteomes* 5, 13. doi: 10.3390/proteomes5020013. Special Issue: "Approaches to Top-Down Proteomics: In Honour of Prof. Patrick H. O'Farrell"
25. Shanu, A., Stimpson, S.E., Coorssen, J.R.\*, and Myers, S.J.\* **(2017)** Autophagy is up regulated in a neuronal model of Charcot-Marie Tooth Disease that overexpresses Dynamin 2 mutant. *Int. J. Neurosci. Res.* 1:3. ISSN 2572-8385 \*Co-corresponding authors.
26. Ormerod, K.G.\*, LePine, O.K., Abbineni, P.S., Bridgeman, J.M., Coorssen, J.R.\*, Mercier, A.J., and Tattersall, G.J.\* **(2017)** *Drosophila* development, physiology, behaviour, and lifespan are influenced by altered dietary composition. *Fly* 11, 153-170 (Epub 9/03/2017). <http://dx.doi.org/10.1080/19336934.2017.1304331> \*Co-corresponding authors
27. Dabral, D., and **Coorssen, J.R. (2017)** Phospholipase A<sub>2</sub>: Potential roles in native membrane fusion. *Int. J. Biochem. Cell Biol.* 85, 1-5 (Epub 25/01/2017).
28. Stroud, L.J., Slapeta, J., Padula, M.P., Druery, D., Tsiotsioris, G., Coorssen, J.R.\*, and Stack, C.M.\* **(2017)** Comparative proteomic analysis of two pathogenic *Trichomonas foetus* genotypes: there is more to the proteome than meets the eye. *Int. J. Parasitology* 47, 203-213 (Epub 17/01/2017). \*Co-corresponding authors.
29. Stimpson, S.E., Shanu, A., Coorssen, J.R.\*, and Myers, S.J.\* **(2016)** Identifying unique protein alterations caused by SPTLC1 mutations in a transfected neuronal cell model. *World J. Neurosci.* 6, 325-347. DOI: 10.4236/wjns.2016.64035. \*Co-corresponding authors.
30. Epstein, J.A., Blank, P.S., Searle, B.C., Catlin, A.D., Cologna, S.M., Olson, M.T., Backlund, P.S., **Coorssen, J.R.**, and Yergey A.L. **(2016)** ProteinProcessor: A Probabilistic Analysis Using Mass Accuracy and the MS Spectrum. *Proteomics* 16, 2480-2490 (Epub 22/08/2016).

31. Stimpson, S.E., Lauto, A., **Coorssen, J.R.\***, and Myers, S.J.\* **(2016)** Isolation and identification of ER associated proteins with unique expression changes specific to the V144D SPTLC1 mutations in HSN-I. *Biochem. Analytical Biochem.* 5(1), 248. doi:10.4172/2161-1009.1000248. \*Co-corresponding authors.
32. Partridge, M.A., Gopinath, S., Myers, S.J., and **Coorssen, J.R.** **(2016)** An initial top-down proteomic analysis of the standard cuprizone mouse model of Multiple Sclerosis. *J. Chem. Biol.* 9, 9-18 (Epub 14/06/2015).
33. Stimpson, S.E., **Coorssen, J.R.\***, and Myers, S.J.\* **(2015)** Proteome alterations associated with the V144D SPTLC1 mutation that causes Hereditary Sensory Neuropathy-1. *Electronic J. Biol.* 11, 176-186. \*Co-corresponding authors.
34. **Coorssen, J.R.** and Yergey, A.L. **(2015)** Proteomics is analytical chemistry: Fitness-for-purpose in top-down and bottom-up analyses. *Proteomes* 3, 440-453. doi:10.3390/proteomes3040440.
35. Rogasevskaja, T. and **Coorssen, J.R.** **(2015)** The role of phospholipase D in regulated exocytosis. *J. Biol. Chem.* 290, 28683-28696 (Epub 2/10/2015). Highlighted in the *Lipid Chronicles* (1/12/2015).
36. Partridge, M.A., Myers, S.J., Gopinath, S., and **Coorssen, J.R.** **(2015)** Proteomics of a conundrum: thoughts on addressing the aetiology vs. progression of Multiple Sclerosis. *Proteomics Clinical Applications* 9, 838-843 (Epub 11/04/2015). Requested Viewpoint.
37. Mahadeo, M., Furber, K.L., Lam, S., **Coorssen, J.R.\***, and Prenner, E.J.\* **(2015)** Secretory vesicle cholesterol: correlating lipid domain organization and Ca<sup>2+</sup> triggered fusion. *BBA (Biomembranes)* 1848, 1165-1174. \*Co-corresponding authors.
38. Stimpson, S.E., **Coorssen, J.R.\***, and Myers, S.J.\* **(2015)** Optimal isolation of mitochondria for proteomic analyses. *Analytical Biochem.* 475, 1-3 (Epub 14/01/2015). \*Co-corresponding authors.
39. Stimpson, S.E., **Coorssen, J.R.\***, and Myers, S.J.\* **(2015)** Mitochondrial protein alterations in a familial peripheral neuropathy caused by the V144D amino acid mutation in the sphingolipid protein, SPTLC1. *J. Chem. Biol.* 8, 25-35 (Epub 14/11/2014). \*Co-corresponding authors.
40. Wright, E., Padula, M., Higgins, V., Aldrich-Wright, J., and **Coorssen, J.R.** **(2014)** A Systems Biology approach to understanding the mechanisms of action of an alternative anticancer compound in comparison to cisplatin. *Proteomes* 2, 501-526.
41. Oliveira, B.M., **Coorssen, J.R.\***, and Martins-de-Souza, D. **(2014)** 2DE: The Phoenix of Proteomics. *J. Proteomics* 104, 140-150 (Epub 2/04/2014). \*Co-corresponding and co-first author. For a special issue on Environmental and Structural Proteomics.
42. Marshall, L.L., Stimpson, S.E., Hyland, R., **Coorssen, J.R.\***, and Myers, S.J.\* **(2014)** Increased lipid droplet accumulation associated with a peripheral sensory neuropathy. *J. Chem. Biol.* 7, 67-76 (Epub 23/03/2014). \*Co-corresponding authors.
43. Wright, E.P., Partridge, M.A., Padula, M.P., Gauci, V.J., Malladi, C.S., and **Coorssen, J.R.** **(2014)** Top-down proteomics: Enhancing 2D gel-electrophoresis from tissue processing to high sensitivity protein detection. *Proteomics* 14, 872-889 (Epub 3/03/2014); invited submission for a special issue on Tissue Proteomics.
44. Wright, E.P., Prasad, K.A.G., Padula, M.P., and **Coorssen, J.R.** **(2014)** Deep Imaging: How much of the Proteome Does Current Top-Down Technology Already Resolve? *PLoS ONE* 9(1): e86058. doi:10.1371/journal.pone.0086058.
45. Butt, R.H., and **Coorssen, J.R.** **(2013)** Coomassie Blue as a near-IR fluorescent stain: a systematic comparison with Sypro Ruby for in-gel protein detection. *Mol. Cell. Proteomics* 12, 3834-3850 (Epub 16/09/2013).
46. Abbineni, P.S., Hibbert, J.E., and **Coorssen, J.R.** **(2013)** Critical role of cortical vesicles in dissecting regulated exocytosis: Overview of insights into fundamental molecular mechanisms. *The Biological Bulletin* 224, 200-217; invited submission for a special 'symposium issue' on Cell Biology.
47. Rituper, B., Chowdhury, H.H., Jorgacevski, J., **Coorssen, J.R.**, Kreft, M., and Zorec, R. **(2013)** Cholesterol-mediated membrane surface area dynamics in neuroendocrine

- cells. *Biochimica et Biophysica Acta - Molecular and Cell Biology of Lipids* 1831, 1228-1238 (Epub 12/04/2013).
48. Gauci, V.J., Padula, M.P., and **Coorsen, J.R. (2013)** Coomassie blue staining for high sensitivity gel-based proteomics. *Journal of Proteomics* 90, 96-106 (Epub 8/02/2013). Invited submission for a special issue.
  49. Hyland, R.A., Rogers, P.J., Higgins, V.J., Myers, S., and **Coorsen, J.R. (2012)** Measuring hydrogen peroxide reduction using a robust, inexpensive, and sensitive method. *J. Chem. Biol.* 5(4), 143-150 (Epub 1/09/2012).
  50. Raveh, A., Valitsky, M., Shani, L., **Coorsen, J.R.**, Blank, P.S., Zimmerberg, J., and Rahamimoff, R. **(2012)** Observations of Calcium Dynamics in Cortical Secretory Vesicles. *Cell Calcium* 52, 217-225 (Epub 27/07/2012).
  51. **Coorsen, J.R.**, and Zorec, R. **(2012)** Regulated exocytosis per partes. *Cell Calcium* 52, 191-195 (Epub 9/07/2012). Editorial for a special issue on Regulated Exocytosis.
  52. Ormerod, K.G., Rogasevskaia, T., **Coorsen, J.R.\*** and Mercier, A.J.\* **(2012)** Cholesterol-independent effects of methyl-beta-cyclodextrin on chemical synapses. *PLoS ONE* 7(5): e36395. doi:10.1371/journal.pone.0036395. \*Co-corresponding authors. **722 views and 157 downloads within 6 months of publication.**
  53. Rogasevskaia, T., Churchward, M.A., and **Coorsen, J.R. (2012)** Anionic lipids in Ca<sup>2+</sup>-triggered membrane fusion. *Cell Calcium* 52, 259-269 (Epub 17/04/2012) Requested submission for a special issue on Regulated Exocytosis.
  54. Lubieniecka, J.M., Streijger, F., Lee, J.H.T., Stoynov, N., Liu, J., Mottus, R., Pfeifer, T., Kwon, B.K., **Coorsen, J.R.**, Foster, L.J., Grigliatti, T.A., and Tetzlaff, W. **(2011)** Biomarkers for Severity of Spinal Cord Injury in the Cerebrospinal Fluid of Rats. *PLoS ONE* 6(4): e19247 (doi:10.1371/journal.pone.0019247).
  55. Rogasevskaia, T. and **Coorsen, J.R. (2011)** A New Approach to the Molecular Analysis of Docking, Priming, and Regulated Membrane Fusion. *J. Chem. Biol.* 4, 117-136 (Epub 8/02/2011). **Noted as the 5<sup>th</sup> most highly downloaded paper in the journal.**
  56. Gauci, V.J., Wright, E.P., and **Coorsen, J.R. (2011)** Quantitative Proteomics: assessing the spectrum of in-gel protein detection methods. *J. Chem. Biol.* 4, 3-29 (Epub 18/06/10). **Noted as highly accessed.**
  57. Chasserot-Golaz, S., **Coorsen, J.R.\***, Meunier, F., and Vitale, N. **(2010)** Lipid Dynamics in Exocytosis. *Cell Mol. Neurobiol.* 30, 1335-1342 (Epub 16/11/2010). \*Co-corresponding author.
  58. Furber, K.L., Dean, K.T., and **Coorsen, J.R. (2010)** Dissecting the mechanism of Ca<sup>2+</sup>-triggered membrane fusion: probing protein function using thiol-reactivity. *Clin Exp Pharmacol Physiol* 37, 208-217 (Epub 4/08/09). Invited paper after the 2008 Australian Physiological Society Meeting; also published on AuPS website (<http://www.aups.org.au>). Highlighted by MDLinx (Pharmacy), a website for physicians and clinician scientists (<http://www.mdlinx.com/pharmacy/news-article.cfm/3023557>).
  59. Churchward, M.A., and **Coorsen, J.R. (2009)** Cholesterol, regulated exocytosis and the physiological fusion machine. *Biochem. J.* 423, 1-14 (Epub 14/09/09). Invited Review.
  60. Feng, Z.-P., Zhang, Z., van Kesteren, R.E., Straub, V.A., van Nierop, P., Jin, K., Nejatbakhsh, N., Goldberg, J.I., Spencer, G.E., Yeoman, M.S., Wildering, W., **Coorsen, J.R.**, Croll, R.P., Buck, L., Syed, N.I., Smit, A.B. **(2009)** Transcriptome analysis of the central nervous system of the mollusc *Lymnaea stagnalis*. *BMC Genomics* 10, 451 (Epub 19/09/09).
  61. Furber, K.A., Brandman, D., and **Coorsen, J.R. (2009)** Enhancement of the Ca<sup>2+</sup>-triggering steps of native membrane fusion via thiol-reactivity. *J. Chem. Biol.* 2, 27-37 (Epub 3/10/08).
  62. Furber, K.A., Churchward, M.A., Rogasevskaia, T., and **Coorsen, J.R. (2009)** Identifying critical components of native Ca<sup>2+</sup>-triggered membrane fusion: integrating studies of proteins and lipids. From: *Mechanisms of Exocytosis. Annals of the New York Academy of Sciences* 1152, 121-134 (accepted 11/07/08).
  63. Churchward, M.A., Brandman, D., Rogasevskaia, T., and **Coorsen, J.R. (2008)** Copper (II) sulfate charring for high sensitivity on-plate fluorescent detection of lipids and sterols:

- Quantitative analyses of the composition of functional secretory vesicles. *J. Chem. Biol.* 1, 79-87 (Epub 17/06/08).
64. Churchward, M.A., Rogasevskaia, T., Brandman, D.M., Khosravani, H., Nava, P., Atkinson, J.K., and **Coorssen, J.R. (2008)** Specific lipids supply critical intrinsic negative curvature – an essential component of native Ca<sup>2+</sup>-triggered membrane fusion. *Biophys. J.* 94, 3976-3995 (Epub 28/01/08). **[117 citations]**
65. Butt, R.H., Pfeifer, T.A., Delaney, A., Grigliatti, T.A., Tetzlaff, W.G., and **Coorssen, J.R. (2007)** Enabling coupled quantitative genomic and proteomic analyses from rat spinal cord samples. *Mol. Cell. Proteomics* 6, 1574-1588 (Epub 17/05/07).
66. Harris, L., Churchward, M.A., Butt, R.H., and **Coorssen, J.R. (2007)** Assessing detection methods for gel-based proteomic analyses. *J. Proteome Res.* 6, 1418-1425. (Epub 17/03/07). **Designated as Highly Accessed (April-June, 2007). Noted in Proteome Commons News (04/07).**
67. Butt, R.H., Lee, M., Amadi Pershahid, S., Backlund, P., Wood, S., and **Coorssen, J.R. (2006)** An initial proteomic analysis of human preterm labour: placental membranes. *J. Proteome Res.* 5, 3161-3172. (Epub 19/10/06).
68. Taylor, R.C., and **Coorssen, J.R. (2006)** Proteome resolution by 2D gel electrophoresis varies with the commercial source of IPG strips. *J. Proteome Res.* 5, 2919-2927 (Epub 7/10/06). **Designated as Highly Accessed (October-December, 2006). Noted in Swiss Proteomics Society Digest (issue 36).**
69. Rogasevskaia, T. and **Coorssen, J.R. (2006)** Rafts define the efficiency of native Ca<sup>2+</sup>-triggered membrane fusion. *J. Cell Sci.* 119, 2688-2694. (Epub 6/06/06). **Highlighted article in this issue.**
70. Howell, J.M., Winstone, T.L., **Coorssen, J.R.,** and Turner, R.J. (2006) An evaluation of in vitro Protein-Protein Interaction techniques: Assessing contaminating background proteins. *Proteomics* 6, 2050-2069. (Epub 6/03/06).
71. Butt, R.H., and **Coorssen, J.R. (2006)** Pre-extraction Sample Handling by Automated Frozen Disruption Significantly Improves Subsequent Proteomic Analyses. *J. Proteome Res.* 5, 437-448. (Epub 22/12/05).
72. Hibbert, J.E., Butt, R.H., and **Coorssen, J.R. (2006)** Actin is not an essential component in the mechanism of Ca<sup>2+</sup>-triggered vesicle fusion. *Int. J. Biochem. Cell Biol.* 38, 461-471 (Epub 08/11/05).
73. Bi, G.-Q., Bolshakov, V., Bu, G., Cahill, C.M., Chen, Z.-F., Collingridge, G.L., Cooper, R.L., **Coorssen, J.R. et al. (2006)** Recent advances in basic neurosciences and brain disease: from synapses to behaviour. *Molecular Pain*, 2, 38 (December 30). **Designated as Highly Accessed.**
74. Churchward, M.A., Rogasevskaia, T., Höfgen, J., Bau, J., and **Coorssen, J.R. (2005)** Cholesterol facilitates the native mechanism of Ca<sup>2+</sup>-triggered membrane fusion. *J. Cell Sci.*, 118, 4833-4848. **Highlighted article in this issue. Cited in Faculty of 1000. [145 citations]**
75. Churchward, M., Butt, R.H., Lang, J.C., Hsu, K.K., and **Coorssen, J.R. (2005)** Enhanced detergent extraction for analysis of membrane proteomes by two-dimensional gel electrophoresis. *Proteome Science*, 3, 5 (June 7). **Designated as Highly Accessed. Most highly accessed article 2005-2006; 9<sup>th</sup> most highly accessed article in this journal.**
76. Butt, R.H., and **Coorssen, J.R. (2005)** Postfractionation for Enhanced Proteomic Analyses: Routine Electrophoretic Methods Increase the Resolution of Standard 2D-PAGE. *J. Proteome Res.*, 4, 982-991 (Epub April 30). **Designated as Highly Accessed.**
77. Szule, J.A. and **Coorssen, J.R. (2004)** Comment on 'Transmembrane Segments of Syntaxin Line the Fusion Pore of Ca<sup>2+</sup>-Triggered Exocytosis.' *Science*, 306, 813.
78. Whalley, T., Timmers, K., **Coorssen, J.R.,** Kingsley, D.H., and Zimmerberg, J. (2004) Membrane fusion of secretory vesicles of the sea urchin egg in the absence of NSF. *J. Cell Sci.*, 117, 2345-2356.
79. Szule, J.A., Jarvis, S., Hibbert, J.E., Spafford, J.D., Braun, J.E.A., Zamponi, G., Wessel, G.M., and **Coorssen, J.R. (2003)** Specific presynaptic proteins function upstream of

- triggered membrane fusion. *J. Biol. Chem.*, 278, 24251-24254 (Epub 21/05/03). **Accelerated Publication.**
80. **Coorssen, J.R.**, Blank, P.S., Albertorio, F., Bezrukov, L., Kolosova, I., Chen, X., Backlund, P. and Zimmerberg, J. (2003) Regulated secretion: SNARE density, vesicle fusion, and calcium dependence. *J. Cell Sci.*, 116, 2087-2097. **Cited as "Must Read" in Faculty of 1000 (Factor 6.0); [www.f1000biology.com/article/12692190/evaluation](http://www.f1000biology.com/article/12692190/evaluation).**
  81. Szule, J.A. and **Coorssen, J.R.** (2003) Revisiting the role of SNAREs in exocytosis and membrane fusion. *Biochim. Biophys. Acta. – Mol. Cell Res.*, 1641, 121-135 (requested for a special issue on membrane fusion).
  82. **Coorssen, J.R.**, Blank, P.S., Albertorio, F., Bezrukov, L., Kolosova, I., Backlund, P. and Zimmerberg, J. (2002) Quantitative femto- to attomole immunodetection of regulated secretory vesicle proteins critical to exocytosis. *Anal. Biochem.*, 307, 54-62.
  83. Yergey, A.L., **Coorssen, J.R.**, Backlund, P.S., Blank, P.S., Humphrey, G.A., Zimmerberg, J., Campbell, J.M., and Vestal, M.L. (2002) De novo sequencing of peptides using MALDI-TOF/TOF. *J. Am. Soc. Mass Spectrom.* 13, 784-791. **Most cited JASMS paper for 2002 [225 citations].**
  84. Zimmerberg, J., Blank, P.S., Kolosova, I., Cho, M.-S., Tahara, M. and **Coorssen, J.R.** (2000) Stage-specific preparations to study the Ca<sup>2+</sup>-triggered fusion steps of exocytosis: rationale and perspectives. *Biochimie* 82, 303-314.
  85. Zimmerberg, J., **Coorssen, J.R.**, Vogel, S.S. and Blank, P.S. (1999) Sea urchin egg preparations as systems for the study of calcium-triggered exocytosis. *J. Physiol.*, 520.1, 15-21.
  86. **Coorssen, J.R.**, Blank, P.S., Tahara, M. and Zimmerberg, J. (1998) Biochemical and functional studies of cortical vesicle fusion: the SNARE complex and Ca<sup>2+</sup> sensitivity. *J. Cell Biol.* 143, 1845-1857. **[146 citations]**
  87. Scepek, S., **Coorssen, J.R.** and Lindau, M. (1998) Fusion pore expansion in horse eosinophils is modulated by Ca<sup>2+</sup> and protein kinase C via distinct mechanisms. *EMBO J.* 17, 4340-4345. **[123 citations]**
  88. Tahara, M., **Coorssen, J.R.**, Timmers, K., Blank, P.S., Whalley, T., Scheller, R. and Zimmerberg, J. (1998) Calcium disrupts the SNARE protein complex on sea urchin egg secretory vesicles without irreversibly blocking fusion. *J. Biol. Chem.* 273, 33667-33673.
  89. **Coorssen, J.R.**, Schmitt, H. and Almers, W. (1996) Ca<sup>2+</sup> triggers massive exocytosis in Chinese hamster ovary cells. *EMBO J.* 15, 3787-3791. **[138 citations]**
  90. **Coorssen, J.R.** (1996) Phospholipase activation and secretion: evidence that PLA<sub>2</sub>, PLC, and PLD are not essential to exocytosis. *Am. J. Physiol.* 270 (Cell Physiol. 39), C1153-C1163.
  91. Parsons\*, T.D., **Coorssen\*, J.R.**, Horstmann, H. and Almers, W. (1995) Docked granules, the exocytic burst, and the need for ATP hydrolysis in endocrine cells. *Neuron* 15, 1085-1096. \*Both authors contributed equally **[353 citations]**
  92. **Coorssen, J.R.** and Rand, R.P. (1995) Structural effects of neutral lipids on divalent cation-induced interactions of phosphatidylserine-containing bilayers. *Biophys. J.* 68, 1009-1018.
  93. **Coorssen, J.R.** and Haslam, R.J. (1993) GTP<sub>γ</sub>S and phorbol ester act synergistically to stimulate both Ca<sup>2+</sup>-independent secretion and phospholipase D activity. Inhibition by BAPTA and analogues. *FEBS Lett.* 316, 170-174.
  94. **Coorssen, J.R.**, Davidson, M.M.L. and Haslam, R.J. (1990) Factors affecting dense and α-granule secretion from electroporabilized human platelets: Ca<sup>2+</sup>-independent actions of phorbol ester and GTP<sub>γ</sub>S. *Cell Regulation* 1, 1027-1041.
  95. **Coorssen, J.R.** and Rand, R.P. (1989) Effects of cholesterol on the structural transitions induced by diacylglycerol in phosphatidyl-choline and -ethanolamine bilayer systems. *Biochemistry and Cell Biology* 68, 65-69.
  96. **Coorssen, J.R.** and Rand, R.P. (1988) Competitive forces between lipid membranes. *Studia Biophysica* 127, 53-60.

Papers in preparation

97. Almuslehi, M.S., Sen, M.K., Shortland, P.J., Mahns, D.A., and **Coorsen, J.R. (2020)** Proteomic analysis of the optic nerve in the cuprizone model of multiple sclerosis.
98. Malladi, C., Gauci, V.J., and **Coorsen, J.R. (2021)** A proteomic analysis of the myometrium in labour and preterm labour.
99. Schaffran, B., Malladi, C., Gauci, V.J., and **Coorsen, J.R. (2021)** CNS proteome changes with the development of hypertension.

iii. BOOKS, CHAPTERS

1. Mansour, V.J., and **Coorsen, J.R. (2018)** Quantitative gel electrophoresis. In: Proteomics in Domestic Animals: from Farm to Systems Biology (de Almeida, A. et al., Eds). Springer. ISBN 978-3-319-69681-2.
2. **Coorsen, J.R. (2017)** Proteomics. In: Reference Module in Life Sciences. Elsevier Inc. ISBN 9780128096338, [http://dx.doi.org/\[Pii\]](http://dx.doi.org/[Pii]).
3. Mansour (née Gauci), V.J., Noaman, N., and **Coorsen, J.R. (2016)** Gel staining techniques – dyeing to know it all. In: eLS (Encyclopedia of Life Sciences). John Wiley & Sons, Ltd: Chichester. DOI: 10.1002/9780470015902.a0002686.pub3
4. Abbineni, P., Wright, E.P., Rogasevskaia, T.P., Killingsworth, M., Malladi, C., and **Coorsen, J.R. (2014)** The Sea Urchin Egg and Cortical Vesicles as Model Systems to Dissect the Fast, Ca<sup>2+</sup>-Triggered Steps of Regulated Exocytosis. In: Neuromethods (Thorn, P., Ed). Humana Press / Springer.
5. **Coorsen, J.R. (2013)** Top-down Proteomics: 2D gels are an integral part of the process. In: Farm Animal Proteomics 2013, pp. 31-33 (de Almeida, A. et al., Eds). Wageningen Academic Publishers / Springer. ISBN 978-90-8686-776-9.
6. **Coorsen, J.R. (2013)** Proteomics. In: Brenner's Encyclopedia of Genetics, 2<sup>nd</sup> Edition, Part 1231 (Maloy, S.; Hughes, K., Editors-in-Chief.). Academic Press / Elsevier.
7. **Coorsen, J.R. (2009)** Synaptic Proteins and Regulated Exocytosis. In: Encyclopedia of Neuroscience, Part 19, pp. 3964-3968 (Binder, Marc D.; Hirokawa, Nobutaka; Windhorst, Uwe, Eds.). Springer Verlag GmbH, Heidelberg.
8. **Coorsen, J.R. (2009)** Proteomics and the Study of the Nervous System. In: Encyclopedia of Neuroscience, Part 16, pp. 3330-3333 (Binder, Marc D.; Hirokawa, Nobutaka; Windhorst, Uwe, Eds.). Springer Verlag GmbH, Heidelberg.
9. Parsons, T.D., **Coorsen, J.R.**, Horstmann, H., Lee, A.K., Tse, F.W. and Almers, W. (1995) The last seconds in the life of a secretory vesicle. In: Cold Spring Harbor Symposia on Quantitative Biology, Volume 60, Protein Kinesis: The Dynamics of Protein Trafficking and Stability (pp. 389-396) CSH Laboratory Press, New York.
10. Haslam, R.J. and **Coorsen, J.R. (1993)** Evidence that activation of phospholipase D mediates Ca<sup>2+</sup>- independent secretion from human platelets. In: Mechanisms of Platelet Activation and Control (Authi, K.S., Watson, S.P. and Kakkar, V.V. eds.) Adv. Exp. Med. Biol. (pp. 149-164) Plenum, New York.

iv. ABSTRACTS (1987 - present)

1. Coish, J.M., Crozier, R.W.E., Schieffelin, J.S., **Coorsen, J.R.**, Hunter, F.F., and Adam J. MacNeil, A.J. (2020) Zika virus infection of mast cells is not associated with a degranulation response. Immunology 2020 (American Association of Immunologist) Honolulu, USA (8/5/2020)
2. Coish, J.M., Crozier, R.W.E., Schieffelin, J.S., **Coorsen, J.R.**, Hunter, F.F., and Adam J. MacNeil, A.J. (2020) Pre-existing Dengue virus immunity augments mast cell infection by Zika virus (abstract R4535). Experimental Biology. San Diego, USA (4/4/2020)
3. Sen, M.K., Almuslehi, M.S., **Coorsen, J.R.**, Mahns, D.A., and Shortland, P.J. (2019) Behavioural and histological changes in cuprizone-fed mice. Macquarie Neurodegeneration Meeting. Sydney, Australia. (19/07/2019)

4. Kurgan, N., Noaman, N., Pergande, M.R, Cologna, S.M., **Coorssen, J.R.**, and Klentrou, N. **(2019)** An anti-inflammatory phenotype following high intensity interval exercise: a sequential top-down proteomic analysis of human serum. Exercise Metabolism: a Cell Symposium. Sitges, Spain.
5. Dabral, D., and **Coorssen J.R. (2019)** Cortical vesicle associated PLA2: Identification and role in native membrane fusion. Biophysical Society (Boston, February). Baltimore, MS, USA. (2-6/03/2019)
6. Almuslehi, M.S., Sen, M.K., Mahns, D.A., Shortland, P.J., and **Coorssen, J.R. (2018)** Blood Brain Barrier Disruption Facilitates CD8 T Cell infiltration into the CNS of Orchiectomized Cuprizone Treated Mice. Annual meeting of the Australian Neuroscience Society. Brisbane, Australia. (3-6/12/2018).
7. Sen, M.K., Almuslehi, M.S., Myers, S.J., Mahns, D.A., **Coorssen, J.R.**, and Shortland, P.J. **(2018)** Detection of early behavioural deficits in cuprizone fed mice. Annual meeting of the Australian Neuroscience Society. Brisbane, Australia. (3-6/12/2018).
8. Almuslehi, M.S., Sen, M.K., Mahns, D.A., Shortland, P.J., and **Coorssen, J.R. (2018)** Bilateral Orchiectomy Preserves Peripheral Immune Organs in Cuprizone Treated Mice. Annual Meeting of the Society for Neuroscience. San Diego, USA.
9. Kurgan, N., Noaman, N., **Coorssen, J.R.**, and Klentrou, N. **(2018)** Sequential top-down analysis of the human serum proteome in response to high intensity exercise and recovery. Annual Meeting of the Canadian Society for Exercise Physiology. Niagara Falls, Canada.
10. Dabral, D., and **Coorssen J.R. (2018)** Cortical vesicle associated PLA<sub>2</sub>: Identification and characterization. Health Beyond Research & Innovation Showcase. Sydney, Australia.
11. Almuslehi, M.S., Sen, M.K., Mahns, D.A., Shortland, P.J., and **Coorssen, J.R. (2018)** Bilateral Orchiectomy Preserves Peripheral Immune Organs in Cuprizone Treated Mice. Health Beyond Research & Innovation Showcase. Sydney, Australia.
12. Sen, M.K., Almuslehi, M.S., Mahns, D.A., **Coorssen, J.R.**, and Shortland, P.J. **(2018)** Detection of early behavioural deficits in Cuprizone treated mice. Health Beyond Research & Innovation Showcase. Sydney, Australia.
13. Backlund, P.S., Blank, P.S., **Coorssen, J.R.**, Cologna, S.M., Klein, C., and Yergey, A.L. **(2018)** Lipid Aggregations Studied using Electrospray Ionization and Ion Mobility. Annual meeting of the American Society for Mass Spectrometry. San Diego, USA.
14. Noaman, N., Abbineni, P.S., Withers, M., and **Coorssen, J.R. (2018)** High Sensitivity Top-down Proteomics: Coomassie for In-gel Proteoform Detection Rivals MS-based Peptide Detection. ASBMB Annual Meeting. San Diego, USA.
15. Sen, M.K., Shortland, P.J., Myers, S.J., **Coorssen, J.R.**, and Mahns, D.A. **(2017)** Treatment with pertussis toxin does not induce a multiple sclerosis-like phenotype in cuprizone-treated mice. 47<sup>th</sup> Meeting of the Society for Neuroscience. Washington, DC.
16. Sen, M.K., Shortland, P.J., Myers, S.J., **Coorssen, J.R.**, and Mahns, D.A. **(2017)** Treatment with pertussis toxin does not induce a multiple sclerosis-like phenotype in cuprizone-treated mice. UNSW Brain Science Symposium. Sydney, Australia.
17. Van Leeuwen, M.P., Thevarajah, J.J., Ward, R., **Coorssen, J.R.**, Castignolles, P., and Gaborieau, M. **(2017)** Characterisation of branching starch by capillary electrophoresis. 67th Australasian Grain Science Conference. Christchurch, New Zealand.
18. Noaman, N., Abbineni, P.S., Withers, M., and **Coorssen, J.R. (2017)** It's how you use it: Coomassie staining enables (sub)femtomole detection of intact gel-resolved proteoforms for Top-down Discovery Proteomics. 5th International Congress on Analytical Proteomics. Lisbon, Portugal.
19. Radzieta, M., Espedido, B., Malladi, C., **Coorssen, J.R.**, Killingsworth, M., van Hal, S.J., Phillips, J., and Jensen, S.O. **(2017)** Investigating the Mechanism of Action and Clinical Utility of the Novel Antimicrobial BDM-I. ASM/ESCMID Conference on Drug Development to Meet the Challenge of Antimicrobial Resistance. Boston, USA.
20. **Coorssen, J.R.**, D'Silva, A., and Hyett, J. **(2017)** Identification of Post Translational Modifications of First Trimester Maternal Serum Proteins that are Predictive for Spontaneous Preterm Birth. 5th International Congress on Analytical Proteomics. Lisbon, Portugal.
21. D'Silva, A., Hyett, J., and **Coorssen, J.R. (2017)** Identification of Post Translational

- Modifications of First Trimester Maternal Serum Proteins that are Predictive for Spontaneous Preterm Birth. 21<sup>st</sup> International Conference, Prenatal Diagnosis & Therapy. San Diego, USA.
22. Van Leeuwen, M.P., Toutounji, M.R., Mata, J., Gilbert, E., Pallas, L., Ward, R., **Coorsen, J.R.**, Castignolles, P., and Gaborieau, M. **(2017)** Exploring the supra(molecular) characterisation of starch in rice as a tool to predict digestibility. Australian Institute of Food Science & Technology Summer School. Sydney, NSW, Australia.
  23. Van Leeuwen, M.P., Toutounji, M.R., Mata, J., Gilbert, E., Pallas, L., Ward, R., **Coorsen, J.R.**, Castignolles, P., and Gaborieau, M. **(2016)** Relating starch structure of rice to its digestibility. GI Taskforce, RACI Analytical and Environmental Chemistry Research and Development meeting. Sydney, NSW, Australia.
  24. Radzieta, M., Espedido, B., Malladi, C., **Coorsen, J.R.**, Phillips, J., and Jensen, S.O. **(2016)** Investigating the Mechanism of Action of the Novel Antimicrobial BDM-I. European Congress for Clinical Microbiology and Infectious Diseases. Amsterdam, NLD.
  25. Van Leeuwen, M.P., Thevarajah, J.J., **Coorsen, J.R.**, Gaborieau, M., and Castignolles, P. **(2016)** Separation of amylose and amylopectin and characterisation of their branching using capillary electrophoresis. Royal Australian Chemical Institute, NSW ssNMR and Polymer Symposium. UNSW, NSW, Australia.
  26. D'Silva, A., Hyett, J., and **Coorsen J.R.** **(2016)** A top-down proteomic approach to the identification of early serum biomarkers predictive of spontaneous preterm birth. 21<sup>st</sup> Lorne Proteomics Symposium. Lorne, Australia.
  27. Van Leeuwen, M.P., Toutounji, M.R., Mata, J., Gilbert, E., Shrestha, A., Pallas, L., Ward, R., **Coorsen, J.R.**, Castignolles, P., and Gaborieau, M. **(2015)** Molecular characterisation of starch by NMR spectroscopy and capillary electrophoresis. Annual International Congress of Pacific Basic Chemical Societies. Honolulu, USA.
  28. Van Leeuwen, M.P., Thevarajah, J.J., **Coorsen, J.R.**, Gaborieau, M., and Castignolles, P. **(2015)** Characterisation of branching in starch using capillary electrophoresis. Royal Australian Chemical Institute, NSW Open Education meeting. Macquarie U., Australia.
  29. Radzieta, M., Espedido, B., Malladi, C., **Coorsen, J.R.**, Phillips, J., and Jensen, S.O. **(2015)** Investigating the Mechanism of Action of the Novel Antimicrobial BDM-I. ASM. Canberra, Australia.
  30. Van Leeuwen, M.P., Toutounji, M.R., **Coorsen, J.R.**, Castignolles, P., and Gaborieau, M. **(2015)** Capillary electrophoresis of carbohydrates: from the quantification of sugars in breakfast cereals to the characterisation of starch in rice. Royal Australian Chemical Institute, NSW Polymer Research Symposium. UNSW, Australia.
  31. Malladi, C.S., D'Silva, A., Sen, M.K., Partridge M.A., and **Coorsen, J.R.** **(2015)** High resolution quantitative top-down proteomics: Resolving protein species as a key to identifying biomarkers and analyzing molecular mechanisms. 4<sup>th</sup> International Congress on Analytical Proteomics. Lisbon, Portugal.
  32. Noaman, N., Abbineni, P.S., and **Coorsen, J.R.** **(2015)** Modified Coomassie staining supports different imaging formats for quantitative top-down Discovery Proteomics. Human Proteome Organization World Congress. Vancouver, Canada.
  33. Toutounji, M., van Leeuwen, M., Shrestha, A., Pallas, L., Ward, R., **Coorsen, J.R.**, Castignolles, P., Gaborieau, M. **(2015)** Starch structure and digestibility in rice and breakfast cereals. 65<sup>th</sup> Australian Cereal Chemistry Conference / Australian Grain Science Association annual meeting. Sydney, Australia.
  34. **Coorsen, J.R.** and Abbineni, P.S. **(2015)** What is the docked state of a release-ready vesicle? 18<sup>th</sup> International Symposium on Chromaffin Cell Biology. Cairns, Australia.
  35. Abbineni, P.S., Dabral, D., and **Coorsen, J.R.** **(2015)** Dissecting phosphoregulation of exocytosis: Assessing the roles of kinases in late steps of the exocytotic pathway. 18<sup>th</sup> International Symposium on Chromaffin Cell Biology. Cairns, Australia.
  36. Partridge, M.A., Gopinath, S., Myers, S., and **Coorsen, J.R.** **(2015)** Understanding Multiple Sclerosis by using a top-down Proteomic approach. 20<sup>th</sup> Lorne Proteomics Symposium. Lorne, Australia.

37. **Coorsen, J.R. (2014)** Lipid regulation of exocytosis. Australian Physiological Society. Brisbane, Australia.
38. Rogasevskai, T.P., and **Coorsen, J.R. (2014)** A role for PLD in the late steps of exocytosis. **18<sup>th</sup> IUPAB Congress**. Brisbane, Australia.
39. Gano, C., Scott, K., Salinovic, M., **Coorsen, J.R.**, Dong, Q., Bucci, J., de Souza, P., and Harman, D.G. **(2014)** Does dietary intervention change urine isoprostane levels in prostate cancer patients? – A pilot study. 2<sup>nd</sup> Australian Lipid Meeting. Wollongong, Australia.
40. Mahadeo, M., Czyz, O., Lam, S., **Coorsen, J.R.**, Zaremborg, V., and Prenner, E. **(2014)** Studying Lateral Organization in complex Eukaryotic membrane extracts using Brewster Angle Microscopy. 18<sup>th</sup> IUPAB Congress. Brisbane, Australia.
41. Ormerod, K., Aksamit, S., **Coorsen, J.R.**, Tattersall, G.J., and Mercier, A.J. **(2014)** Cholesterol and its metabolites in *Drosophila* development. Southern Ontario Neuroscience Association Annual Meeting. Western University, London, ON, Canada.
42. Abomughaid, M., Tay, E., **Coorsen, J.R.**, George, J., and Mark W. Douglas, M.W. **(2014)** Detailed lipidomic analysis of hepatitis C virus infected cells to identify new drug targets. 10<sup>th</sup> Annual Scientific Workshop of the Australian Centre for HIV and Hepatitis Virology Research (ACH<sup>2</sup>). Katoomba, Australia.
43. Krahn-Roldan, A., Esmores, L., Aksamit, S., Ormerod, K., **Coorsen, J.R.**, and Mercier, A.J. **(2014)** Cholesterol removal elicits contraction in *Drosophila* muscles. Southern Ontario Neuroscience Association Annual Meeting. Western University, London, ON, Canada.
44. Stroud, L.C., Padula, M., Slapeta, J., **Coorsen, J.R.**, and Stack, C.M. **(2014)** *Tritrichomonas foetus* the master manipulator. 50<sup>th</sup> Annual Meeting of the Australian Society of Parasitology. Canberra, Australia.
45. Abbineni, P., and **Coorsen, J.R. (2014)** A proteomic and ultra-structural examination of the late stages of regulated exocytosis. 19<sup>th</sup> Lorne Proteomics Symposium. Lorne, Australia.
46. **Coorsen, J.R. (2014)** Vesicular 'Omics: Defining the release-ready and Ca<sup>2+</sup>-triggering fusion steps of regulated exocytosis. 19<sup>th</sup> Lorne Proteomics Symposium. Lorne, Australia.
47. Cronin, L., Slapeta, J., Padula, M., **Coorsen, J.R.**, and Stack, C. **(2013)** *Tritrichomonas foetus*: Identifying critical proteins involved in host-parasite interaction. 24<sup>th</sup> International Conference of the World Association for the Advancement of Veterinary Parasitology. Perth, Australia.
48. Pickford, R., Abomughaid, M., Tay, E., **Coorsen, J.R.**, and Douglas, M.W. **(2013)** Changes in the Lipidome during Hepatitis C virus infection measured with mass spectrometry. American Society for Mass Spectrometry annual meeting. Minneapolis, USA.
49. Abomughaid, M., Eslam, M., Tay, E., **Coorsen, J.R.**, George, J., and Mark W. Douglas, M.W. **(2013)** Lipidomics profiling identifies PEMT as a key enzyme in metabolic reprogramming in chronic hepatitis C infection. 20<sup>th</sup> International Symposium on Hepatitis C Virus and Related Viruses (HCV meeting). Melbourne, Australia.
50. Abbineni, P., Wright, E.P., and **Coorsen, J.R. (2013)** The Omics of exocytosis: defining the release-ready and Ca<sup>2+</sup>-triggering fusion steps. 3<sup>rd</sup> International Congress on Analytical Proteomics. San Pedro, Brazil.
51. Abomughaid, M., Tay, E., **Coorsen, J.R.**, George, J., and Mark W. Douglas, M.W. **(2013)** Using lipidomics to identify new drug targets against the hepatitis C virus. 9<sup>th</sup> Annual Scientific Workshop of the Australian Centre for HIV and Hepatitis Virology Research (ACH<sup>2</sup>). Katoomba, Australia.
52. Cronin, L., **Coorsen, J.R.**, and Stack, C. **(2013)** Dissecting the roles of key molecules involved in the host-parasite interaction of *Tritrichomonas foetus*. Joint Academic Microbiology Seminars. Sydney, NSW.
53. LePine, O., Ormerod, K.G., **Coorsen, J.R.**, Mercier, A.J., and Tattersall, G.J. **(2013)** Behaviour of larval and adult *Drosophila* is significantly altered by different commercially available, standard media. Annual meeting of the Canadian Association of Neuroscience. Toronto, Canada.
54. Abomughaid, M., Tay, E., **Coorsen, J.R.**, George, J., and Mark W. Douglas, M.W. **(2013)** Detailed lipidomic analysis of hepatitis C virus infected cells to identify new drug targets. Marie Bashir Institute Annual Meeting. Sydney, Australia.

55. Milward, E.A., Johnstone, D.M., Graham, R.G., **Coorssen, J.R.**, Abbott, C.A., Mate, K., Wouters, M., and Myers, M. (for CUBEnet Health Futures Working Group) **(2012)** An overview of biomedical and 'omics' content in the curricula of leading Australian universities. Game On: Preparing our biology and biomedical graduates for the future; joint CUBEnet/ VIBEnet/Quantitative Skills in Science Education Forum. U. Sydney, NSW, Australia.
56. Malladi, C.S., Gauci, V., Padula, M.P., Wood, S., and **Coorssen, J.R.** **(2012)** Understanding human labour: myometrial proteomics. Biomarker Discovery Conference. Shoal Bay, NSW, Australia.
57. Wright, E.P., Harper, B.W., Padula, M.P., Aldrich-Wright, J.R., and **Coorssen, J.R.** **(2012)** Cancer proteomics: learning from cell lines and new anticancer agents. Biomarker Discovery Conference. Shoal Bay, NSW, Australia.
58. Gauci, V.J., and **Coorssen, J.R.** **(2012)** Sensitive in-gel protein detection using Coomassie. Proteomics and Beyond meeting. Macquarie U., Sydney.
59. Malladi, C.S., Gauci, V., Padula, M.P., Wood, S., and **Coorssen, J.R.** **(2012)** Understanding human labour: myometrial proteomics. Proteomics and Beyond meeting. Macquarie U., Sydney.
60. Wright, E.P., Harper, B.W., Padula, M.P., Aldrich-Wright, J.R., and **Coorssen, J.R.** **(2012)** Initial studies suggest that a novel anticancer agent may enhance protein degradation. Proteomics and Beyond meeting. Macquarie U., Sydney.
61. Malladi, C.S., Gauci, V., Padula, M.P., Wood, S., and **Coorssen, J.R.** **(2012)** Understanding human labour: myometrial proteomics. Human Proteome World Congress. Boston, USA.
62. Abomughaid, M., Tay, E., **Coorssen, J.R.**, George, J., and Mark W. Douglas, M.W. **(2012)** Detailed lipidomic analysis of hepatitis C virus infected cells to identify new drug targets. Annual Meeting of the Sydney Emerging Infections and Biosecurity Institute. Sydney, Australia.
63. Wright, E.P., Harper, B.W., Padula, M.P., Aldrich-Wright, J.R., and **Coorssen, J.R.** **(2012)** Initial studies suggest that a novel anticancer agent may enhance protein degradation. Human Proteome Organization World Congress. Boston, USA.
64. Gauci, V.J., and **Coorssen, J.R.** **(2012)** Sensitive in-gel protein detection using Coomassie. 9<sup>th</sup> Siena Meeting - From Genome to Proteome: Open Innovations. Siena, Italy.
65. Rogasevskaia, T., and **Coorssen, J.R.** **(2012)** The Critical Role of Lipids in the Ca<sup>2+</sup>-triggered Fusion Pathway. Biomarkers and Clinical Research. Las Vegas, USA.
66. Ormerod, K.G., Rogasevskaia, T., **Coorssen, J.R.** and Mercier, A.J. **(2012)** Cholesterol-independent effects of methyl-beta-cyclodextrin on chemical synapses. East Coast Nerve Net annual meeting. Amherst, USA.
67. Maxwell, K.A., Butt, R.H., **Coorssen, J.**, and Dyck, R.H. **(2012)** Characterization of pro- and anti-regenerative processes in the neonatal mouse brain. International Society of Developmental Neuroscience. Mumbai, India.
68. Gauci, V.J., and **Coorssen, J.R.** **(2011)** Optimised Coomassie brilliant blue staining for infrared fluorescence detection is a competitive alternative for sensitive gel-based proteomic analyses. P1423. Human Proteome World Congress. Geneva, Switzerland.
69. Gauci, V.J., and **Coorssen, J.R.** **(2011)** Optimisation of CBB staining for sensitive near IR protein detection. 2<sup>nd</sup> International Congress, Analytical Proteomics. Ourense, Spain.
70. Rogasevskaia, T., and **Coorssen, J.R.** **(2011)** Alternate Approaches to Analysing Lipid Contributions to Docking, Priming, and Fusion. 16<sup>th</sup> ISCCB Meeting / International Symposium on Chromaffin Cell Biology. Beijing, China.
71. Ormerod, K., Rogasevskaia, T., **Coorssen, J.R.**, and Mercier, A.J. **(2011)** Postsynaptic actions of a cholesterol chelator depend on temperature acclimatization. Annual meeting of the Canadian Association of Neuroscience. Quebec City, Canada.
72. Furber, K.L., and **Coorssen, J.R.** **(2010)** Thiol-reactivity: A small molecule approach to indentifying proteins involved in regulating the calcium sensing steps of native membrane fusion. Australian Physiological & Biophysics Societies joint meeting. (Adelaide, November).

73. Butt, R.H., Gauci, V.J., and **Coorssen, J.R. (2010)** High sensitivity in-gel protein detection using coomassie blue as a near-IR dye. Human Proteome World Congress; 2010 Sep 19-23; Sydney, Australia. p 398-399. HUPO 2010 (Sydney, September). Abstract # PO208.
74. Ormerod, K., **Coorssen, J.R.**, and Mercier, A.J. **(2010)** Postsynaptic actions of a cholesterol chelator depend on temperature-acclimatization. Southern Ontario Neuroscience Association Annual Meeting, Brock University (St.Catharines, ON, Canada, May).
75. Furber, K.L., and **Coorssen, J.R. (2010)** An Unbiased Approach to Identifying Proteins Critical to the Mechanism of Ca<sup>2+</sup>-Triggered Membrane Fusion. Australian Physiological and Neuroscience Societies joint annual meeting. (Sydney, January). Proceedings of Australian Physiol Soc. 40, 43-54.
76. Butt, R.H., and **Coorssen, J.R. (2009)** Refining gel-based proteomics: Towards optimized methods of analysis. 1<sup>st</sup> International Congress on Analytical Proteomics/5<sup>th</sup> Congress of the Portuguese Proteomics Network. (Lisbon, November).
77. Butt, R.H., Pfeifer, T.A., Lubieniecka, J.M., Liu, J., Grigliatti, T.A., Foster, L.J., Tetzlaff, W., and **Coorssen, J.R. (2009)** Molecular dissection of spinal cord injury using a refined proteomic approach. HUPO 2009 (Toronto, September). MCP 8 (Suppl 1)
78. Furber, K.L., and **Coorssen, J.R. (2009)** Thiol-Reactivity: A tool to dissect the mechanism of calcium-triggered membrane fusion. Annual Hunter Cellular Biology Meeting (Hunter Valley, March).
79. Furber, K.L., and **Coorssen, J.R. (2009)** Using Thiol-Reactivity to Identify Proteins Involved in the Ca<sup>2+</sup>-Triggering Steps of Native Membrane Fusion. Annual Meeting of the Biophysical Society (Boston, February). Biophys. J. 96 (Suppl 1), 361a.
80. Rogasevskaia, T., and **Coorssen, J.R. (2008)** Manipulations of intact sea urchin eggs *in vitro*: a new paradigm for molecular studies of regulated membrane fusion. Annual Meeting of the American Society for Cell Biology (San Francisco, December). Mol Biol Cell 19.
81. **Coorssen, J.R. (2008)** Integrating studies of proteins and lipids: dissecting the mechanism of Ca<sup>2+</sup>-Triggered membrane fusion. Annual Meeting of the Australian Physiological Society. (<http://www.aups.org.au/Proceedings/39/4P>).
82. Maxwell, K.A., Toole, C., Butt, R.H., **Coorssen, J.**, and Dyck, R.H. **(2008)** Pro- and anti-regenerative processes characterized in the neonatal mouse brain. 6<sup>th</sup> Hershey Conference on Developmental Brain Injury (Paris, June)
83. Furber, K.L., Churchward, M.A., Rogasevskaia T.P and **Coorssen, J.R. (2008)** Identifying critical components of native Ca<sup>2+</sup>-triggered membrane fusion: Integrating studies of proteins and lipids. International EU Meeting on Mechanism(s) of Exocytosis (Ljubljana, Slovenia, May).
84. **Coorssen, J.R. (2008)** Enhancing resolution and detection for gel-based proteomics. 8th Annual International Conference of the Canadian Proteomics Initiative; [http://cpicanada.org/CPI08\\_Program\\_Booklet.pdf](http://cpicanada.org/CPI08_Program_Booklet.pdf). (Vancouver, Canada, May).
85. Maxwell, K.A., Toole, C., Butt, R.H., **Coorssen, J.**, and Dyck, R.H. **(2008)** Pro- and anti-regenerative processes characterized in the neonatal mouse brain. 5<sup>th</sup> International Symposium on Neuroprotection and Neurorepair: Cerebral Ischemia and Stroke (Magdeburg, Germany, May).
86. Rogasevskaia, T., Churchward, M.A., and **Coorssen, J.R. (2007)** Exploring the roles of specific phospholipids in fast calcium-triggered membrane fusion. Annual Meeting of the American Society for Cell Biology (Washington DC, December). Mol Biol Cell 18
87. Valitski, M., Blank, P.S., Raveh, A., **Coorssen, J.R.**, Zimmerberg, J.J., and Rahamimoff, R. **(2007)** Non Stationary fluctuation analysis of calcium in secretory vesicles. Annual Meeting of the Israeli Society for Neurosciences.
88. Maxwell, K.A., Toole, C., Butt, R.H., **Coorssen, J.**, and Dyck, R.H. **(2007)** Characterization of pro- and anti-regenerative processes in the neonatal mouse brain. Society for Neuroscience Annual Meeting (San Diego, November).
89. Churchward, M.A., Rogasevskaia, T., Taylor, R.C., and **Coorssen, J.R. (2007)** Specific lipids provide critical negative curvature to enable Ca<sup>2+</sup>-triggered native membrane fusion. Society of General Physiology Annual Symposium: "Membrane biophysics of fusion, fission, and rafts in health and disease (Woods Hole, MA, USA, September).

90. Churchward, M.A., Furber, K.L., Dunn, T.W., Rogasevskaia, T., and **Coorssen, J.R. (2007)** Identifying an essential component of calcium-triggered vesicular release: Specific lipids contribute a critical negative curvature. IBRO 2007 (Melbourne, Australia, July).
91. Furber, K.L., Brandman, D.M., and **Coorssen, J.R. (2007)** Enhancing the Ca<sup>2+</sup>-Sensitivity of Triggered Membrane Fusion using Thiol-Reactive Reagents. The Secretory Vesicle Cycle and Novel Approaches to its Analysis: IBRO Satellite Meeting (Brisbane, Australia, July).
92. Maxwell, K.A., Butt, R.H., **Coorssen, J.**, and Dyck, R.H. **(2007)** Characterization of pro- and anti-regenerative processes in the neonatal mouse brain. Canadian Stroke Network, Annual General Meeting (Quebec City, June).
93. Lehmann, A., **Coorssen, J.**, and Mercier, A.J. **(2007)** Effects of a cholesterol chelator on synaptic transmission in crayfish depend on acclimatization temperature. Canadian Association of Neuroscience Annual Meeting (Toronto, May).
94. Churchward, M.A., Taylor, R.C., Furber, K.L., Rogasevskaia, T., and **Coorssen, J.R. (2007)** A critical negative curvature contributed by specific lipids is an essential component of Ca<sup>2+</sup>-triggered native membrane fusion. Keystone Symposium: Bioactive Lipids in the Lipidomics Era (Taos, February).
95. Butt, R.H., Churchward, M.A., and **Coorssen, J.R. (2006)** Assessing dogma: recent advances in 2D-PAGE. **12<sup>th</sup> Lorne Proteomics Symposium** (annual meeting of the Australasian Proteomics Society, Victoria, Australia, February).
96. Furber, K.L., Brandman, D.M., Yergey, A.L., and **Coorssen, J.R. (2006)** Thiol-reactivity as a route to identifying proteins involved in Ca<sup>2+</sup>-triggered membrane fusion. Annual Meeting of the American Society for Cell Biology (San Diego, December). Mol Biol Cell 17
97. Churchward, M.A., Rogasevskaia, T., Taylor, R.C., and **Coorssen, J.R. (2006)** Negative curvature contributed by specific lipids is an essential component of Ca<sup>2+</sup>-triggered native membrane fusion. Annual Meeting of the American Society for Cell Biology. Mol Biol Cell 17
98. Maxwell, K.A., Butt, R.H., **Coorssen, J.R.**, and Dyck, R.H. **(2006)** Inflammation and neural regeneration after neonatal brain injury. Annual Meeting of the Canadian Stroke Network (St. John's, May, 2006).
99. Khan, O., Krioutchkova, S., Ranganathan, P., Butt., R.H., and **Coorssen, J.R. (2005)** Proteomic analyses investigate possible long-term effects of exposure to microwave radiation. Canadian Proteome Society International Meeting: Enhanced Technologies for Proteomics (Calgary, Sept., 2005). **Awarded first prize in the first annual CPS student research competition.**
100. Furber, K.L., Dunn, T.W., Wiersma-Meems, Syed, N.I., and **Coorssen, J.R. (2005)** Cholesterol facilitates neurotransmitter release. Annual Meeting of the Society for Neuroscience E43; 380.12.
101. Butt, R.H., Lee, M.Y., Pirshahid, S.A., Backlund, P.S., Wood, S., and **Coorssen, J.R. (2005)** Proteomic analyses to define the molecular mechanisms underlying human preterm labour. Annual Meeting of the Canadian Proteomics Initiative (Toronto, May, 2005).
102. Weselake, R., Kazala, C., Marshall, K., Shi, Y., Furukawa-Stoffer, T., Urban, S., Thibault, J., Foroud, N., **Coorssen, J.**, Moloney, M., Laroche, A., Olson, D., and Ross, A. **(2005)** Gene expression in microspore-derived oil-forming cell suspension cultures of oilseed rape exposed to low temperature or osmotic stress. Annual Meeting of the American Society of Plant Biology (Seattle).
103. Szule, J.A., Churchward, M.A., Butt, R.H., and **Coorssen, J.R. (2005)** Cholesterol is an essential component of the minimal native fusion machine. Biophys. J. 88, 379A (Suppl. S).
104. Dyck, R.H., Maxwell, K.A., Maherali, N.A., and **Coorssen, J.R. (2005)** Comparison of the regenerative capacity of the cerebral cortex in neonatal mice following traumatic or ischemic injury. J. Cerebral Blood Flow & Metabolism 25, S232.
105. Butt, R.H., Churchward, M.A., Ernst, M.G., and **Coorssen, J.R. (2004)** 2D-PAGE Protocols for High Resolution Analyses of Membrane Proteomes. For the Proteome Society Meeting "New Technologies, Novel Approaches in Proteomics Research" (December; [www.proteome.org/3Events/f\\_events.htm](http://www.proteome.org/3Events/f_events.htm)).
106. McRory, J.E., Latour, I., Beedle, A., Hamid, J., Chen, L., Lamb, J., **Coorssen, J.R.**, and Zamponi, G.W. **(2004)** T-type calcium channel splice isoform variant association with

- Annexin III in neuronal tumors. For the XL National Meeting of the Argentine Society for Biochemistry & Molecular Biology (SAIB).
107. McRory, J.E., Latour, I., Beedle, A., Hamid, J., Chen, L., Lamb, J., **Coorsen, J.R.**, and Zamponi, G.W. (2004) T-type calcium channel splice variant association with Annexin III in neuronal tumors. Abstract for the Annual Meeting of the Society for Neuroscience.
  108. Maxwell, K.A., **Coorsen, J.R.**, and Dyck, R.H. (2004) Proteomic analysis of factors enhancing and inhibiting regeneration following cortical injury in neonatal mice. Abstract for the Annual Meeting of the Society for Neuroscience.
  109. Weselake, R., Kazala, C., Urban, S., Huff, P., Marshall, K., Furukawa-Stoffer, T., Moloney, M., Lamb, J., **Coorsen, J.**, Ross, A., and Olson, D. (2004) Proteomics of abiotic stress in triacylglycerol-producing cultures of oilseed rape. Submitted for the annual meeting of the American Oil Chemists' Society (Cincinnati, 05/04).
  110. Hibbert, J.E., Szule, J.A., Butt, R.H., and **Coorsen, J.R.** (2003) Models-to-mechanism: Testing hypotheses concerning Ca<sup>2+</sup>-triggered membrane fusion. Cell Biology of the Neuron meeting (New Orleans), Neuron (Ref. Cellbio 111).
  111. Hibbert, J.E., Albertorio, F., and **Coorsen, J.R.** (2003) Testing models of Ca<sup>2+</sup>-triggered membrane fusion. Mol. Biol. Cell 14, 356a (#1988).
  112. Szule, J.A., Jarvis, S.E., Hibbert, J.E., Spafford, J.D., Braun, J.E., Zamponi, G.W., Wessel, G.M., and **Coorsen, J.R.** (2003) Calcium-triggered membrane fusion proceeds independently of specific presynaptic proteins. Mol. Biol. Cell 14, 356a (#1993).
  113. **Coorsen, J.R.**, Butt, R.H., Hibbert, J.E., and Szule, J.A. (2003) Integrated functional-molecular analyses to explore models of regulated membrane fusion. HUPO/IUBMB meeting (Montreal). Mol. Cell. Proteomics 2, 695 (Abstract 22.4).
  114. Weselake, R.J., Kazala, E.C., Urban, S., Huff, P.W., Marshall, K., Moloney, M.M., J.C. Lamb, and **Coorsen, J.R.** (2003) Proteomic investigation of oil-forming membranes from cell suspension cultures of *Brassica Napus*. Annual meeting of the Canadian Section of the American Oil Chemists' Society (Edmonton, 09/03).
  115. Weselake, R.J., Kazala, E.C., Urban, S., Huff, P.W., Moloney, M.M., J.C. Lamb, and **Coorsen, J.R.** (2003) Effect of low temperature on microsomal protein expression and oil formation activity in cell suspension cultures of *Brassica Napus*. 1<sup>st</sup> Canadian Plant Genomics Workshop (Saskatoon).
  116. Raveh, A., Blank, P.S., **Coorsen, J.R.**, Epstein, J., Shani, L., Zimmerberg, J., and Rahamimoff, R. (2001) Calcium dynamics in secretory vesicles. Annual meeting of the Israeli Physiological Society.
  117. Verma, A., **Coorsen, J.R.**, Yin, S-R., Poustka, A., Blank, P.S., Backlund, P., and Zimmerberg, J. (2001) Identification of the calcium-binding proteins regulating the membrane fusion steps of exocytosis. ASCB Annual Meeting. Mol. Biol. Cell 12, 378.
  118. **Coorsen, J.R.**, Zimmerberg, J., and Blank, P.S. (2001) Kinetics of homotypic cortical vesicle fusion. Biophys. J. 80, 141a (Abstract 627.13).
  119. Zimmerberg, J., Frolov, V., Reese, T.S., Blank, P.S., Chernomordik, L., and **Coorsen, J.R.** (2000) Membrane fusion complexes in exocytosis and viral entry. Presented at the International Juan March Meeting (Spain): Comparison of the mechanisms of cellular and viral membrane fusion. November, 2000.
  120. Blank, P.S., **Coorsen, J.R.**, Albertorio, F. and Zimmerberg, J. (2000) Vesicle protein modification alters the rate and extent of calcium-triggered fusion. Biophys. J. 78, 316A (Abstract 1871).
  121. Blank, P.S., **Coorsen, J.R.**, Epstein, J., Harari, E., Meiri, H., Rahamimoff, R., Raveh, A. and Zimmerberg, J. (1999) Fluo-4 fluorescence dynamics in docked secretory vesicles of sea urchin eggs. Neurosci. Lett., Supplement 54, S9.
  122. **Coorsen, J.R.**, Blank, P.S., Kolosova, I., Backlund, P. and Zimmerberg, J. (1999) Are SNARE proteins necessary & sufficient for membrane fusion? Mol. Biol. Cell 10, 219a, #1269.
  123. Rahamimoff, R., Blank, P.S., **Coorsen, J.R.**, Epstein, J., Fendyur, A., Harari, E., Kachalsky, S.G., Kaiserman, I., Krausz, D., Melamed-Book, N., Yagodin, S., and Zimmerberg, J. (1999) Calcium microdynamics as studied by fluctuation analysis of

- confocal images: oscillations, sparks and noise. Presented at the Annual Meeting of the Israeli Society of Physiology, Oct., 1999.
124. Blank, P.S., **Coorsen, J.R.**, Malley, J., Vogel, S. and Zimmerberg, J. **(1999)** Combined biochemical and functional studies of exocytosis. German Society for Neuroscience conference proceedings.
  125. **Coorsen, J.R.**, Blank, P.S., Tahara, M. and Zimmerberg, J. **(1998)** The final steps of exocytosis. *Mol. Biol. Cell* 9, 201a, #1164.
  126. **Coorsen, J.R.**, Blank, P.S., Tahara, M. and Zimmerberg, J. **(1998)** Using vesicle-vesicle fusion to study the final steps of Ca<sup>2+</sup>-triggered exocytosis. Presented at the 1998 Gordon Conference on "The Cell Biology of the Neuron," Plymouth, NH, U.S.A., June 14-19, 1998.
  127. **Coorsen, J.R.**, Blank, P.S., Tahara, M. and Zimmerberg, J. **(1998)** Vesicle-Vesicle fusion: A model system for the final steps of Ca<sup>2+</sup>-triggered exocytosis. Presented at the 1998 International Conference Jacques Monod on "Mechanisms of Exocytosis," France, April.
  128. **Coorsen, J.R.**, Blank, P.S., Tahara, M. and Zimmerberg, J. **(1998)** SNARE complex dissociation does not initiate membrane fusion. *Biophysical J.* 74, A95 (Pos303).
  129. **Coorsen, J.R.**, Tahara, M., Blank, P.S. and Zimmerberg, J. **(1997)** Role of SNARE proteins in membrane fusion. *Mol. Biol. Cell* 8, 296a, #1715.
  130. Tahara, M., **Coorsen, J.R.**, Timmers, K., Whalley, T., Scheller, R., and Zimmerberg, J. **(1997)** Role of SNARE proteins in the fusion of sea urchin cortical granules. *J. Gen. Physiology* 110, 24a, #67.
  131. **Coorsen, J.R.**, Schmitt, H. and Almers, W. **(1996)** Ca-triggered exocytosis in an epithelial cell line. *Biophys. J.* 70, A85.
  132. Scepek, S. **Coorsen, J.R.** and Lindau, M. **(1996)** Involvement of protein phosphorylation/dephosphorylation in dilation of the exocytotic fusion pore in eosinophils. German Society for Cell Biology Meeting. *European J. Cell Biol.* (abstracts '96).
  133. Parsons, T.D., **Coorsen, J.R.**, Horstmann, H. and Almers, W. **(1995)** Multiple pools of docked vesicles: Life after ATP hydrolysis in pituitary melanotrophs. Thirty-ninth annual meeting of the Biophysical Society, San Francisco. *Biophys. J.* 68, A117.
  134. **Coorsen, J.R.** and Haslam, R.J. **(1992)** Synergistic activation of phospholipase D (PLD) in permeabilized platelets by GTP[S] and phorbol ester or by GTP[S] and Ca<sup>2+</sup>; close correlation with secretion. Eighth International Conference on Second Messengers and Phosphoproteins, Glasgow, Aug. 3-8, 1992.
  135. Davidson, M.M.L., **Coorsen, J.R.** and Haslam, R.J. **(1992)** Stimulation of platelet Ca<sup>2+</sup>-independent protein kinase activities by phosphatidic acid and analogues. 8<sup>th</sup> International Conference on Second Messengers and Phosphoproteins, Glasgow, Aug. 3-8, 1992.
  136. **Coorsen, J.R.**, Davidson, M.M.L. and Haslam, R.J. **(1992)** Correlations between protein phosphorylation, phospholipase D activity and Ca<sup>2+</sup>-independent secretion from permeabilized platelets. ASBMB/ Biophysical Society. *FASEB J.* 6, A225.
  137. **Coorsen, J.R.** and Haslam, R.J. **(1991)** Correlation between GTP[S]-induced exocytosis from permeabilized platelets and activation of phospholipase D (PLD). *J. Cell Biol.* 115, 257a (#1490).
  138. **Coorsen, J.R.**, Davidson, M.M.L. and Haslam, R.J. **(1990)** GTP<sub>γ</sub>S and protein kinase C synergistically promote the Ca<sup>2+</sup>-independent secretion of dense and α-granule constituents from permeabilized platelets. *J. Cell Biol.* 111, 76a.
  139. **Coorsen, J.R.** and Haslam, R.J. **(1990)** Comparison of dense and α-granule secretion from permeabilized platelets: Effects of Ca<sup>2+</sup>, guanine nucleotides, thrombin and phorbol ester. *FASEB J.* 4, A2310.
  140. **Coorsen, J.R.**, Fuller, N.L. and Rand, R.P. **(1989)** Ca<sup>2+</sup>-induced interactions between PS-containing bilayers acts to dehydrate neighbouring PC molecules. *Biophys. J.* 55, 111a.
  141. **Coorsen, J.R.** and Rand, R.P. **(1987)** Effects of cholesterol on the structural transitions induced by diacylglycerol in phosphatidyl-choline (PC) and -ethanolamine (PE) bilayer systems. *Biophys. J.* 51, 157a.

iv. COMMUNICATIONS - unpublished (not peer-reviewed)

1. Kurgan, N., Noaman, N., **Coorssen, J.R.**, and Klentrou, N. **(2018)** Sequential top-down analysis of the human serum proteome in response to high intensity exercise. Saltin International Graduate Course in Exercise & Clinical Physiology. Blue Mountains, ON, Canada (24-27/09/2018).
2. Sen, M.K., Almuslehi, M.S., Myers, S.J., Mahns, D.A., **Coorssen, J.R.**, and Shortland, P.J. **(2018)** Variability of oligodendrocyte and myelin loss in Cuprizone-fed mice. WSU Higher Degree by Research Showcase. Sydney, Australia (22-26/10/2018).
3. Sen, M.K., Almuslehi, M.S., Shortland, P.J., Myers, S.J., **Coorssen, J.R.**, and Mahns, D.A. **(2017)** Identification of novel proteoforms in pertussis toxin-injected cuprizone-ingesting mice. Inter-University mental health and neuroscience conference. Sydney, Australia.
4. Sen, M.K., Almuslehi, M.S., Shortland, P.J., Myers, S.J., **Coorssen, J.R.**, and Mahns, D.A. **(2017)** Identification of differently expressed proteins in pertussis toxin injected cuprizone treated mice. WSU Higher Degree Research Symposium. Sydney, Australia.
5. Gano, C., Bucci J., Failes, T., Arndt, G., Dong, Q., Mahns, D., **Coorssen, J.R.**, Saedisomelia, A., Scott, K., and de Souza, P. **(2016)** Plant derived phytochemicals synergise in dual combination to suppress prostate cancer cell line growth at achievable plasma concentrations. Ingham Institute and SSWLHD Research & Teaching Showcase (10/16)
6. Killingsworth, M.C., Abbineni, P.S., and **Coorssen, J.R.** **(2015)** "Bending nature" to improve the visualisation of cancer cells and their microenvironment. Ingham Institute and SSWLHD Research & Teaching Showcase (11/15).
7. Gano, C., Scott, K., Salinovic, M., **Coorssen, J.R.**, Wright, E., Dong, Q., Bucci, J., de Souza, P., and Harman, D.G. **(2015)** Does dietary intervention change urine isoprostane levels in prostate cancer patients? – A pilot study. Ingham Institute and SSWLHD Research & Teaching Showcase (11/15).
8. Myers, S., and **Coorssen, J.R.** **(2015)** Multiple Sclerosis: autoimmunity or neurodegeneration. UWS/SWSLHD Conjoint Research Forum (03/15).
9. Noaman, N., and **Coorssen, J.R.** **(2015)** Getting the biggest biomedical bang out of established biobanks. UWS Summer Research Fellowship Program (02/15).
10. Partridge, M., and **Coorssen, J.R.** **(2014)** A Top-Down Proteomic Approach to Understanding Multiple Sclerosis. Ingham Institute Research & Teaching Showcase (11/14).
11. Noaman, N., Abbineni, P.S., and **Coorssen, J.R.** **(2014)** Improving protein detection methods for top-down proteomics. Ingham Institute Research & Teaching Showcase (11/14).
12. Malladi, C.S., Padula, MP, Wood, S., and **Coorssen, J.R.** **(2014)** Proteomic approach to understanding human labour. Ingham Institute Research & Teaching Showcase (11/14).
13. Partridge, M., Myers, S.J., Gopinath, S., and **Coorssen, J.R.** **(2014)** A Top-Down Proteomic Approach to Understanding Multiple Sclerosis. UWS MMRG Annual Meeting (11/14).
14. Noaman, N., Abbineni, P.S., and **Coorssen, J.R.** **(2014)** Improving protein detection methods for top-down proteomics. 2<sup>nd</sup> Proteomics & Beyond Symposium (Macquarie U., 11/14).
15. Abbineni, P.S., Myers, S., and **Coorssen, J.R.** **(2013)** Identifying membrane protein complexes critical for regulated exocytosis: a bias-free proteomics approach. UWS MMRG Annual Meeting (09/13).
16. Wright, E.P., Harper, B.W., Padula, M.P., Higgins, V.J., Aldrich-Wright, J.R., and **Coorssen, J.R.** **(2013)** A systems biology approach to investigating the effects of a novel platinum anticancer compound. UWS MMRG Annual Meeting (09/13).
17. Malladi, C.S., Schaffran, B., Gauci, V., Padula, M.P., and **Coorssen, J.R.** **(2013)** A proteomic approach to understanding hypertension. UWS MMRG Annual Meeting (09/13).
18. Cronin, L., Slapeta, J., Padula, M., **Coorssen, J.R.**, and Stack, C. **(2013)** *Tritrichomonas foetus*: Identifying critical proteins involved in host-parasite interaction. UWS MMRG Annual Meeting (09/13).

19. Stimpson, S., **Coorssen, J.R.**, Myers, S. (2013) Effects of SPTLC1 mutations on mitochondria: proteomic investigations. UWS MMRG Annual Meeting (09/13).
20. M. Featon, **Coorssen, J.R.**, Myers, S. (2013) Linking autophagy and peripheral neurodegeneration. UWS MMRG Annual Meeting (09/13).
21. Abbineni, P. and **Coorssen, J.R.** (2013) Identifying protein / lipid interactions that mediate regulated exocytosis. UWS SoM/SSH Research Futures Forum (07/13).
22. Cronin, L., **Coorssen, J.R.**, Stack, C. (2013) Using proteomics to dissect proteins expressed at the host-parasite interface of *Tritrichomonas foetus*. Joint Academic Microbiology Seminar, Sydney.
23. Wright, E.P., and **Coorssen, J.R.** (2012) Proteomic studies suggest that a novel anticancer agent may enhance protein degradation in *S. cerevisiae*. Proteomics & Beyond Symposium (Macquarie U., 11/14).
24. Gauci, V.J., and **Coorssen, J.R.** (2012) Sensitive in-gel protein detection using Coomassie. Proteomics & Beyond Symposium (Macquarie U., 11/14).
25. Malladi, C.S., and **Coorssen, J.R.** (2013) Understanding human labour: Myometrial proteomics. Proteomics & Beyond Symposium (Macquarie U., 11/14).
26. Mukkaddam, T., Chang, D., **Coorssen, J.R.**, Sucher, N. (2012) Molecular mechanisms underlying the effect of caloric-restriction and rapamycin on the chronological lifespan of *Saccharomyces cerevisiae*. UWS HDR Research Forum.
27. Abbineni, P. and **Coorssen, J.R.** (2011) Isolation and identification of cholesterol-binding proteins in a secretory vesicle membrane. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
28. Papalia, S., and **Coorssen, J.R.** (2011) Prion protein-dependent alterations in brain proteomes. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
29. Wisinski-Bokiniec, P., **Coorssen, J.R.**, Myers, S. (2011) The role of autophagy in dominant intermediate Charcot-Marie-Tooth disease. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
30. Lim, S., **Coorssen, J.R.**, O'Connor, M. (2011) The role of RBBP9 in the maintenance of human pluripotent stem cells. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
31. Hyland, R., **Coorssen, J.R.**, Witting, P., Myers, S. (2011) Characterising the cell biological changes in axonal degeneration caused by mutations in housekeeping proteins. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
32. Harper, B., **Coorssen, J.R.**, Aldrich-Wright, J. (2011) The synthesis, characterisation and biological activity of novel platinum (II) intercalators as potential anticancer compounds. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
33. Stimpson, S., **Coorssen, J.R.**, Myers, S. (2011) Investigating SPTLC1 mutations on protein and lipid profiles in HSN-1 patient lymphoblasts. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
34. Wright, E.P., Myers, S., and **Coorssen, J.R.** (2011) Enhancing in-gel protein detection to promote coupled proteomic-genomic analyses. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
35. Gauci, V.J., and **Coorssen, J.R.** (2011) Optimised CBB staining for IR fluorescence is a competitive alternative for sensitive gel-based proteomic analyses. Research Future Postgraduate Forum. UWS College of Health & Science (06/11).
36. Wright, E.P., Myers, S., and **Coorssen, J.R.** (2010) Investigating the use of reactive dyes as post-electrophoretic stains in both one- and two-dimensional proteomic analyses. Research Future Postgraduate Forum. UWS College of Health & Science (06/10).
37. Gauci, V.J., Myers, S., and **Coorssen, J.R.** (2010) Quantitation capacity of infrared dyes for protein detection in polyacrylamide gels. Research Future Postgraduate Forum. UWS College of Health & Science (06/10).
38. Smith, K., **Coorssen, J.R.**, Myers, S. (2010) Mitochondria – masters of malady! Research Future Postgraduate Forum. UWS College of Health & Science (06/10).

39. Stimpson, S., **Coorssen, J.R.**, Myers, S. (2010) Investigating SPTLC1 mutations on protein and lipid profiles in neuronal cells. Research Future Postgraduate Forum. UWS College of Health & Science (06/10).
40. **Coorssen, J.R.** (2009) Dissecting Molecular Mechanisms – Molecular Physiology in the UWS School of Medicine. Annual colloquium of the Nanoscale Organisation and Research Group. UWS 06/09.
41. **Coorssen, J.R.** (2009) Dissecting Molecular Mechanisms – Basic, Clinical & Translational Research. College of Health & Science Staff Research Futures Form, UWS (03/09)
42. Butt, R.H., Lee, J.Y., and **Coorssen, J.R.** (2008) Toward eating your cake and having it too: High sensitivity made affordable with infrared fluorescence detection of Coomassie Brilliant Blue. Presented by Dr. Butt at his awards talk for the Biochemistry and Molecular Biology Celebration of Excellence Symposium. U. Calgary.
43. Butt, R.H., Lee, J.Y., and **Coorssen, J.R.** (2008) High sensitivity detection of gel-based proteomes that won't break the bank: Infrared fluorescence detection of Coomassie Brilliant Blue. Presented to the Department of Biochemistry and Molecular Biology. U. Calgary.
44. Butt, R.H., Rogasevskajia, T. and **Coorssen, J.R.** (2008) A practical introduction to discovery proteomics: the theories, technologies, methods and more. Presented to the Masters of Biotechnology Program, Faculty of Medicine, U. Calgary.
45. Butt, R.H., Pfeifer, T.A., Delaney, A., Grigliatti, T.A., Tetzlaff, W. and **Coorssen, J.R.** (2008) Towards Understanding Injury: A discovery proteomics approach. Presented at the NeuroConnections / Hotchkiss Brain Institute annual retreat (Banff), U. Calgary.
46. Butt, R.H., Pfeifer, T.A., Delaney, A., Grigliatti, T.A., Tetzlaff, W. and **Coorssen, J.R.** (2007) A first look at spinal injury with optimized methods for coupled genomic and proteomic analyses. Presented at the Canadian Spring Conference on Brain and Behaviour. (Fernie, British Columbia).
47. Butt, R.H., Churchward M.A. and **Coorssen J.R.** (2007) Two dimensional gel electrophoresis dissected: theory, practice and recent advances driving discovery proteomics. Presented to the Masters of Biotechnology Program, Faculty of Medicine, U. Calgary.
48. Butt, R.H., and **Coorssen, J.R.** (2007) Are rigorous analyses of both proteins and RNA possible from one tissue sample? Presented at the NeuroConnections / Hotchkiss Brain Institute annual retreat (Banff), U. Calgary.
49. Churchward, M.A., Taylor, R.C., Rogasevskaia, T., and **Coorssen, J.R.** (2007) Specific lipids provide negative curvature to enable native Ca<sup>2+</sup>-triggered vesicle fusion. Presented at the NeuroConnections / Hotchkiss Brain Institute annual retreat (Banff), U. Calgary.
50. Furber, K.L., and **Coorssen, J.R.** (2007) Using thiol-reactive reagents to identify proteins involved in the Ca<sup>2+</sup>-triggered steps of membrane fusion. Presented at the NeuroConnections / Hotchkiss Brain Institute annual retreat (Banff), U. Calgary.
51. Taylor, R., Teskey, C., and **Coorssen, J.R.** (2007) Pre-seizure proteomics: identifying a continuum of molecular alterations. Presented at the NeuroConnections / Hotchkiss Brain Institute annual retreat (Banff), U. Calgary.
52. **Coorssen, J.R.** (2006) Proteomics and a new approach to understanding epilepsy. Teaching paper for a continuing medical education seminar as part of "Scientific and Technical Advances in Epilepsy," given in conjunction with the 41<sup>st</sup> Meeting of the Canadian Congress of Neurological Sciences, Montreal, Quebec. June, 2006.
53. Taylor, R., Teskey, C., Wiebe, S., and **Coorssen, J.R.** (2006) A proteomic approach to understanding epilepsy. Presented at the Hotchkiss Brain Institute / Molecules of the Mind retreat on Epilepsy (Banff), U. Calgary. January 2006.
54. Butt, R.H., Lee, M.Y., Luft, W.A., Ahmadi Pirshahid, S., Yergey, A.L., Wood, S., and **Coorssen, J.R.** (2004) Proteomic analyses to define the molecular mechanisms underlying human preterm labour. Presented at the opening of the Institute for Maternal and Child Health, Faculty of Medicine, U. Calgary. Nov. 2004.
55. Butt, R.H. and **Coorssen, J.R.** (2004) Routine electrophoretic methods for increasing the effective resolution of traditional 2D-PAGE. Presented at the BMB Annual Meeting (Banff), U. Calgary. Oct. 2004.

56. Churchward, M.A., Butt, R.H., Hsu, K.K., Lang, J.C., and **Coorssen, J.R. (2004)** Optimized extraction of membrane proteins for one- and two-dimensional electrophoresis. Presented at the BMB Annual Meeting (Banff), U. Calgary. Oct. 2004.
57. **Coorssen, J.R. (2004)** Biomolecular analyses and fundamental synaptic mechanisms: The Coorssen Lab. Presented at the pre-launch announcement of the Calgary Brain Institute, U. Calgary. March 2004.
58. Butt, R.H, Hibbert, J.E., Szule, J.A, and **Coorssen J.R. (2003)** A coupled functional-molecular approach to understanding calcium-triggered membrane fusion. Presented at the CMNRG Annual Retreat (Banff), U. Calgary. Oct. 2003.
59. Szule, J.A, and **Coorssen J.R. (2003)** Calcium-triggered membrane fusion proceeds independently of specific presynaptic proteins. Presented at the CMNRG Annual Retreat (Banff), U. Calgary. Oct. 2003.
60. Butt, R.H, Hibbert, J.E., Szule, J.A, and **Coorssen J.R. (2003)** A proteomic approach to the analysis of calcium-triggered membrane fusion. Presented at the BMB Annual Meeting (Banff), U. Calgary. Oct. 2003.
61. **Coorssen, J.R.**, Blank, P.S., Albertorio, F., Bezrukov, L., Backlund, P. and Zimmerberg, J. **(2001)** Identifying proteins essential to calcium regulated membrane fusion. Presented at the annual symposium of the Cell Biology and Metabolism Branch, NICHD, NIH.
62. Blank, P.S., **Coorssen, J.R.**, Albertorio, F., and Zimmerberg, J. **(2001)** Calcium triggered exocytosis: A kinetic synthesis. Presented at the annual symposium of the Cell Biology and Metabolism Branch, NICHD, NIH.
63. Verma, A., **Coorssen, J.R.** , Backlund, P., and Zimmerberg, J. **(2001)** Progress in identifying  $Ca^{2+}$  binding proteins regulating the membrane fusion steps of exocytosis. Presented at the annual symposium of the Cell Biology and Metabolism Branch, NICHD, NIH.
64. **Coorssen, J.R.**, Blank, P.S., Albertorio, F., Bezrukov, L., Kolosova, I., Fong, X., Backlund, P. and Zimmerberg, J. **(2001)** The  $Ca^{2+}$ -triggered membrane fusion steps of regulated exocytosis occur independently of SNARE proteins. Presented at the annual symposium of the Laboratory of Cellular & Molecular Biophysics, NICHD, NIH.
65. **Coorssen, J.R.**, Blank, P.S., Albertorio, F., Humphrey, G., Backlund, P. and Zimmerberg, J. **(2001)** Selective sulfhydryl modification and labeling of proteins essential to  $Ca^{2+}$ -triggered exocytosis. Presented at the annual symposium of the Laboratory of Cellular & Molecular Biophysics, NICHD, NIH.
66. Albertorio, F., Blank, P.S., Zimmerberg, J., and **Coorssen, J.R. (2001)** Testing the Calmodulin / V-ATPase hypothesis of membrane fusion. Presented at the annual symposium of the Laboratory of Cellular & Molecular Biophysics, NICHD, NIH.
67. Verma, A., **Coorssen, J.R.**, Backlund, P., and Zimmerberg, J. **(2001)** Identifying calcium binding proteins regulating exocytosis. Presented at the annual symposium of the Laboratory of Cellular & Molecular Biophysics, NICHD, NIH.
68. Miroslava, S., **Coorssen, J.R.**, Albertorio, F., Backlund, P., Zimmerberg, J. **(2001)** Identifying proteolipids in a  $Ca^{2+}$ -regulated exocytotic preparation. Presented at the annual symposium of the Laboratory of Cellular & Molecular Biophysics, NICHD, NIH.

#### v. MEDIA COVERAGE

1. Cutting-edge research equipment purchased thanks to Brock community effort. Brock News 19/07/2019; <https://brocku.ca/brock-news/2019/07/cutting-edge-research-equipment-purchased-thanks-to-brock-community-effort/>
2. International student improves lab processes to help scientists conduct protein research. Brock News 14/06/2018; <https://brocku.ca/brock-news/2018/06/international-student-improves-lab-processes-to-help-scientists-conduct-protein-research/>
3. Brock-led study examines role of protein levels in preterm birth. Brock News 10/04/2018; <https://brocku.ca/brock-news/2018/04/brock-led-study-shows-differences-in-protein-levels-that-could-predict-preterm-birth/>
4. Faculty of Graduate Studies celebrates 10<sup>th</sup> anniversary. Brock News 21/03/2017; <https://brocku.ca/brock-news/2017/03/faculty-of-graduate-studies-celebrates-10th-anniversary/>

5. Brock University takes a 21st century approach to graduate studies (by M.T. Bitti). National Post. January 9, 2017.
6. Television interview on Graduate Studies at Brock University. Taking Niagara by Storm, with John Storm - Niagara Region news and local affairs show. COGECO TV, Oct 26 (6:30pm), Oct 28 (6:00pm), and Oct 30 (12:30pm) 2016.
7. Coorssen to head Brock faculty of graduate studies. ASBMB Today (the member magazine of the American Society for Biochemistry & Molecular Biology) 15 (9), 2016.
8. A homecoming for Brock's new Dean of Graduate Studies. The Brock Press 4/10/2016 (Vol 52; [https://issuu.com/thebrockpress/docs/october\\_4\\_\\_2016/1?e=17030302/39344165](https://issuu.com/thebrockpress/docs/october_4__2016/1?e=17030302/39344165)).
9. New Brock dean pushing for well-rounded grads. St. Catharines Standard, Niagara Falls Review, Welland Tribune, and Fort Erie Times. Hardcopy and E-editions. Aug 22, 2016 (<http://www.stcatharinesstandard.ca/2016/08/21/new-brock-dean-pushing-for-well-rounded-grads>).
10. Renowned protein analyst speaking at Brock. Brock News, Aug 4, 2016.
11. Brock grad returning from Australia to become new Dean of Graduate Studies. Brock News, April 25, 2016 (<https://brocku.ca/brock-news/2016/04/brock-grad-returning-from-australia-to-become-new-dean-of-graduate-studies/>)
12. Cell study challenges laws of biology and opens doors to new treatments (regarding our paper in JBC: 'The role of phospholipase D in regulated exocytosis.' Western News, Nov 11, 2015 ([http://www.westernsydney.edu.au/newscentre/news\\_centre/more\\_news\\_stories/cell\\_study\\_challenges\\_laws\\_of\\_biology\\_and\\_opens\\_doors\\_to\\_new\\_treatments](http://www.westernsydney.edu.au/newscentre/news_centre/more_news_stories/cell_study_challenges_laws_of_biology_and_opens_doors_to_new_treatments)))
13. Meeting Report: "Proteomics from Discovery to Function" 6<sup>th</sup> Annual Meeting of the Proteomics Society, Indian and International Conference – A Milestone for the Indian Proteomics Community. OMICS 19, 329-331 (2015).
14. Vesicular 'omics' (by Susan Williamson; Lorne Proteomics/Functional Proteomics feature). Australian Life Scientist 11, 14-16 (January/February 2014). Highlighting my combined exocytosis/proteomics/lipidomics research. (<http://lifescientist.com.au/content/molecular-biology/article/vesicular-39-omics-39--425851700>).
15. Funding a potential game-changer in MS research. Highlighting my community initiated and funded research program on Multiple Sclerosis. WSU Foundation Impact Report 2014.
16. UWS Honour Roll. UWS Foundation Impact Report 2013.
17. Wheels of Rotary move us closer to medical breakthrough (regarding Multiple Sclerosis research funded by the Rotary Club of Narellan), and Investing in the future of health – Mechanisms of calcium dependent membrane functions (regarding exocytosis research funded by Bellberry Ltd.). UWS Foundation Impact Report 2012.
18. Alumni of Distinction Award Recipients; Top 35 in the last 35 years. Surgite Magazine 4, 12-13 (2012). Brock University website (<http://www.brocku.ca/alumni-association/awards/alumni-of-distinction-awards/award-recipients>). February, 2012.  
<http://www.youtube.com/watch?v=l58sldpqaQg>
19. Brock working with University of Western Sydney. Brock University News. Sept. 22, 2011.
20. The cutting edge at Campbelltown (by M. Taverniti). Campbelltown-Macarthur newspaper. June 23, 2010.
21. Uni in the 'machine age' (by M. Taverniti). Camden newspaper. June 23, 2010.
22. Launch of the new Brain Circuits and Epilepsy Program. U of C News. Oct. 18, 2007.
23. U of C research team discovers proteins associated with preterm labour (by Laurie Wang). Faculty of Medicine News Digest. Sept. 25, 2007.
24. Named in the national funding announcement made by the Alberta Heritage Foundation for Medical Research. The Globe and Mail newspaper. March 18, 2006.
25. Go ahead, eat it up. U of C study discovers cholesterol's redeeming qualities (by Rene Bodack). Gauntlet student newspaper. Oct. 27, 2005.
26. High school students sample a taste of research (by Meghan Sired). OnCampus newspaper article highlighting one of my AHFMR HYRS summer students. Oct. 21, 2005 (Vol. 3, No. 4).

27. U of C has good news for bad cholesterol (by Eva Ferguson). Article in the A section of the Calgary Herald, Your Health – trends / discoveries spot, Oct. 21, 2005 (p. A17).
28. Research unclogs cholesterol myth (by Michelle Mark). Article in the Calgary Sun, News section, Oct. 21, 2005 (p. 10).
29. CITY TV (CHUM), Oct. 20, 2005. Cholesterol may not be as bad as we think (by Leah Sarich); 01:50 interview segment and coverage of the lab.
30. CTV (CFTO Toronto) News, Oct. 20, 2005. Cholesterol may not be as bad as we once thought; 02:40 news segment on our cholesterol findings.
31. CTV (CFCN) News, Oct. 20, 2005. Cholesterol may not be as bad as we once thought; 00:45 news segment on our cholesterol findings.
32. Global News, Oct. 20, 2005. Are statin drugs knocking your cholesterol *too* low? 01:20 news segment on our lab and findings concerning cholesterol.
33. CBC Radio (AM). Oct. 20, 2005. U of C researchers hoping study influences cholesterol treatments. Three 1 min radio segments concerning our cholesterol findings.
34. QR77 (CHQR-AM). U of C research suggests even the lowest doses of drugs used to treat cholesterol may be too much. 00:55 radio segment.
35. Give Cholesterol a break! U of C press release, Oct. 20, 2005. Highlighting the publication of *Cholesterol facilitates the native mechanism of Ca<sup>2+</sup>-triggered membrane fusion* in the Journal of Cell Science.
36. Summer jobs. Newspaper article in Life section of Red Deer Advocate (Aug., 2004) concerning my involvement with the Alberta Heritage Foundation for Medical Research Summer Student training program.
37. Global National News, Aug. 16, 2002. Two minute segment (by Francis Silvaggio) on my involvement with the AHFMR Summer Student training program.
38. Young innovator awards: Young U of C profs get \$20,000 boost. Calgary Herald, City Section, Jan. 12, 2002.
39. Young innovator awards help promising young researchers. Double page write up on my work and that of three others. Jan 14, 2002, University of Calgary Gazette (p. 10-11).
40. CTV News (National), Dec. 28, 2001. 2:35 minute segment on Proteomics (by A. Favro), including interviews with Dr. Power and me, and images of the facility.
41. U of C boasts 'cutting edge' lab (by Wendy-Anne Thompson). Banner title of major article in the Calgary Herald, City Section, Oct. 6, 2001.
42. Canadian proteomics researchers examine synaptic function (by Heather Kent). October 2001 issue of the *Chronicle of Neurology and Psychiatry*.
43. Global National News, Oct. 5, 2001. Two minute segment (by Kevin Newman and Leanne Niblock) on my use of sea urchins as model systems and the new proteomics facility. Plus two additional segments in other time slots.
44. CKAL-TV News (Calgary), Oct. 5, 2001. 2:20 minute segment (by Tom Njegovan and Bryan Labby) on my new proteomics facility and enhanced government funding at the U of C. Plus one additional segment in another time slot.
45. CFCN-TV News (Calgary), Oct. 5, 2001. Two minute segment (by Higgins/Janz and Kevin Rich) on my new proteomics facility and its proposed basic 'neurological' research. Plus one additional segment in another time slot.
46. CFRE-TV News (Regina), Oct. 5, 2001. Two minute segment (by Kevin Newman and Leanne Niblock) on my use of sea urchins as model systems & the new proteomics facility.
47. New lab is Canada's first integrated proteomics facility. Full page Research & Innovation article in the Oct. 15, 2001, University of Calgary Gazette (p. 16).
48. More medical marvels: New facility provides insight to CNS afflictions (by Andrea Brundon) in the Oct. 11, 2001, University of Calgary Gauntlet (p. 3).
49. Interview on SAIT radio, discussing the new proteomics facility and the planned applications of this newly acquired technology. Oct. 8, 2001.

vi. COMMUNITY ACTIVITIES

Christmas fundraiser. Brain Injury Community Re-entry Program. St. Catharines (12/2019).

Food for Thought. Brain Injury Association of Niagara annual fundraiser. St. Catharines (6/2019).

Assisted the Mentorship Program, Niagara Folk Arts Multicultural Centre, St. Catharines (9/2019).

Invited speaker, St. Catharines Central Rotary Club (04/2017).

Counsellors Day - represented Brock University Graduate Studies

- Part of a panel presentation to regional high school and college guidance counsellors; my role was to emphasize the importance of early consideration of graduate school as part of a student's career trajectory (8/12/2016).

CSIRO Scientist in Schools Program. Partner Scientist with Wollondilly Anglican College, NSW, 2013-2016; with Camden High School, NSW, 2011-2013.

- "Scientist in Residence" at Wollondilly Anglican College (2013-2016).
  - 2 hour seminar/discussion concerning Proteomics and potential experiments with students interested in pursuing a research project in my group (2013 - 2016).
  - 3 hour demo/practical lab session at UWS School of Medicine on sample preparation, gel electrophoresis & quantitative image analysis (2013 - 2016).
  - 2 hour seminar/discussion concerning gel electrophoresis data; what are 'resolution,' 'sensitivity' & 'coverage' and why are they important to analyses (2013 - 2016).

Guest speaker, the Rotary Club of Macarthur Sunrise. 'Multiple Sclerosis: What do we really know? How to proceed? Campbelltown, NSW. March 2016.

Various Community Engagement / Meet-and-Greet for Supporters of WSU. Throughout 2008-2016 including:

- Requested panel member, Rotary Club of Narellan Medical Student Scholarship (2009-2016); also reviewed scholarships provided by Campbelltown City Council (2014).
- Rotary Club of Narellan Annual Ball (2009-2015 & 2018); grant recipient in 2012-15.
- Hosting and lab tours, Rotary Club of Narellan annual WSU visit/dinner, 2009-2015
  - Brief update talk and lab tour + informal Q & A with members.
  - Introductory research talk 2012: Identifying Molecular and Cellular Mechanisms of MS using a Functional Proteomic Approach. Part of an acknowledgement to the Rotary Club of Narellan for funding to the MMRG to initiate a research program targeting Multiple Sclerosis.

Guest lecturer / community outreach at area primary school; Glenbrook Primary, 05/2015, a presentation on 'Molecules-to-Cells.'

Successfully nominated Miltenyi Biotec and Ecobiotics Ltd for WSU Partnership Awards (11/2010); Rotary Club of Narellan (11/2013).

Guest speaker, the Camden Science Cafe. 'Systems Biology: What it is, and where it is taking biomedical research and healthcare.' Camden, NSW. March 2013.

Award Presenter, Year 11 Achievement Awards, including WSU Most Outstanding Year 11 Student. Elderslie High School graduation ceremonies. Camden, NSW. December, 2013.

Brief research overview for official opening of the WSU/CHS Mass Spectrometry Facility (07/2010).

Hosting and lab tours, Rotary Club of Camden, 2009.

Brief overview of my training and scientific research, Dean of Medicine's Annual Reception for Donors and Friends of the University, 2009.

Introduction to research and why I joined WSU + lab tours, Rotary Club of Narellan, 2008.

Adjudicator for NSW Brain Bee, 2008 and 2011; short talk: Intro to Exocytosis & Endocytosis

Guest speaker, PEARLS (Preeclampsia Research Laboratories) fundraising dinner, Sydney (08/2008).

Guest lecturer / community outreach at area high school; Chevalier College, 06/09, 2 lectures ('Molecules-to-Cells' and 'Imaging').

Assistant coach, Foothills T-ball League for ages 3-7 (2002).